

# Expression analysis - water potential .alpha=0.05 .r=r

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# 1 Task

Final document in folder /reports:

```
> fileName(outputFile)
```

```
[1] "40_Expression-water-stress_0.05-r"
```

## 2 Information from pISA

Data directory

```
> .inroot
```

```
[1] "../..input"
```

Results directory

```
> .oroot
```

```
[1] "../..output/40_Expression-water-stress-0.05-r"
```

```
project:      _p_VinskaTrta
```

```
Investigation: _I_EnViRoS
```

```
Study:        _S_01_Integ
```

```
Assay:        _A_01_Desc-R
```

### 3 Data

Priporočljivo je najprej prebrati phenodata in feature-data, potem pa podatke. To omogoča izbor spremenljivk in vzorcev takoj za tem, ko podatke preberemo.

```
> cat(knit_child(file.path("../doc", "10a_Read-Data.Rnw"), quiet=TRUE))
```

List of data objects:

```
> my.objects
      name                description      class length ncol
1      t18      Transcripts for 2018 data.frame  15242   48
2      t19      Transcripts for 2019 data.frame  15242   32
3    t1819 Transcripts for 2018 and 2019 data.frame  15242   80
4    pdata18      Phenodata for 2018 data.frame    48   21
5    pdata19      Phenodata for 2019 data.frame    32   21
6  pdata1819 Phenodata for 2018 and 2019 data.frame    80   21
```

Factor for days within the year

```
> pdata1819$day <- factor(as.character(pdata1819$day))
```

#### 3.1 Water potential

On Dec 12 we decided to include water stress as a prediction factor.

In addition to expression data, we need water stress data.

```
> (wsfn <- getMeta(.adesc, "Water potential data"))
[1] "/input/Stem water potential 2018 2019.txt"
> wpdata <- read.table(file.path(.aroot,wsfn), sep="\t",
+   header=TRUE,
+   row.names=1)
> str(wpdata)
'data.frame':      78 obs. of  5 variables:
 $ variey      : chr  "Cabernet Volos" "Cabernet Volos" "Cabernet Volos" "Cab
 $ date        : chr  "4.06.2018" "15.06.2018" "21.06.2018" "29.06.2018" ...
 $ treatment   : chr  "WW" "WW" "WW" "WW" ...
 $ SWP.mean    : num  -0.34 -0.384 -0.34 -0.2 -0.4 -0.41 -0.455 -0.33 -0.455
 $ SWP.cumulated: num  -0.34 -0.724 -1.064 -1.264 -1.664 ...
```

Tidy factors and dates. Be careful, variety in the header is misspelled:

```
> names(wpdata)
[1] "variey"      "date"        "treatment"   "SWP.mean"
[5] "SWP.cumulated"
> head(wpdata)
      variey      date treatment SWP.mean SWP.cumulated
C18_3d_WW Cabernet Volos 4.06.2018      WW    -0.340      -0.340
C18_14d_WW Cabernet Volos 15.06.2018     WW    -0.384      -0.724
C18_20d_WW Cabernet Volos 21.06.2018     WW    -0.340      -1.064
C18_28d_WW Cabernet Volos 29.06.2018     WW    -0.200      -1.264
C18_33d_WW Cabernet Volos  4.07.2018     WW    -0.400      -1.664
C18_39d_WW Cabernet Volos 10.07.2018     WW    -0.410      -2.074
```

```

> X <- wpdata
> X$treat <- factor(X$treatment, levels=c("WW", "WS"))
> X$variety <- factor(X$variety)
> # Date of sampling might be useful as well
> require(lubridate)
> X$date <- as_date(X$date, format="%d.%m.%Y")
> X$year <- year(X$date)
> table(data.frame(X$date, "Count"=1))

```

```

          Count
X.date    1
2018-06-04 4
2018-06-15 4
2018-06-21 4
2018-06-29 4
2018-07-04 4
2018-07-10 4
2018-07-16 4
2018-07-23 4
2018-07-31 4
2018-08-07 4
2019-06-18 4
2019-06-26 4
2019-07-08 4
2019-07-12 4
2019-07-17 4
2019-07-22 2
2019-07-30 4
2019-08-06 4
2019-08-12 4
2019-08-19 4

```

```

> str(X)

```

```

'data.frame':      78 obs. of  8 variables:
 $ variety      : chr  "Cabernet Volos" "Cabernet Volos" "Cabernet Volos" "Cab
 $ date         : Date, format: "2018-06-04" ...
 $ treatment    : chr  "WW" "WW" "WW" "WW" ...
 $ SWP.mean     : num  -0.34 -0.384 -0.34 -0.2 -0.4 -0.41 -0.455 -0.33 -0.455
 $ SWP.cumulated: num  -0.34 -0.724 -1.064 -1.264 -1.664 ...
 $ treat       : Factor w/ 2 levels "WW","WS": 1 1 1 1 1 1 1 1 1 1 ...
 $ variety     : Factor w/ 2 levels "Cabernet Volos",...: 1 1 1 1 1 1 1 1 1 1
 $ year        : num  2018 2018 2018 2018 2018 ...

```

Order by date

```

> X <- X[order(X$date), ]

```

```

> wpdata <- X

```

Tidy dates ...

```

> table(wpdata$date, wpdata$variety)

```

	Cabernet Volos	Fleurtai
2018-06-04	2	2
2018-06-15	2	2
2018-06-21	2	2
2018-06-29	2	2
2018-07-04	2	2
2018-07-10	2	2
2018-07-16	2	2
2018-07-23	2	2
2018-07-31	2	2
2018-08-07	2	2
2019-06-18	2	2
2019-06-26	2	2
2019-07-08	2	2
2019-07-12	2	2
2019-07-17	2	2
2019-07-22	0	2
2019-07-30	2	2
2019-08-06	2	2
2019-08-12	2	2
2019-08-19	2	2

```

> table(pdata1819$date)
< table of extent 0 >
> pdata1819$date <- as_date(pdata1819$Date, format="%d.%m.%Y")
> table(pdata1819$date)

```

```

2018-06-11 2018-06-12 2018-07-05 2018-08-07 2019-06-26 2019-07-18
      8           8           16           16           16           16

```

```

> pdata1819$Year <- factor(paste0("20",pdata1819$year))

```

Water potential and transcriptomics are not measured at the same dates. We will interpolate the water potential to matching dates.

```

> wpas <- NULL
> par(mfrow=c(2,2),mar=c(3,4,2,1))
> for( yr in unique(wpdata$year)){
+ for (vari in levels(wpdata$variety)) {
+   with(wpdata, {
+     filter <- (wpdata$year==yr) & (variety== vari)
+     plot(date[filter], SWP.mean[filter], type="n",ylim=range(SWP.mean))
+     for(tr in levels(treat)){
+       select <- filter & (treat %in% tr)
+       lines(date[select], SWP.mean[select], type="b", pch=16, col=treat[select])
+       title(paste(yr,vari))
+       unidata <- unique(pdata1819$date[pdata1819$Year==yr])
+       abline(v=unidata, col=3)
+       wpa <- (approx(date[select], SWP.mean[select], unidata))
+       # print(data.frame(wpa))
+       # if(is.null(wpas)) wpas <- data.frame(wpa) else
+       wpas <-- rbind(wpas, data.frame(wpa, yr,
+       variety=substr(vari,1,1),

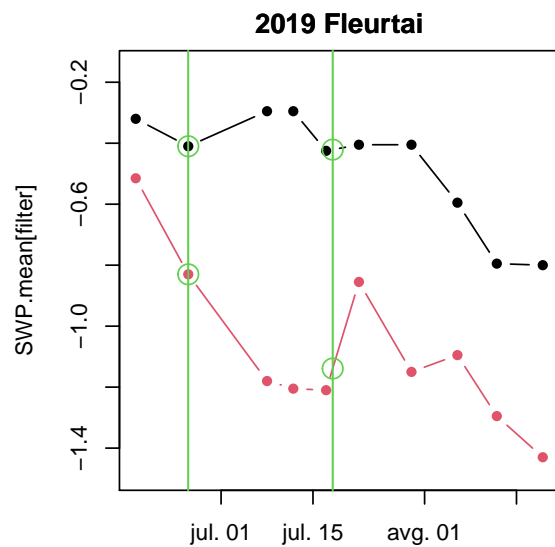
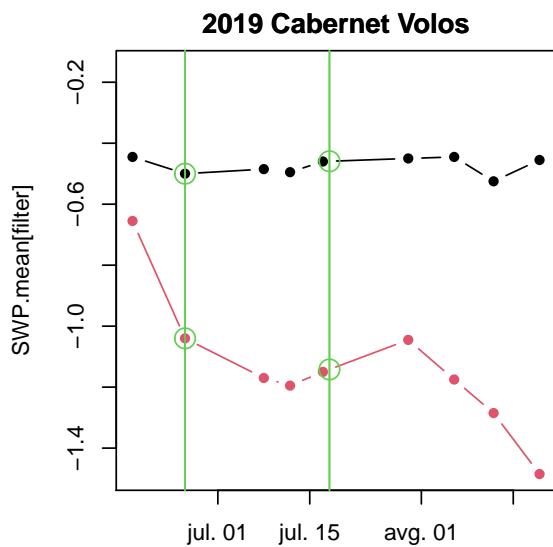
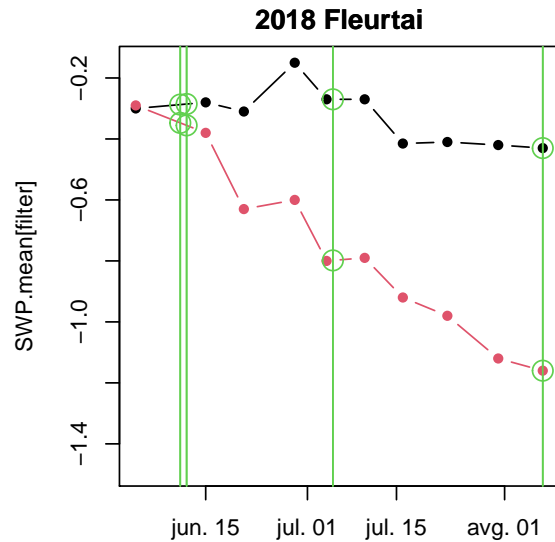
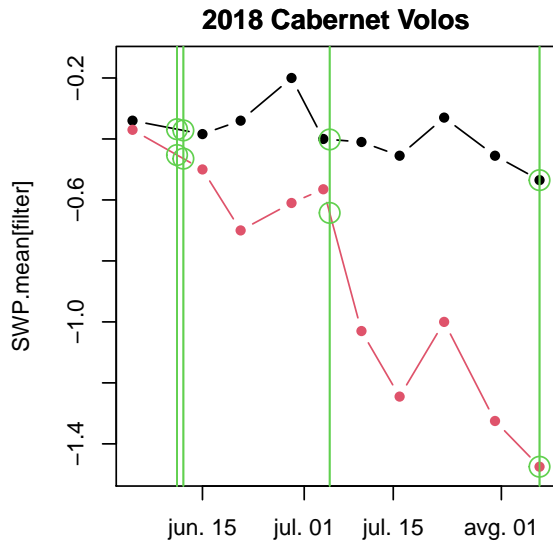
```



```

+   treat=tr))
+   points(wpa, col=3, cex=2)
+   }
+   })
+ }
+ }

```



```

> colnames(wpas) [1:2] <- c("date", "swp")
> #rownames(wpas) <- wpas[,1]
> #,by.x=c("date", "variety", "treat"),by.y=c("date", "variety", "treat"))

```

@

Order water potential data by date

```
> diff(wpas$date)
```

Time differences in days

```
[1] 23 33 -57 1 23 33 -57 1 23 33 -57 1 23 33 -57 380
[17] 22 -22 22 -22 22 -22 22
```

```
> wpas <- wpas[order(wpas$date),]
> diff(wpas$date)
```

```
Time differences in days
[1] 0 0 0 1 0 0 0 23 0 0 0 33 0 0 0 323
[17] 0 0 0 22 0 0 0
```

Convert treatment and variety into factors

```
> (wpas$treat <- factor(wpas$treat, levels=c("WW", "WS")))
[1] WW WS WW WS WW WS WW WS WW WS WW WS WW WS WW WS WW WS WW WS WW WS
[23] WW WS
Levels: WW WS

> (wpas$variety <- factor(wpas$variety))
[1] C C F F C C F F C C F F C C F F C C F F C C F F
Levels: C F
```

```
> with(wpas, ftable(yr, variety, treat, date))
              date 2018-06-11 2018-06-12 2018-07-05 2018-08-07 2019-06-26
yr  variety treat
2018 C      WW      1          1          1          1          0
      WS      1          1          1          1          0
      F      WW      1          1          1          1          0
      WS      1          1          1          1          0
2019 C      WW      0          0          0          0          1
      WS      0          0          0          0          1
      F      WW      0          0          0          0          1
      WS      0          0          0          0          1
```

Check date consistency

```
> all(pdata1819$date %in% wpas$date)
[1] TRUE
```

Add interpolated WP values to measured date to phenodata.

```
> mapdate <- match(pdata1819$date, wpas$date)
> pdata1819w <- pdata1819
> pdata1819w$swp <- wpas[mapdate, c("swp")]
> dim(pdata1819w)
[1] 80 24
> head(pdata1819w)
```

	ID	Variety	Date	variety	year	day
C18_11d_WS1	C18_11d_WS1	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WS2	C18_11d_WS2	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WS3	C18_11d_WS3	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WS4	C18_11d_WS4	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WW1	C18_11d_WW1	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WW2	C18_11d_WW2	Cabernet Volos	12.06.2018	C	18	11
	treat	rep	project.name	species	plant.name	tissue
C18_11d_WS1	WS	1	EnViros	Vitis vinifera	grapevine	leaf
C18_11d_WS2	WS	2	EnViros	Vitis vinifera	grapevine	leaf
C18_11d_WS3	WS	3	EnViros	Vitis vinifera	grapevine	leaf
C18_11d_WS4	WS	4	EnViros	Vitis vinifera	grapevine	leaf
C18_11d_WW1	WW	1	EnViros	Vitis vinifera	grapevine	leaf
C18_11d_WW2	WW	2	EnViros	Vitis vinifera	grapevine	leaf

```

health.status plant.number growth.location
C18_11d_WS1    water stress      R1          Udine
C18_11d_WS2    water stress      R2          Udine
C18_11d_WS3    water stress      R3          Udine
C18_11d_WS4    water stress      R4          Udine
C18_11d_WW1    well watered      R1          Udine
C18_11d_WW2    well watered      R2          Udine

```

```

growth.conditions
C18_11d_WS1    outside
C18_11d_WS2    outside
C18_11d_WS3    outside
C18_11d_WS4    outside
C18_11d_WW1    outside
C18_11d_WW2    outside

```

```

C18_11d_WS1    several leaves ground together and stored at -80oC; an aliquot was
C18_11d_WS2    several leaves ground together and stored at -80oC; an aliquot was
C18_11d_WS3    several leaves ground together and stored at -80oC; an aliquot was
C18_11d_WS4    several leaves ground together and stored at -80oC; an aliquot was
C18_11d_WW1    several leaves ground together and stored at -80oC; an aliquot was
C18_11d_WW2    several leaves ground together and stored at -80oC; an aliquot was

```

```

Metabolites.Order      Metabolites.File.Name
C18_11d_WS1            2 1019Cabernet Volos_6_12_WS1.D
C18_11d_WS2            4 1019Cabernet Volos_6_12_WS2.D
C18_11d_WS3           102 1219Cabernet Volos_6_12_WS3.D
C18_11d_WS4           100 1219Cabernet Volos_6_12_WS4.D
C18_11d_WW1            6 1019Cabernet Volos_6_12_WW1.D
C18_11d_WW2            8 1019Cabernet Volos_6_12_WW2.D

```

```

Metabolites.File Transcripts.ID      date Year
C18_11d_WS1 Cabernet Volos6_12WS      C1_S1 2018-06-12 2018
C18_11d_WS2 Cabernet Volos6_12WS      C1_S2 2018-06-12 2018
C18_11d_WS3 Cabernet Volos6_12WS      C1_S3 2018-06-12 2018
C18_11d_WS4 Cabernet Volos6_12WS      C1_S4 2018-06-12 2018
C18_11d_WW1 Cabernet Volos6_12WW      C1_W1 2018-06-12 2018
C18_11d_WW2 Cabernet Volos6_12WW      C1_W2 2018-06-12 2018

```

```

swp
C18_11d_WS1 -0.372
C18_11d_WS2 -0.372
C18_11d_WS3 -0.372
C18_11d_WS4 -0.372
C18_11d_WW1 -0.372
C18_11d_WW2 -0.372

```

```
> all(colnames(t1819)==rownames(pdata1819w))
```

```
[1] TRUE
```

One plot:

```
> t1819[1,]
```

	C18_11d_WS1	C18_11d_WS2	C18_11d_WS3	C18_11d_WS4
Vitvi15g01736	4.664602	4.109897	4.44822	4.013955
	C18_11d_WW1	C18_11d_WW2	C18_11d_WW3	C18_11d_WW4
Vitvi15g01736	4.717536	4.154237	4.475261	4.466465
	C18_34d_WS1	C18_34d_WS2	C18_34d_WS3	C18_34d_WS4
Vitvi15g01736	4.400333	4.641164	4.419123	4.534438
	C18_34d_WW1	C18_34d_WW2	C18_34d_WW3	C18_34d_WW4
Vitvi15g01736	4.924801	5.08569	5.339735	5.263629
	C18_67d_WS1	C18_67d_WS2	C18_67d_WS3	C18_67d_WS4
Vitvi15g01736	2.058717	2.289503	2.923483	2.575657
	C18_67d_WW1	C18_67d_WW2	C18_67d_WW3	C18_67d_WW4
Vitvi15g01736	3.891121	3.040087	4.216897	3.763086
	F18_10d_WS1	F18_10d_WS2	F18_10d_WS3	F18_10d_WS4
Vitvi15g01736	4.859141	4.866943	4.98059	4.945395
	F18_10d_WW1	F18_10d_WW2	F18_10d_WW3	F18_10d_WW4
Vitvi15g01736	5.705006	4.619238	4.808548	5.276124
	F18_34d_WS1	F18_34d_WS2	F18_34d_WS3	F18_34d_WS4
Vitvi15g01736	4.064718	3.850458	5.141832	4.626909
	F18_34d_WW1	F18_34d_WW2	F18_34d_WW3	F18_34d_WW4
Vitvi15g01736	4.096173	4.802221	4.329152	4.441903
	F18_67d_WS1	F18_67d_WS2	F18_67d_WS3	F18_67d_WS4
Vitvi15g01736	1.583128	1.740858	2.978406	4.166406
	F18_67d_WW1	F18_67d_WW2	F18_67d_WW3	F18_67d_WW4
Vitvi15g01736	3.558043	3.569471	4.411209	4.359744
	C19_22d_WS1	C19_22d_WS2	C19_22d_WS3	C19_22d_WS4
Vitvi15g01736	3.18	3.5	3.19	3.61
	C19_22d_WW1	C19_22d_WW2	C19_22d_WW3	C19_22d_WW4
Vitvi15g01736	5.42	5.35	4.68	4.92
	C19_44d_WS1	C19_44d_WS2	C19_44d_WS3	C19_44d_WS4
Vitvi15g01736	5.13	5.19	5.71	5.29
	C19_44d_WW1	C19_44d_WW2	C19_44d_WW3	C19_44d_WW4
Vitvi15g01736	6.19	5.62	5.62	6.19
	F19_22d_WS1	F19_22d_WS2	F19_22d_WS3	F19_22d_WS4
Vitvi15g01736	3.03	3.93	4.24	2.85
	F19_22d_WW1	F19_22d_WW2	F19_22d_WW3	F19_22d_WW4
Vitvi15g01736	5	4.97	4.47	5.51
	F19_44d_WS1	F19_44d_WS2	F19_44d_WS3	F19_44d_WS4
Vitvi15g01736	5.84	5.83	4.56	5.23
	F19_44d_WW1	F19_44d_WW2	F19_44d_WW3	F19_44d_WW4
Vitvi15g01736	5.44	6.13	5.96	6.16

```
> (varname <- rownames(t1819)[1])
```

```
[1] "Vitvi15g01736"
```

```
> .testing <- !TRUE
```

```
> pd <- pdata1819w
```

```
> par(mfrow=c(2,2),mar=c(3,4,2,1))
```

```
> (varname <- rownames(t1819)[1])
```

```
[1] "Vitvi15g01736"
```

```
> (yr <- unique(pd$year)[1])
```

```
[1] 18
```

```
Levels: 18 19
```

```

> (vari <- levels(pd$variety)[1])
[1] "C"
> for( yr in unique(pd$year)){
+ for (vari in levels(pd$variety)) {
+   filter <- (pd$year %in% yr) & (pd$variety %in% vari)
+   print(filter)
+   print(pd$date[filter])
+ ( y <- unlist(t1819[varname,]))
+ with(pd, {
+   plot(pd$swp[filter], y[filter],
+        type="p",
+        bg=pd$treat[filter],
+        pch=21,
+        cex=1,
+        col="white",
+        xlim=range(pd$swp)
+   )
+   for(i in 1:length(levels(treat))){
+   tr <- levels(pd$treat)[i]
+   select <- filter & (pd$treat %in% tr)
+   fit <- lm(y[select]~pd$swp[select])
+   print(fit)
+   abline(fit, col=i)
+ #   lines(date[select], swp[select], type="b", pch=16, col=treat[select])
+   title(paste(varname, yr, vari))
+   }
+ })
+ }

```

[1] TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE  
[12] TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE  
[23] TRUE TRUE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE  
[34] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE  
[45] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE  
[56] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE  
[67] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE  
[78] FALSE FALSE FALSE

[1] "2018-06-12" "2018-06-12" "2018-06-12" "2018-06-12" "2018-06-12"  
[6] "2018-06-12" "2018-06-12" "2018-06-12" "2018-07-05" "2018-07-05"  
[11] "2018-07-05" "2018-07-05" "2018-07-05" "2018-07-05" "2018-07-05"  
[16] "2018-07-05" "2018-08-07" "2018-08-07" "2018-08-07" "2018-08-07"  
[21] "2018-08-07" "2018-08-07" "2018-08-07" "2018-08-07"

```

Call:
lm(formula = y[select] ~ pd$swp[select])

```

```

Coefficients:
(Intercept) pd$swp[select]
       7.219          6.359

```

```

Call:

```

```
lm(formula = y[select] ~ pd$swp[select])
```

Coefficients:

```
(Intercept)  pd$swp[select]
      9.226           12.539
```

```
[1] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[12] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[23] FALSE FALSE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE
[34] TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE
[45] TRUE TRUE TRUE TRUE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[56] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[67] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[78] FALSE FALSE FALSE
[1] "2018-06-11" "2018-06-11" "2018-06-11" "2018-06-11" "2018-06-11"
[6] "2018-06-11" "2018-06-11" "2018-06-11" "2018-07-05" "2018-07-05"
[11] "2018-07-05" "2018-07-05" "2018-07-05" "2018-07-05" "2018-07-05"
[16] "2018-07-05" "2018-08-07" "2018-08-07" "2018-08-07" "2018-08-07"
[21] "2018-08-07" "2018-08-07" "2018-08-07" "2018-08-07"
```

Call:

```
lm(formula = y[select] ~ pd$swp[select])
```

Coefficients:

```
(Intercept)  pd$swp[select]
      7.011           5.778
```

Call:

```
lm(formula = y[select] ~ pd$swp[select])
```

Coefficients:

```
(Intercept)  pd$swp[select]
      9.935           13.685
```

```
[1] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[12] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[23] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[34] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[45] FALSE FALSE FALSE FALSE TRUE TRUE TRUE TRUE TRUE TRUE TRUE
[56] TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE FALSE FALSE
[67] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[78] FALSE FALSE FALSE
[1] "2019-06-26" "2019-06-26" "2019-06-26" "2019-06-26" "2019-06-26"
[6] "2019-06-26" "2019-06-26" "2019-06-26" "2019-07-18" "2019-07-18"
[11] "2019-07-18" "2019-07-18" "2019-07-18" "2019-07-18" "2019-07-18"
[16] "2019-07-18"
```

Call:

```
lm(formula = y[select] ~ pd$swp[select])
```

Coefficients:

```
(Intercept)  pd$swp[select]
```

15.06

19.93

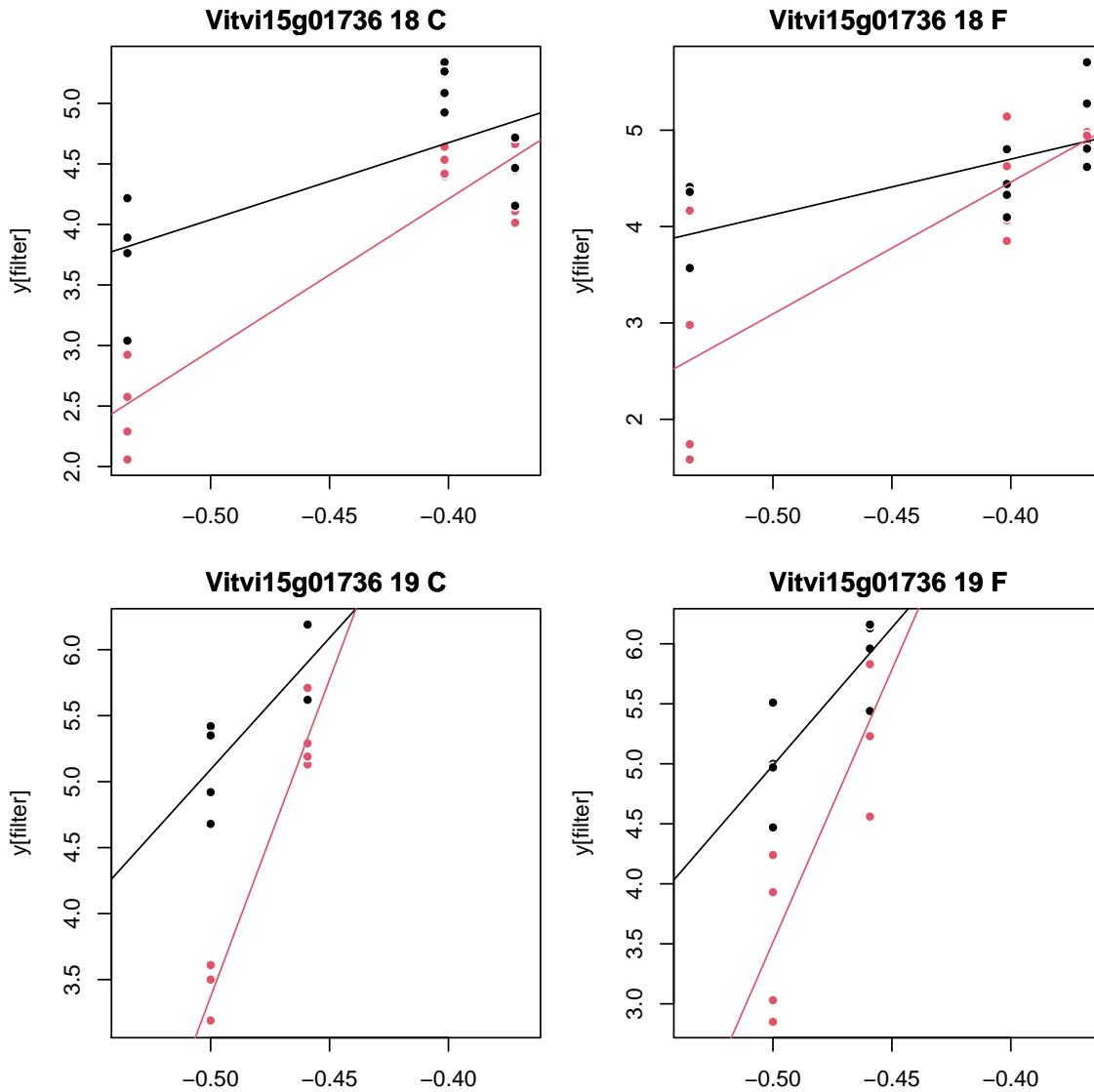
Call:

```
lm(formula = y[select] ~ pd$swp[select])
```

Coefficients:

```
(Intercept) pd$swp[select]
      27.41          48.08
```

```
[1] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[12] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[23] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[34] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[45] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[56] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE TRUE TRUE
[67] TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE
[78] TRUE TRUE TRUE
[1] "2019-06-26" "2019-06-26" "2019-06-26" "2019-06-26" "2019-06-26"
[6] "2019-06-26" "2019-06-26" "2019-06-26" "2019-07-18" "2019-07-18"
[11] "2019-07-18" "2019-07-18" "2019-07-18" "2019-07-18" "2019-07-18"
[16] "2019-07-18"
```



```
Call:
lm(formula = y[select] ~ pd$swp[select])
```

```
Coefficients:
 (Intercept)  pd$swp[select]
      16.45          22.93
```

```
Call:
lm(formula = y[select] ~ pd$swp[select])
```

```
Coefficients:
 (Intercept)  pd$swp[select]
      26.23          45.44
```

```
> .testing <- FALSE
```

## 4 Expression and water potential analysis



```

> varname <- rownames(t1819)[10]
> pd <- pdata1819
> (y <- unlist(t1819[varname,]))

C18_11d_WS1 C18_11d_WS2 C18_11d_WS3 C18_11d_WS4 C18_11d_WW1
  4.088427   3.961216   4.127170   3.916955   4.472212
C18_11d_WW2 C18_11d_WW3 C18_11d_WW4 C18_34d_WS1 C18_34d_WS2
  4.309540   4.212060   4.154032   4.086692   3.914995
C18_34d_WS3 C18_34d_WS4 C18_34d_WW1 C18_34d_WW2 C18_34d_WW3
  4.065776   4.081348   4.313965   4.041339   4.318997
C18_34d_WW4 C18_67d_WS1 C18_67d_WS2 C18_67d_WS3 C18_67d_WS4
  4.238240   4.441597   4.248104   4.242540   3.969094
C18_67d_WW1 C18_67d_WW2 C18_67d_WW3 C18_67d_WW4 F18_10d_WS1
  4.160356   4.676748   4.467201   4.359372   4.102511
F18_10d_WS2 F18_10d_WS3 F18_10d_WS4 F18_10d_WW1 F18_10d_WW2
  4.307786   4.261419   4.248699   4.239323   4.289799
F18_10d_WW3 F18_10d_WW4 F18_34d_WS1 F18_34d_WS2 F18_34d_WS3
  4.620659   4.532309   4.114948   4.149386   4.350818
F18_34d_WS4 F18_34d_WW1 F18_34d_WW2 F18_34d_WW3 F18_34d_WW4
  4.075166   4.239367   4.109161   4.460006   4.507653
F18_67d_WS1 F18_67d_WS2 F18_67d_WS3 F18_67d_WS4 F18_67d_WW1
  4.443768   4.392282   4.258226   4.311732   4.659167
F18_67d_WW2 F18_67d_WW3 F18_67d_WW4 C19_22d_WS1 C19_22d_WS2
  4.584738   4.404701   4.488189   5.160000   5.040000
C19_22d_WS3 C19_22d_WS4 C19_22d_WW1 C19_22d_WW2 C19_22d_WW3
  4.950000   4.920000   5.060000   4.710000   5.410000
C19_22d_WW4 C19_44d_WS1 C19_44d_WS2 C19_44d_WS3 C19_44d_WS4
  5.070000   4.420000   4.480000   4.490000   4.570000
C19_44d_WW1 C19_44d_WW2 C19_44d_WW3 C19_44d_WW4 F19_22d_WS1
  4.930000   5.000000   4.820000   4.770000   5.560000
F19_22d_WS2 F19_22d_WS3 F19_22d_WS4 F19_22d_WW1 F19_22d_WW2
  5.150000   5.130000   5.690000   5.060000   5.160000
F19_22d_WW3 F19_22d_WW4 F19_44d_WS1 F19_44d_WS2 F19_44d_WS3
  5.250000   4.940000   4.710000   4.320000   4.910000
F19_44d_WS4 F19_44d_WW1 F19_44d_WW2 F19_44d_WW3 F19_44d_WW4
  4.540000   5.030000   4.700000   4.820000   4.720000

> means <- tapply(y,pd[,c("date","treat","variety")],mean,na.rm=TRUE)
> means

```

```
, , variety = C
```

	treat	
date	WW	WS
2018-06-11	NA	NA
2018-06-12	4.286961	4.023442
2018-07-05	4.228135	4.037203
2018-08-07	4.415919	4.225334
2019-06-26	5.062500	5.017500
2019-07-18	4.880000	4.490000

```
, , variety = F
```

	treat	
date	WW	WS

```

2018-06-11 4.420523 4.230104
2018-06-12      NA      NA
2018-07-05 4.329047 4.172579
2018-08-07 4.534199 4.351502
2019-06-26 5.102500 5.382500
2019-07-18 4.817500 4.620000

> tfilter <- wpas$treat %in% "WW"
> vfilter <- wpas$variety %in% "C"
> (swp <- wpas$swp[as.character(wpas$date) %in% dimnames(means)[[1]]])
[1] -0.3680000 -0.4527273 -0.2872727 -0.3472727 -0.3720000 -0.4645455
[7] -0.2854545 -0.3554545 -0.4016667 -0.6425000 -0.2700000 -0.7983333
[13] -0.5350000 -1.4750000 -0.4300000 -1.1600000 -0.5000000 -1.0400000
[19] -0.4100000 -0.8300000 -0.4592308 -1.1419231 -0.4210000 -1.1390000

> varname <- rownames(t1819)[10]
> #varname <- rownames(t1819)[runif(1,1,10000)]
> (y <- unlist(t1819[varname,]))
C18_11d_WS1 C18_11d_WS2 C18_11d_WS3 C18_11d_WS4 C18_11d_WW1
4.088427 3.961216 4.127170 3.916955 4.472212
C18_11d_WW2 C18_11d_WW3 C18_11d_WW4 C18_34d_WS1 C18_34d_WS2
4.309540 4.212060 4.154032 4.086692 3.914995
C18_34d_WS3 C18_34d_WS4 C18_34d_WW1 C18_34d_WW2 C18_34d_WW3
4.065776 4.081348 4.313965 4.041339 4.318997
C18_34d_WW4 C18_67d_WS1 C18_67d_WS2 C18_67d_WS3 C18_67d_WS4
4.238240 4.441597 4.248104 4.242540 3.969094
C18_67d_WW1 C18_67d_WW2 C18_67d_WW3 C18_67d_WW4 F18_10d_WS1
4.160356 4.676748 4.467201 4.359372 4.102511
F18_10d_WS2 F18_10d_WS3 F18_10d_WS4 F18_10d_WW1 F18_10d_WW2
4.307786 4.261419 4.248699 4.239323 4.289799
F18_10d_WW3 F18_10d_WW4 F18_34d_WS1 F18_34d_WS2 F18_34d_WS3
4.620659 4.532309 4.114948 4.149386 4.350818
F18_34d_WS4 F18_34d_WW1 F18_34d_WW2 F18_34d_WW3 F18_34d_WW4
4.075166 4.239367 4.109161 4.460006 4.507653
F18_67d_WS1 F18_67d_WS2 F18_67d_WS3 F18_67d_WS4 F18_67d_WW1
4.443768 4.392282 4.258226 4.311732 4.659167
F18_67d_WW2 F18_67d_WW3 F18_67d_WW4 C19_22d_WS1 C19_22d_WS2
4.584738 4.404701 4.488189 5.160000 5.040000
C19_22d_WS3 C19_22d_WS4 C19_22d_WW1 C19_22d_WW2 C19_22d_WW3
4.950000 4.920000 5.060000 4.710000 5.410000
C19_22d_WW4 C19_44d_WS1 C19_44d_WS2 C19_44d_WS3 C19_44d_WS4
5.070000 4.420000 4.480000 4.490000 4.570000
C19_44d_WW1 C19_44d_WW2 C19_44d_WW3 C19_44d_WW4 F19_22d_WS1
4.930000 5.000000 4.820000 4.770000 5.560000
F19_22d_WS2 F19_22d_WS3 F19_22d_WS4 F19_22d_WW1 F19_22d_WW2
5.150000 5.130000 5.690000 5.060000 5.160000
F19_22d_WW3 F19_22d_WW4 F19_44d_WS1 F19_44d_WS2 F19_44d_WS3
5.250000 4.940000 4.710000 4.320000 4.910000
F19_44d_WS4 F19_44d_WW1 F19_44d_WW2 F19_44d_WW3 F19_44d_WW4
4.540000 5.030000 4.700000 4.820000 4.720000

> means <- tapply(y, pd[,c("date", "treat", "variety")], mean, na.rm=TRUE)
> swps <- tapply(pd$swp, pd[,c("date", "treat", "variety")], unique)

```

```

Error in tapply(pd$swp, pd[, c("date", "treat", "variety")], unique): argument
> means
, , variety = C

      date      treat      WW      WS
1 2018-06-11      NA      NA      NA
2 2018-06-12 4.286961 4.023442
3 2018-07-05 4.228135 4.037203
4 2018-08-07 4.415919 4.225334
5 2019-06-26 5.062500 5.017500
6 2019-07-18 4.880000 4.490000

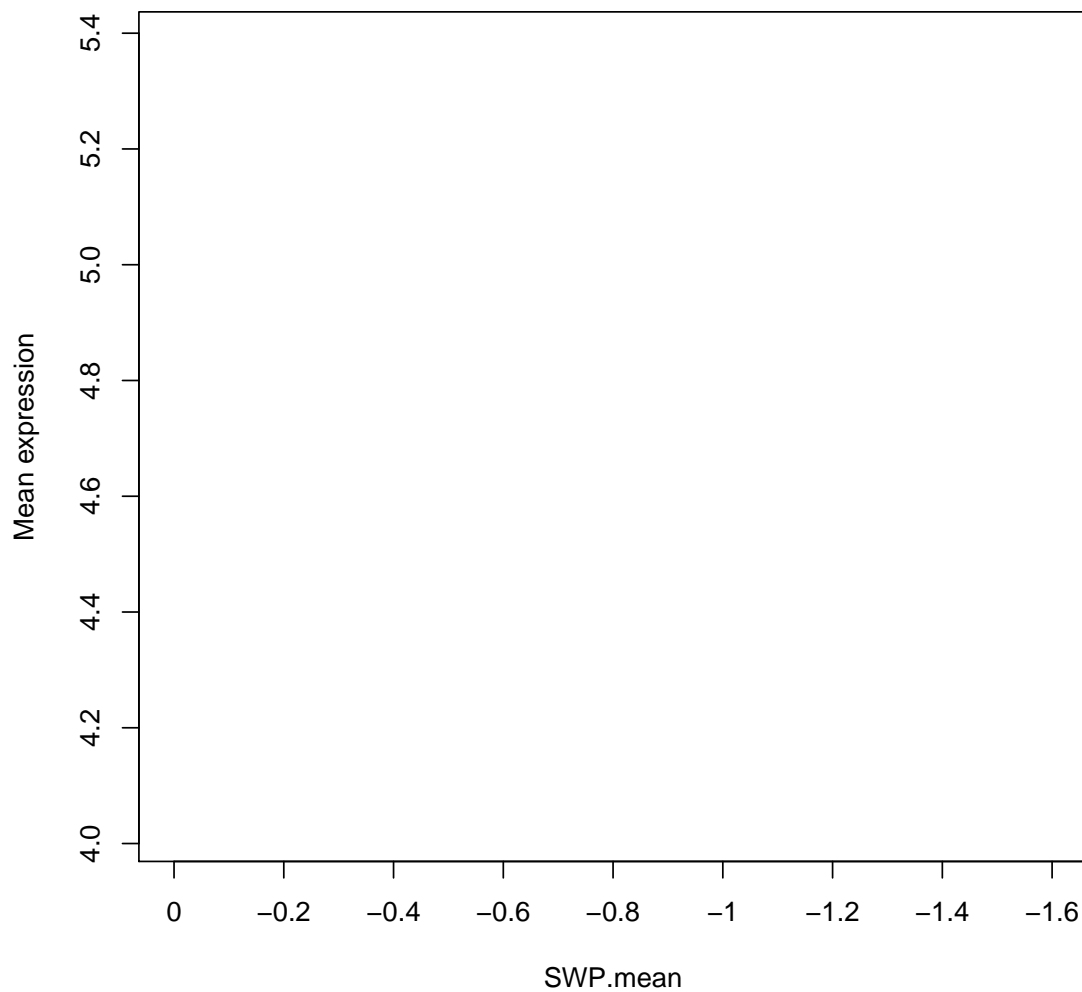
, , variety = F

      date      treat      WW      WS
1 2018-06-11 4.420523 4.230104
2 2018-06-12      NA      NA
3 2018-07-05 4.329047 4.172579
4 2018-08-07 4.534199 4.351502
5 2019-06-26 5.102500 5.382500
6 2019-07-18 4.817500 4.620000

> ylim <- range(means, na.rm=TRUE)
> xlim <- c(min(wpas$swp, -1.6, na.rm=TRUE), max(wpas$swp, 0, na.rm=TRUE))
> vlvl <- unique(wpas$variety)
> tlvl <- rev(unique(wpas$treat))
> cols <- c(4, 2)
> pchs <- c(16, 16)
> plot(1, 1, #wpas$swp[tfilter & vfilter], means[, "WS", vlvl[i]],
+      ylim=ylim,
+      xlim=-rev(xlim),
+      xlab = "SWP.mean",
+      ylab = "Mean expression",
+      type="n",
+      axes=FALSE
+      )
> axis(2)
> box()
> axis(1, at=seq(0, 2, 0.2), labels=- (seq(0, 2, 0.2)))
> title(varname)

```

## Vitvi07g02767



```
> legend("topright", bty="n", pch=pchs, col=cols, legend <- vlvs)
Error in as.graphicsAnnot(legend): argument "legend" is missing, with no default
> #
> for(i in 1:length(vlvs)){
+ for( j in 1:length(tlvls)){
+ tfilter <- wpa$treat %in% tlvls[j]
+ vfilter <- wpa$variety %in% vlvs[i]
+ yy <- means[as.character(wpa$date), tlvls[j], vlvs[i]]
+ wpa <- wpa[tfilter & vfilter, ]
+ all(wpa$date==names(yy))
+ points(-wpa$swp, yy,
+       col = cols[j],
+       pch = pchs[j]
+     )
+   }
+ }
Error in eval(expr, envir, enclos): object 'wpa' not found
```

## 4.1 Expression vs water potential

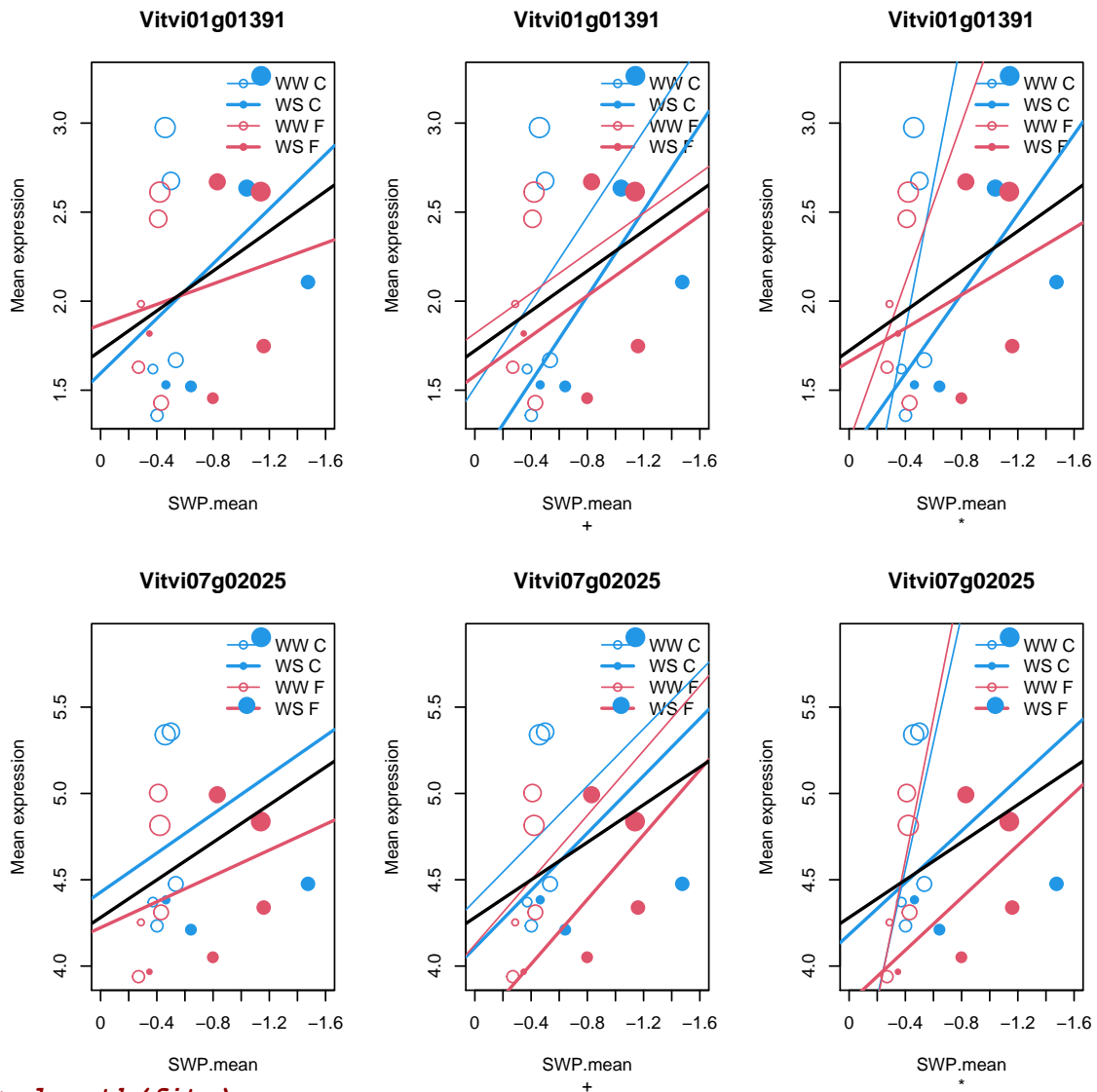
```
> plot.ewp <- function(varname, cols=c(4, 2), pchs=c(1, 16), lwd=c(1, 2, 1, 2)
+   , model=c("*"), verbose=FALSE) {
+   if(verbose) print(varname)
+   (y <- unlist(t1819[varname, ]))
+   means <- tapply(y, pd[, c("date", "treat", "variety")], mean, na.rm=TRUE)
+   means
+   swps <- tapply(wpas$swp, wpas[, c("date", "treat", "variety")], function(x) x[1])
+   swps
+   ylim <- range(means, na.rm=TRUE)
+   xlim <- c(min(wpas$swp, -1.6, na.rm=TRUE), max(wpas$swp, 0, na.rm=TRUE))
+   vlvs <- levels(wpas$variety)
+   tlvs <- levels(wpas$treat)
+   #cols <- c(4, 2)
+   #pchs <- c(1, 16)
+   plot(1, 1, #wpas$swp[tfilter & vfilter], means[, "WS", vlvs[i]],
+     ylim=ylim,
+     xlim=-rev(xlim),
+     xlab = "SWP.mean",
+     ylab = "Mean expression",
+     type="n",
+     axes=FALSE
+   )
+   axis(2)
+   box()
+   axis(1, at=seq(0, 2, 0.2), labels=-seq(0, 2, 0.2))
+   title(varname, model)
+   legend("topright", bty="n", pch=pchs, col=rep(cols, each=2), lwd=lwd,
+     legend= outer(tlvs, vlvs, paste))
+   #
+   allx <- NULL
+   ally <- NULL
+   fits <- NULL
+   fits <- list()
+   for(i in 1:length(vlvs)) {
+     #for( j in 1:length(tlvs)) {
+     #tfilter <- wpas$treat %in% tlvs[j]
+     vfilter <- wpas$variety %in% vlvs[i]
+     wpa <- wpas[vfilter, ]
+     means[, , vlvs[i]]
+     mns <- melt(means[, , vlvs[i]])
+     mns
+     wp <- melt(swps[, , vlvs[i]])
+     wp
+     all(wpa$date==dimnames(mns)$date)
+     yy <- mns[, "value"]
+     xx <- wp[, "value"]
+     points(-xx , yy,
+       col = cols[i],
+       pch = pchs[as.numeric(mns$treat)]
+     , cex=0.5+as.numeric(mns$date)/3
+   )
+ }
```

```

+ #      }
+   xx <- -xx
+   if(model=="") fit <- lm(yy~xx)
+   if(model=="+") fit <- lm(yy~xx+mns$treat)
+   if(model=="*") fit <- lm(yy~xx*mns$treat)
+   if(verbose) print(summary(fit))
+   coef <- fit$coefficients
+   if(length(coef)<3) coef[3] <- 0
+   if(length(coef)<4) coef[4] <- 0
+   abline(coef[1:2], col=cols[i],lwd=1)
+   abline(coef[1:2]+coef[3:4],col=cols[i],lwd=2)
+   ally <- c(ally,yy)
+   allx <- c(allx,xx)
+   fits[[vlvls[i]]] <- fit
+ }
+ fit <- lm(ally~allx)
+ fits[["all"]] <- fit
+ fits[["r"]] <- cor(ally,allx,use="complete")
+ if(verbose) print(summary(fit))
+ abline(fit$coefficients,lwd=2)
+ #points(-swps,means,cex=1.5)
+ invisible(fits)
+ }

> par(mfrow=c(2,3))
> #
> #varname <- rownames(t1819)[10]
> varnames <- c("Vitvi03g01254")
> set.seed(1234)
> for(vari in 1:2){
+ varname <- rownames(t1819)[runif(1,1,10000)]
+ plot.ewp(varname,model="")
+ plot.ewp(varname,model="+")
+ fits <- plot.ewp(varname,model="*")
+ }

```



```

> length(fits)
[1] 4
> names(fits)
[1] "C" "F" "all" "r"

```

## 4.2 Water potential vs Expression

```

> plot.wpe <- function(varname, cols=c(4, 2), pchs=c(1, 16), lwd=c(1, 2, 1, 2)
+ , model=c("*"), verbose=FALSE) {
+   if(verbose) print(varname)
+   (y <- unlist(t1819[varname, ]))
+   means <- tapply(y, pd[, c("date", "treat", "variety")], mean, na.rm=TRUE)
+   means
+   swps <- tapply(wpas$swp, wpas[, c("date", "treat", "variety")], function(x) x[1])
+   swps
+   ylim <- range(means, na.rm=TRUE)
+   xlim <- c(min(wpas$swp, -1.6, na.rm=TRUE), max(wpas$swp, 0, na.rm=TRUE))
+   vlvs <- levels(wpas$variety)
+   tlvs <- levels(wpas$treat)
+   #cols <- c(4, 2)

```

```

+ #pchs <- c(1,16)
+ plot(1,1, #wpas$swp[tfilter & vfilter], means[, "WS", vlvls[i]],
+      xlim=ylim,
+      ylim=-rev(xlim),
+      ylab = "SWP.mean",
+      xlab = "Mean expression",
+      type="n",
+      axes=FALSE
+      )
+ axis(1)
+ box()
+ axis(2, at=seq(0, 2, 0.2), labels=-(seq(0, 2, 0.2)))
+ title(varname, model)
+ legend("topright", bty="n", pch=pchs, col=rep(cols, each=2), lwd=lwd,
+       legend= outer(tlvls, vlvls, paste))
+ #
+ ally <- NULL
+ allx <- NULL
+ fits <- NULL
+ for(i in 1:length(vlvls)){
+ #for( j in 1:length(tlvls)){
+ #tfilter <- wpas$treat %in% tlvls[j]
+ vfilter <- wpas$variety %in% vlvls[i]
+ wpa <- wpas[vfilter, ]
+ means[, , vlvls[i]]
+ mns <- melt(means[, , vlvls[i]])
+ mns
+ wp <- melt(swps[, , vlvls[i]])
+ wp
+ all(wpa$date==dimnames(mns)$date)
+ yy <- mns[, "value"]
+ xx <- wp[, "value"]
+ points(yy, -xx,
+        col = cols[i],
+        pch = pchs[as.numeric(mns$treat)]
+        , cex=0.5+as.numeric(mns$date)/3
+        )
+ #      }
+   xx <- -xx
+   if(model=="") fit <- lm(xx~yy)
+   if(model=="+") fit <- lm(xx~yy+mns$treat)
+   if(model=="*") fit <- lm(xx~yy*mns$treat)
+ #
+   if(verbose) print(summary(fit))
+   coef <- fit$coefficients
+   if(length(coef)<3) coef[3] <- 0
+   if(length(coef)<4) coef[4] <- 0
+   abline(coef[1:2], col=cols[i], lwd=1)
+   abline(coef[1:2]+coef[3:4], col=cols[i], lwd=2)
+   ally <- c(ally, yy)
+   allx <- c(allx, xx)
+   fits[[vlvls[i]]] <- fit
+ }

```

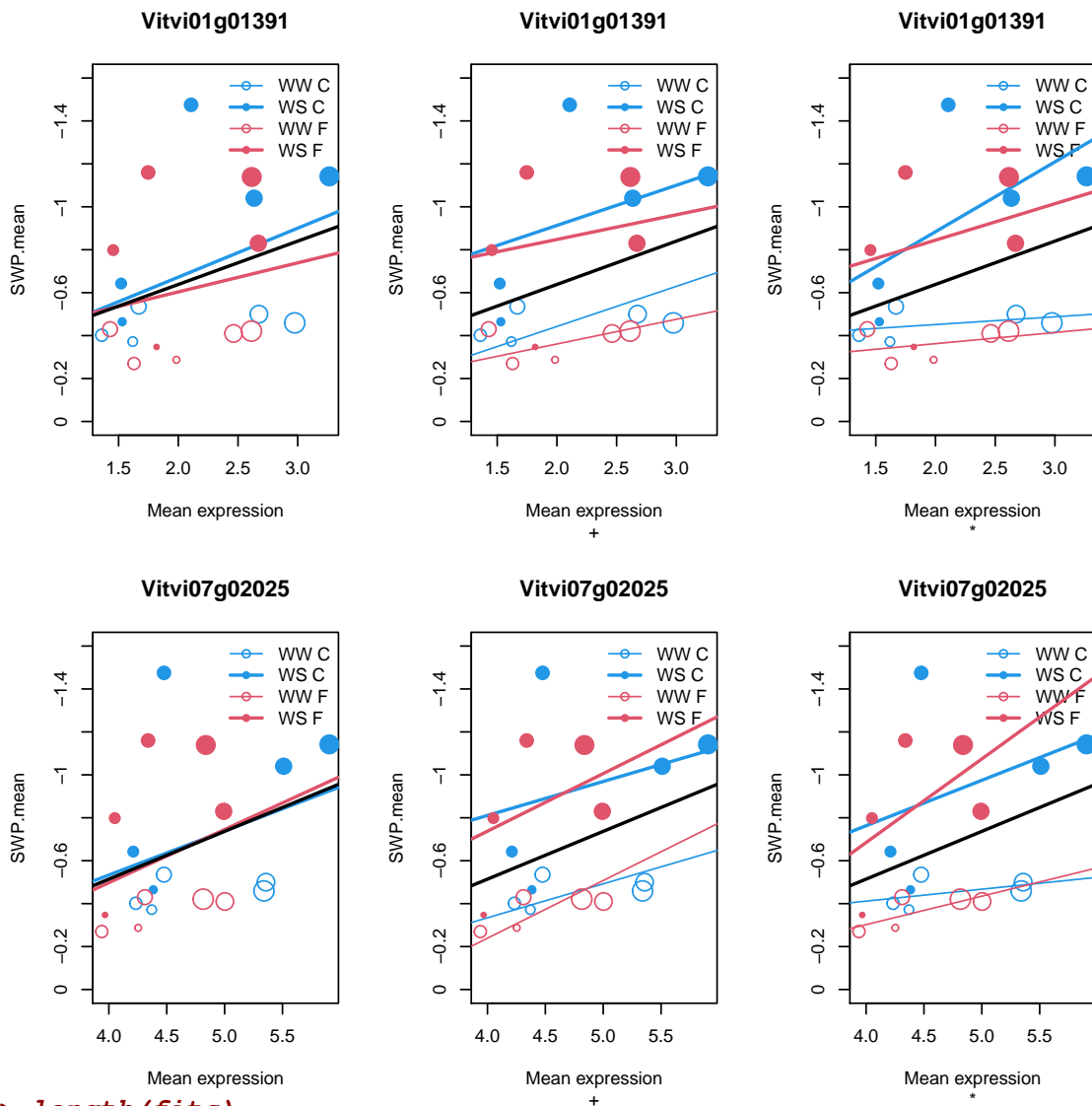


```

+ fit <- lm(allx~ally)
+ fits[["all"]] <- fit
+ if (verbose) print(summary(fit))
+ abline(fit$coefficients, lwd=2)
+ #points(-swps, means, cex=1.5)
+ invisible(fits)
+ }

> par(mfrow=c(2, 3))
> #
> #varname <- rownames(t1819)[10]
> varname <- c("Vitvi03g01254")
> set.seed(1234)
> for(vari in 1:2){
+ varname <- rownames(t1819)[runif(1, 1, 10000)]
+ plot.wpe(varname, model="")
+ plot.wpe(varname, model="+")
+ fits <- plot.wpe(varname, model="*")
+ }

```



```

> length(fits)

```

```
[1] 3
```

```
> names(fits)
```

```
[1] "C" "F" "all"
```

```
> par(mfrow=c(2,3))
```

```
> #
```

```
> #varname <- rownames(t1819)[10]
```

```
> varnames <- c("Vitvi03g01254")
```

```
> #set.seed(1234)
```

```
> varname <- rownames(t1819)[runif(1,1,10000)]
```

```
> plot.wpe(varname,model="")
```

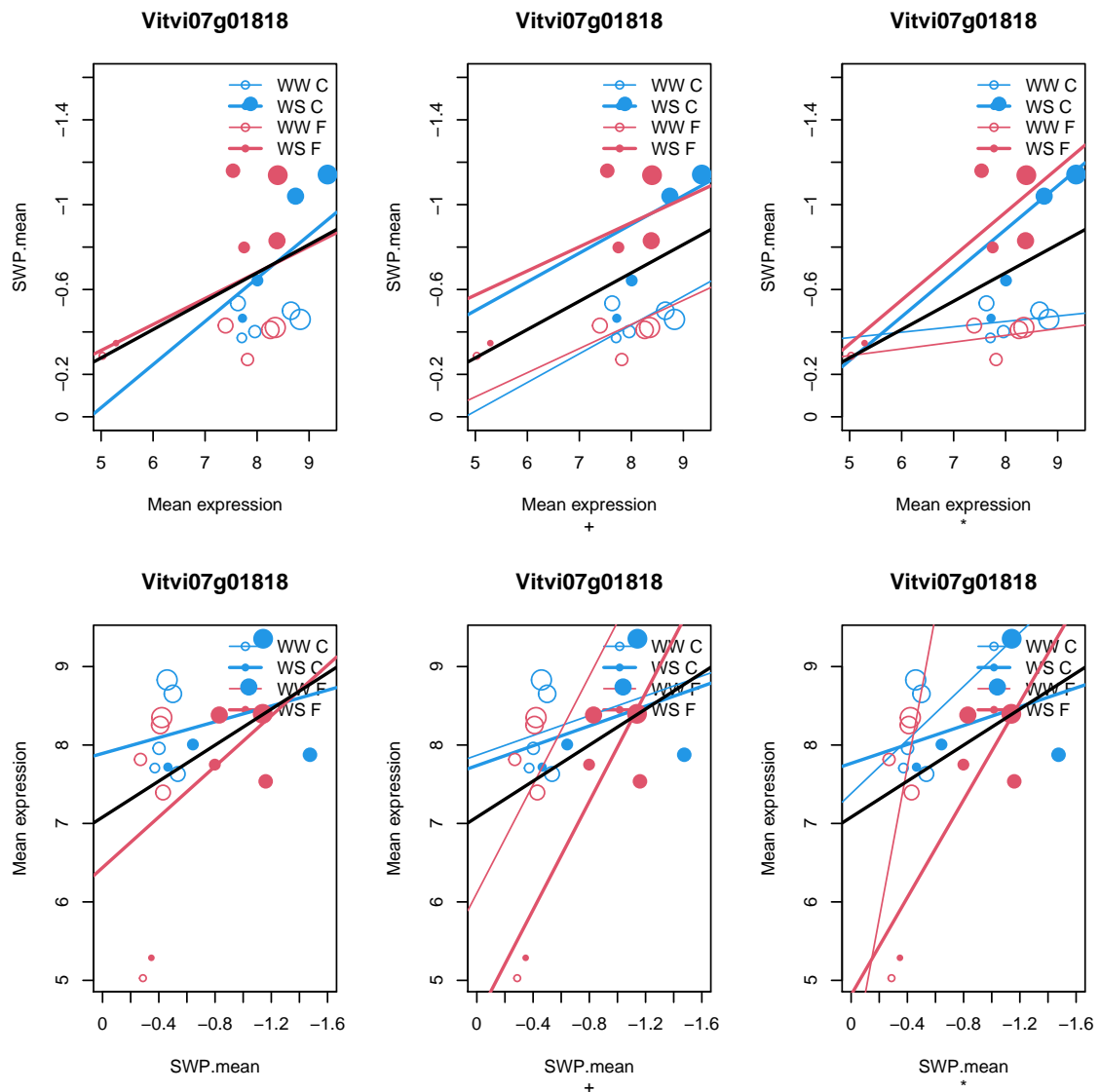
```
> plot.wpe(varname,model="+")
```

```
> plot.wpe(varname,model="*")
```

```
> plot.ewp(varname,model="")
```

```
> plot.ewp(varname,model="+")
```

```
> plot.ewp(varname,model="*")
```



```

> for(i in 1:2){
+   ( yy <- as.vector( ( means[as.character(wpa$date), , vlvls[i]] ) ) )
+   xx <- -rep(wpa$swp, length(tlvls))[!is.na(yy)]
+   yy <- yy[!is.na(yy)]
+   plot(xx, yy, pch=16, col=cols[i], xlim=-xlim, ylim=ylim)
+   fit <- lm(yy ~ xx)
+   abline(fit, col=cols[i])
+ }
Error in as.vector((means[as.character(wpa$date), , vlvls[i]])): object 'wpa'

```

## 5 Linear models for water potential

```

> library(limma)
Warning: package 'limma' was built under R version 4.0.3

Attaching package: 'limma'

The following object is masked from 'package:BiocGenerics':

    plotMA

> exprs <- t1819
> pd <- pdata1819w
> cat(knit_child(file.path("../doc/", "40a_limma-fit-swp.Rnw"), quiet=TRUE))

```

## 5.1 Model design

Simple model with water potential for all data:

```
> design <- with(pd, model.matrix(~ swp))
> design[,2] <- (-design[,2])
> head(design)
  (Intercept)    swp
1           1 0.372
2           1 0.372
3           1 0.372
4           1 0.372
5           1 0.372
6           1 0.372

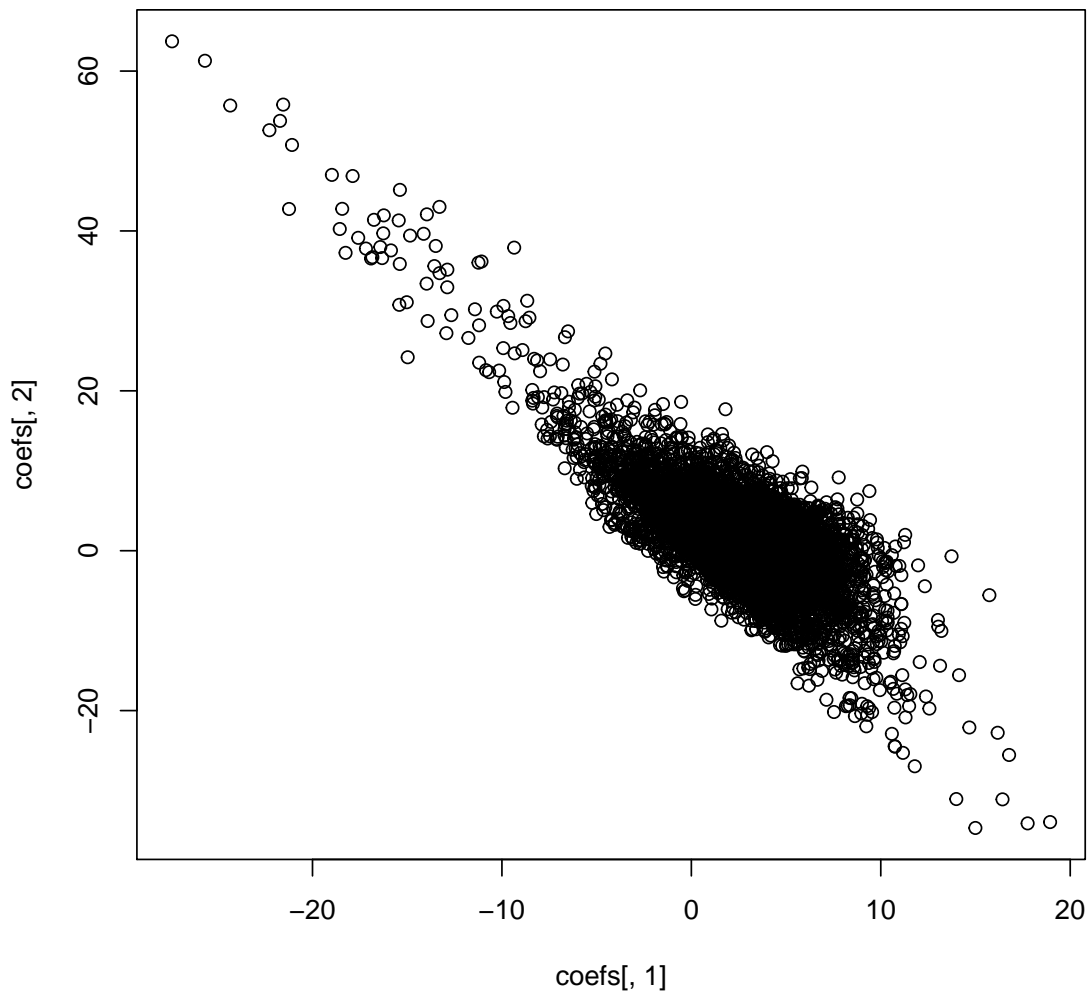
> tail(design)
  (Intercept)    swp
75           1 0.4592308
76           1 0.4592308
77           1 0.4592308
78           1 0.4592308
79           1 0.4592308
80           1 0.4592308
```

Linear models

```
> fit1 <- lmFit(exprs, design)
> fit1 <- eBayes(fit1)
> head(fit1[1:6,]$coefficients)
              (Intercept)            swp
Vitvi15g01736  7.7878435 -7.3063828
Vitvi07g02832  0.3638262 -1.2060744
Vitvi07g02830  1.6475442 -4.3513888
Vitvi07g02812 -0.1783863 -0.3706094
Vitvi07g02811  2.9235068 -0.9628080
Vitvi09g02033 -0.1698934 -1.3624505

> fit <- fit1

> coefs <- fit$coefficients
> plot(coefs[,1], coefs[,2])
```



```
> coefs[abs(coefs[, 2])>20, ]
```

	(Intercept)	swp
Vitvi01g01048	11.306956	-20.85587
Vitvi01g01512	-11.078026	36.17683
Vitvi01g01708	-7.456518	23.93461
Vitvi01g02286	7.528961	-20.17774
Vitvi02g00025	-14.133401	39.64396
Vitvi02g00220	9.289836	-20.53148
Vitvi02g00263	-9.877397	21.10158
Vitvi02g00374	-10.270324	29.90913
Vitvi02g00436	-10.674876	22.33529
Vitvi02g01446	-12.881222	35.15716
Vitvi02g01447	-21.237525	42.74610
Vitvi02g01121	-5.533024	20.87824
Vitvi03g00325	17.753139	-34.10975
Vitvi03g00717	-6.673047	26.70677
Vitvi04g01792	-8.549161	29.15129
Vitvi04g01794	-17.179458	37.79848
Vitvi04g01795	-16.420240	37.98288

Vitvi04g01799	-16.324341	36.60426
Vitvi04g01801	-17.586092	39.13847
Vitvi04g01802	-15.392201	35.87135
Vitvi04g00135	-25.677235	61.28924
Vitvi04g00501	10.734803	-24.45325
Vitvi04g00522	-5.970696	20.70966
Vitvi04g01654	-12.671731	29.46865
Vitvi05g00170	-18.560034	40.24486
Vitvi05g01833	-8.396689	20.10726
Vitvi05g00486	-13.565366	35.60306
Vitvi05g01933	-10.842490	22.58200
Vitvi05g01353	16.171054	-22.77537
Vitvi06g00443	-4.540227	24.67175
Vitvi06g00561	-21.081489	50.75508
Vitvi07g02123	11.170024	-25.28741
Vitvi07g00380	-18.432074	42.76737
Vitvi07g02214	-13.919915	28.73382
Vitvi07g00445	10.585392	-22.90773
Vitvi07g00457	-16.236313	41.95000
Vitvi07g02242	-14.847715	39.42011
Vitvi07g00657	-9.328816	24.68773
Vitvi07g02433	-6.787508	23.28713
Vitvi07g01120	-10.145850	22.53412
Vitvi08g00689	-8.300061	24.00508
Vitvi08g00731	-18.253897	37.26034
Vitvi08g01143	-5.114917	22.39484
Vitvi08g02274	-22.273550	52.60215
Vitvi09g00045	-8.748078	28.70774
Vitvi09g00599	-16.755009	41.39061
Vitvi10g00027	16.773920	-25.55267
Vitvi10g00436	8.632955	-20.69019
Vitvi10g00496	-4.796882	23.39236
Vitvi10g01881	-15.416105	30.75833
Vitvi10g01886	10.748013	-24.51282
Vitvi11g00724	-15.384832	45.13777
Vitvi11g01645	11.792997	-26.96392
Vitvi12g00574	-14.974719	24.19775
Vitvi12g00576	-15.448071	41.31687
Vitvi12g02138	-21.714093	53.77021
Vitvi13g00278	-9.919179	30.62073
Vitvi13g02005	16.426164	-31.10367
Vitvi13g02014	-9.551786	28.48403
Vitvi13g02016	-15.857617	37.56527
Vitvi13g02017	-16.263225	39.70164
Vitvi13g02018	-13.493238	38.12168
Vitvi13g00405	-16.906999	36.55759
Vitvi13g00409	-13.290217	34.71895
Vitvi13g00410	-13.296353	43.00946
Vitvi13g02024	-13.962153	42.07724
Vitvi13g00451	-11.236659	36.03119
Vitvi13g00471	-16.839013	36.71473
Vitvi13g00490	-12.884408	32.95523
Vitvi13g01107	-9.667692	29.34786

```

Vitvi13g01839 -11.200693 23.52176
Vitvi14g00063 -15.023680 31.08035
Vitvi14g01381 14.994303 -34.68278
Vitvi14g01469 18.938154 -33.94468
Vitvi14g02990 -11.425592 30.20039
Vitvi14g01942 -18.975683 47.01313
Vitvi14g02026 -2.702445 20.05851
Vitvi15g00714 9.238437 -21.96631
Vitvi15g01034 -9.352648 37.91364
Vitvi16g00681 -6.508296 27.43906
Vitvi16g00890 9.542375 -20.18448
Vitvi16g01103 -17.880477 46.87118
Vitvi16g01985 -12.930117 27.21067
Vitvi16g01352 -21.545840 55.80582
Vitvi17g01318 -13.974271 33.40743
Vitvi17g00601 13.987170 -31.06433
Vitvi17g00695 -8.661915 31.26828
Vitvi17g01596 8.975235 -20.36075
Vitvi18g02468 -8.914837 25.09923
Vitvi18g00268 -9.928831 25.34296
Vitvi18g01510 -8.136715 23.79546
Vitvi18g01966 -5.065014 20.56758
Vitvi18g02066 -4.194130 21.43313
Vitvi18g02423 -27.414763 63.72049
Vitvi19g00041 14.675960 -22.11331
Vitvi19g00514 -7.985757 22.44871
Vitvi19g02145 -11.206279 28.19821
Vitvi19g01048 -11.768042 26.61329
Vitvi19g01760 -24.342365 55.69011

```

Top table

```

> which <- 2
> tt <- topTable(fit,coef=which, number=Inf)
> head(tt)

```

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi02g00374	29.90913	3.283880	18.79698	2.300407e-31	3.506281e-27
Vitvi04g00522	20.70966	3.414499	17.94327	4.900852e-30	3.734939e-26
Vitvi09g01273	17.79031	3.745035	17.75236	9.830851e-30	3.831175e-26
Vitvi11g00724	45.13777	5.070678	17.74622	1.005426e-29	3.831175e-26
Vitvi01g01943	12.14569	4.556800	17.51369	2.361916e-29	6.767539e-26
Vitvi08g00689	24.00508	2.578551	17.48107	2.664036e-29	6.767539e-26

B

Vitvi02g00374	55.20282
Vitvi04g00522	52.74622
Vitvi09g01273	52.18207
Vitvi11g00724	52.16383
Vitvi01g01943	51.46908
Vitvi08g00689	51.37095

### 5.1.1 Export output files

Export fit in a form:

```
> fitfn <- "lm-fit-statistics.txt"
> write.fit(head(fit), file="", adjust="BH")

      A          Coef. (Intercept)          Coef. swp          t. (Intercept)
Vitvi15g01736      4.47674068175      7.78784348921542      -7.306382793
Vitvi07g02832     -0.1827419310625      0.363826241712366      -1.20607
Vitvi07g02830     -0.3244159185      1.64754420617268      -4.351388755
Vitvi07g02812     -0.3463388427625     -0.178386273275306      -0.3706
Vitvi07g02811      2.4871819448375      2.92350680207491      -0.9628080
Vitvi09g02033     -0.78732802375     -0.169893388093836     -1.362450

> write.fit(fit, file=file.path(.oroot, fitfn), adjust="BH")
```

Export top table

```
> ttfn <- "TopTable.txt"
> my.write.table(tt, file=file.path(.oroot, ttfn))
```

Warning in write.table(x, file = file, col.names = col.names, sep = sep, : appending column names to file Object: tt

Label:

File :

.././output/40\_Expression-water-stress-0.05-r/TopTable.txt

```
> par(mfrow=c(2, 3))
> cf <- 10
> ns <- c(500, 300, 200, 100, 50, 25)
> for(i in 1:length(ns)) {
+ plot(coefs[, 1], coefs[, 2], xlab="Intercept", ylab="Slope")
+ filter <- rownames(tt)[1:ns[i]]
+ points(coefs[filter, 1], coefs[filter, 2], col=i+1, pch=16)
+ abline(h=c(-cf, cf), col="white")
+ title(paste("Top ", ns[i]))
+ }
```

Many significant genes:

```
> .alpha <- 0.05
> sum(tt$adj.P.Val < .alpha)
[1] 9004
> sum((tt$adj.P.Val < .alpha) & (abs(tt$logFC) > cf))
[1] 607
> sum((tt$adj.P.Val < .alpha) & (abs(tt$logFC) > 2*cf))
[1] 99
```

We should consider to set a threshold for effect (slope), e.g. absolute slope larger than certain threshold. In the figure a cutoff 10 is used.

Z apregled je boljše pogledati prvih nekaj in zadnjih nekaj.

```
> topn <- 20
```



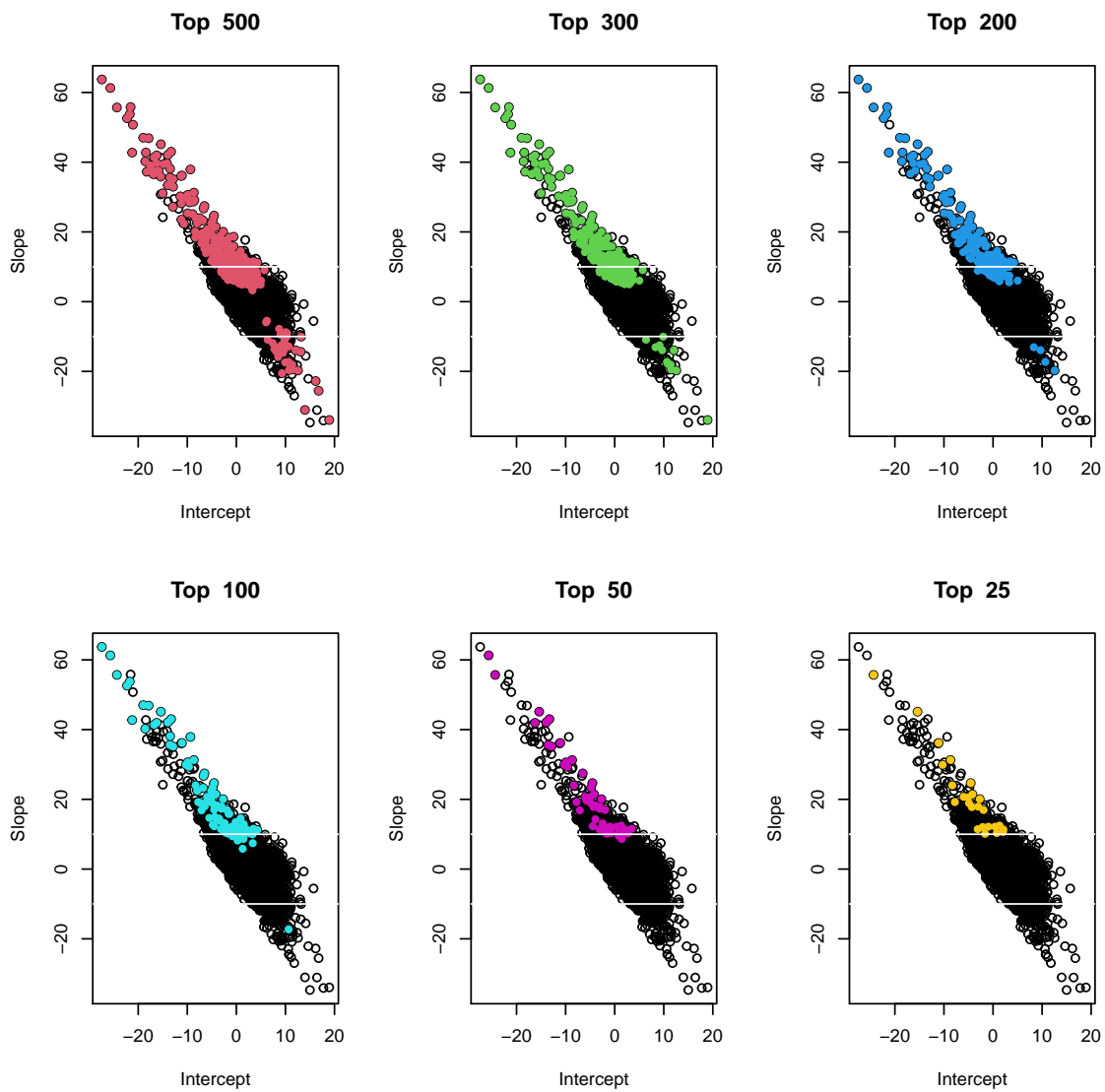


Figure 1: Coefficients for labeled number of top significant genes are indicated by colors. White line is a proposed threshold.

## 5.2 Top 20 genes

Plots of situation for top 20 genes.

Figures caption:

Mean expression (E) and mean water potential (WP). Upper panels show WP vs E and regression lines for three models. Left: all C (blue) and all F (blue). Middle: separate models for combinations of treatment and variety, same slope (+). Right: separate for combinations, different slopes (interaction, \*). Black line is regression of all data, regardless of treatment and variety. Lower panels have swapped variables: E vs WP. Symbol sizes are related to time, large is later.

```
> out <- ""
> (cname <- colnames(coefs)[2])
[1] "swp"
> varnames <- rownames(head(tt,topn))
> varname <- varnames[1]
> for(varname in varnames)
+ out <- paste(out,knit_child(file.path("../doc/","40aa_one-gene-plots.Rnw"),
```

## 5.2.1 Vitvi02g00374

```
> cat.feature(fdata[fdata$geneID %in% varname, 2:5])
```

Vitvi02g00374

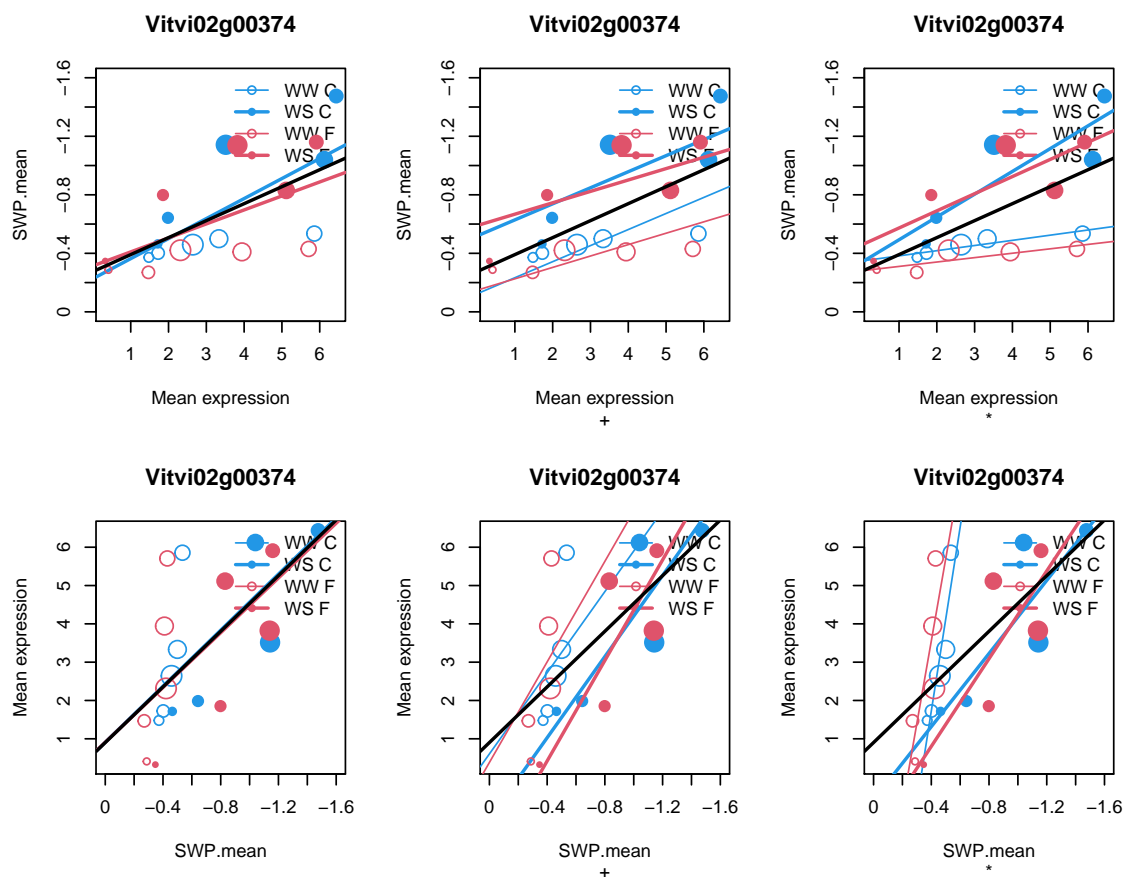
29.05.2011

protein.degradation.ubiquitin

UBX domain-containing protein IPR001012 UBX

Coefficient swp for Vitvi02g00374.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi02g00374	29.90913	3.28388	18.79698	2.300407e-31	3.506281e-27
B					
Vitvi02g00374	55.20282				



Correlation of all measurements (not just means):  $r = 0.9026$ ,  $r^2 = 0.8147$

Correlation of mean values (plots):  $r_2 = 0.6490$

## 5.2.2 Vitvi04g00522

```
> cat.feature(fdata[fdata$geneID %in% varname, 2:5])
```

Vitvi04g00522

20.02.2001

stress.abiotic.heat

casein lytic proteinase B4 |

Chr2:10697877-10701998 REVERSE LENGTH=964 |

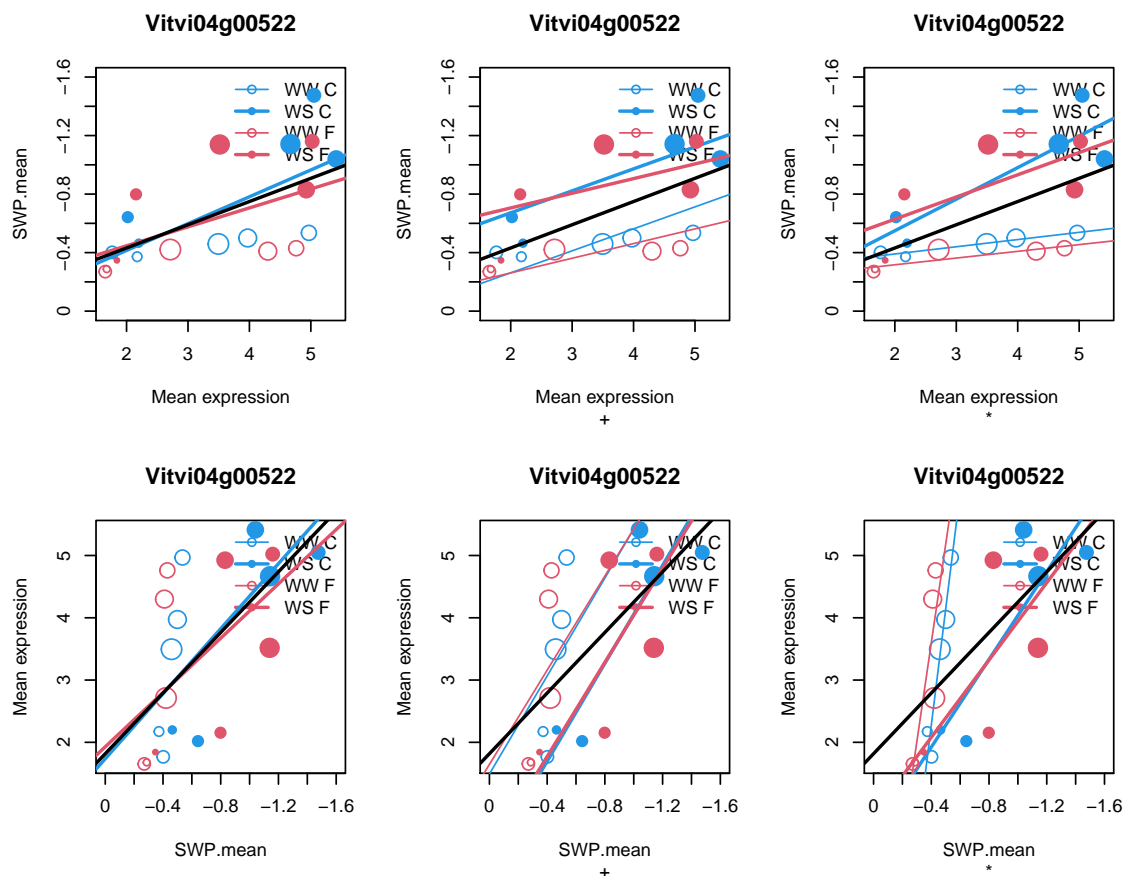
201606

Coefficient swp for Vitvi04g00522.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi04g00522	20.70966	3.414499	17.94327	4.900852e-30	3.734939e-26

B

Vitvi04g00522 52.74622



Correlation of all measurements (not just means):  $r = 0.8957, r^2 = 0.8022$

Correlation of mean values (plots):  $r_2 = 0.6215$

### 5.2.3 Vitvi09g01273

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi09g01273

35.2

not assigned.unknown

ATP-dependent DNA helicase |

Chr4:14990523-14992855 FORWARD LENGTH=589 |

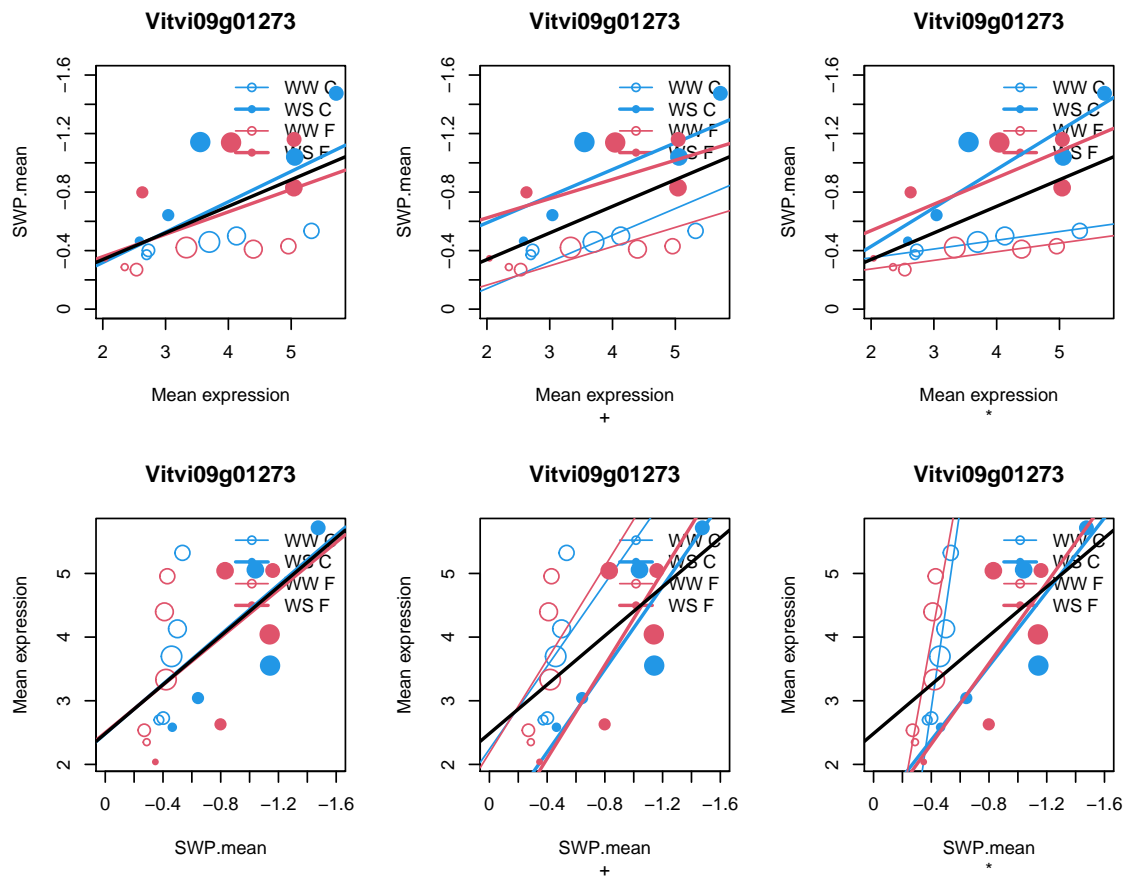
201606

Coefficient swp for Vitvi09g01273.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi09g01273	17.79031	3.745035	17.75236	9.830851e-30	3.831175e-26

B

Vitvi09g01273 52.18207



Correlation of all measurements (not just means):  $r = 0.8945$ ,  $r^2 = 0.8002$

Correlation of mean values (plots):  $r_2 = 0.5909$

## 5.2.4 Vitvi11g00724

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi11g00724

20.02.2001

stress.abiotic.heat

heat shock protein 70B |

Chr1:5502386-5504326 REVERSE LENGTH=646 |

201606

Vitvi11g00724

29.6

protein.folding

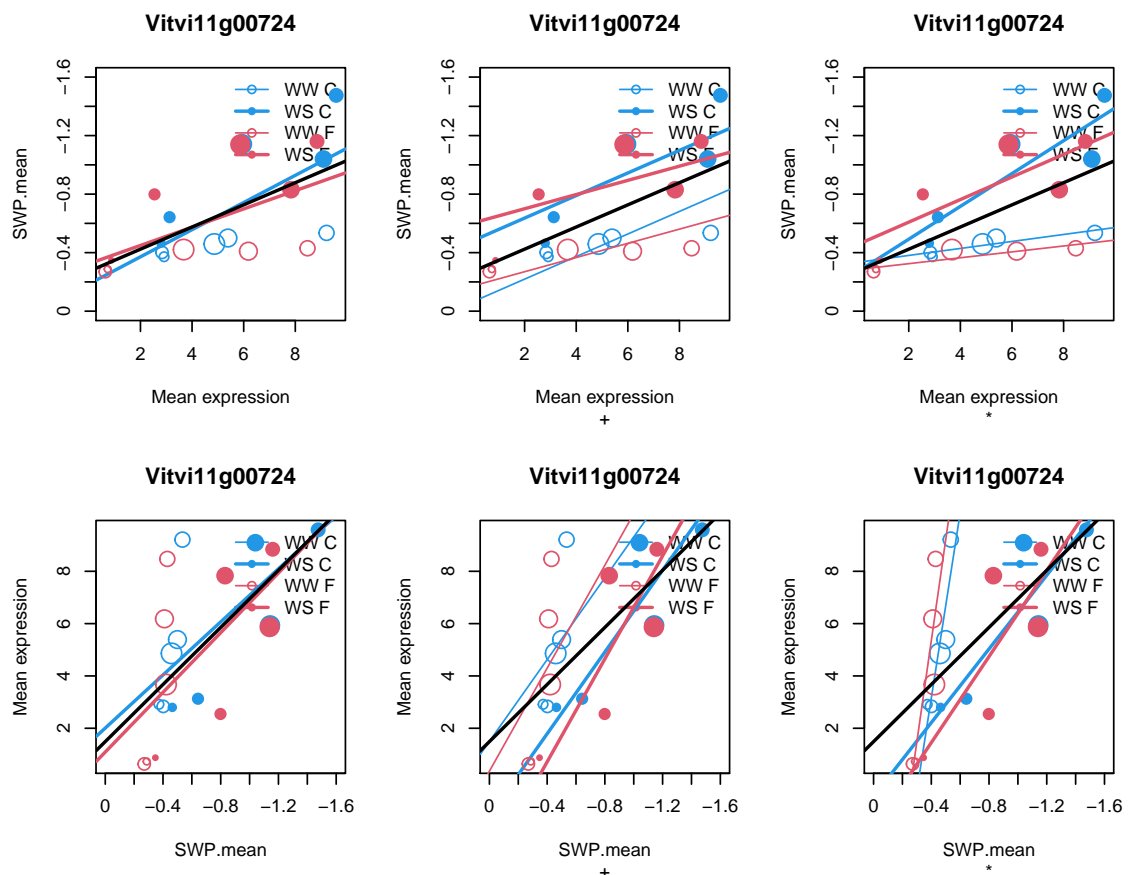
heat shock protein 70B |

Chr1:5502386-5504326 REVERSE LENGTH=646 |

201606

Coefficient swp for Vitvi11g00724.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi11g00724	45.13777	5.070678	17.74622	1.005426e-29	3.831175e-26
B					
Vitvi11g00724	52.16383				



Correlation of all measurements (not just means):  $r = 0.8918$ ,  $r^2 = 0.7953$   
 Correlation of mean values (plots):  $r_2 = 0.6429$

## 5.2.5 Vitvi01g01943

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

```
Vitvi01g01943
```

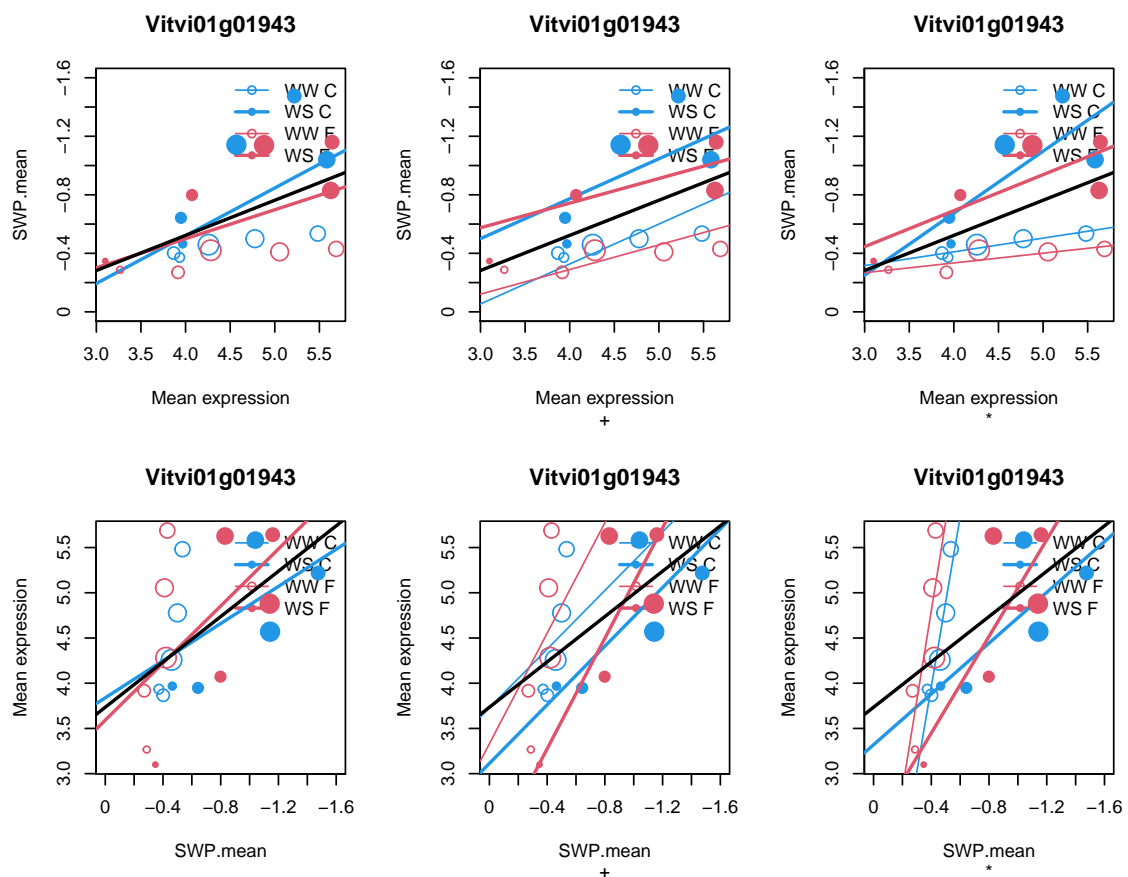
```
20
```

```
stress
```

```
NA
```

Coefficient swp for Vitvi01g01943.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi01g01943	12.14569	4.5568	17.51369	2.361916e-29	6.767539e-26
			B		
Vitvi01g01943	51.46908				



Correlation of all measurements (not just means):  $r = 0.8956$ ,  $r^2 = 0.8021$

Correlation of mean values (plots):  $r_2 = 0.5485$



## 5.2.6 Vitvi08g00689

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi08g00689

20.02.2001

stress.abiotic.heat

DNAJ heat shock N-terminal domain-containing protein |

Chr3:2737589-2740265 FORWARD LENGTH=572 |

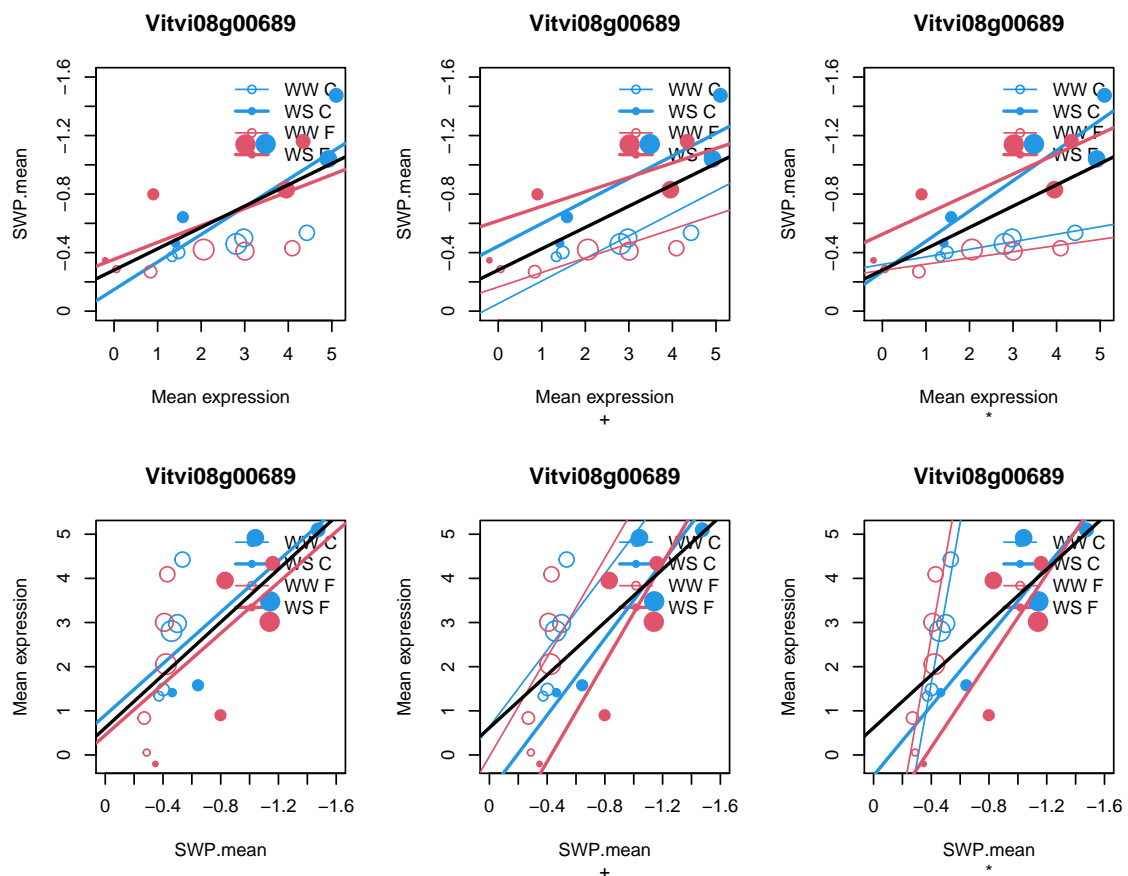
201606

Coefficient swp for Vitvi08g00689.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi08g00689	24.00508	2.578551	17.48107	2.664036e-29	6.767539e-26

B

Vitvi08g00689	51.37095
---------------	----------



Correlation of all measurements (not just means):  $r = 0.8902$ ,  $r^2 = 0.7925$

Correlation of mean values (plots):  $r_2 = 0.6601$

## 5.2.7 Vitvi18g01966

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi18g01966

35.1

not assigned.no ontology

T-complex protein 11 |

Chr1:8118049-8121854 FORWARD LENGTH=1020 |

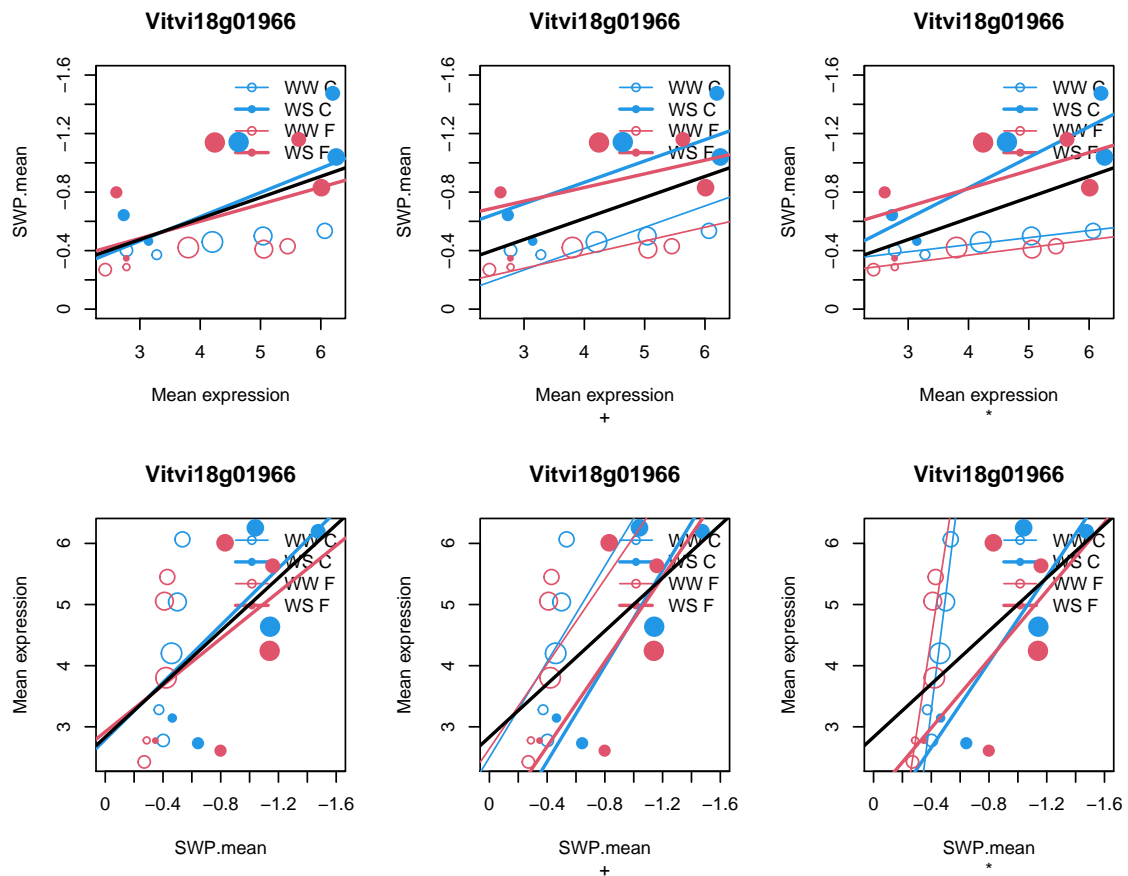
201606

Coefficient swp for Vitvi18g01966.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi18g01966	20.56758	4.255793	17.39113	3.714865e-29	8.088852e-26

B

Vitvi18g01966	51.09958
---------------	----------



Correlation of all measurements (not just means):  $r = 0.8899$ ,  $r^2 = 0.7919$

Correlation of mean values (plots):  $r_2 = 0.5587$

## 5.2.8 Vitvi16g01290

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi16g01290

20.02.2001

stress.abiotic.heat

HSP20-like chaperones superfamily protein |

Chr5:21661588-21663383 FORWARD LENGTH=304 |

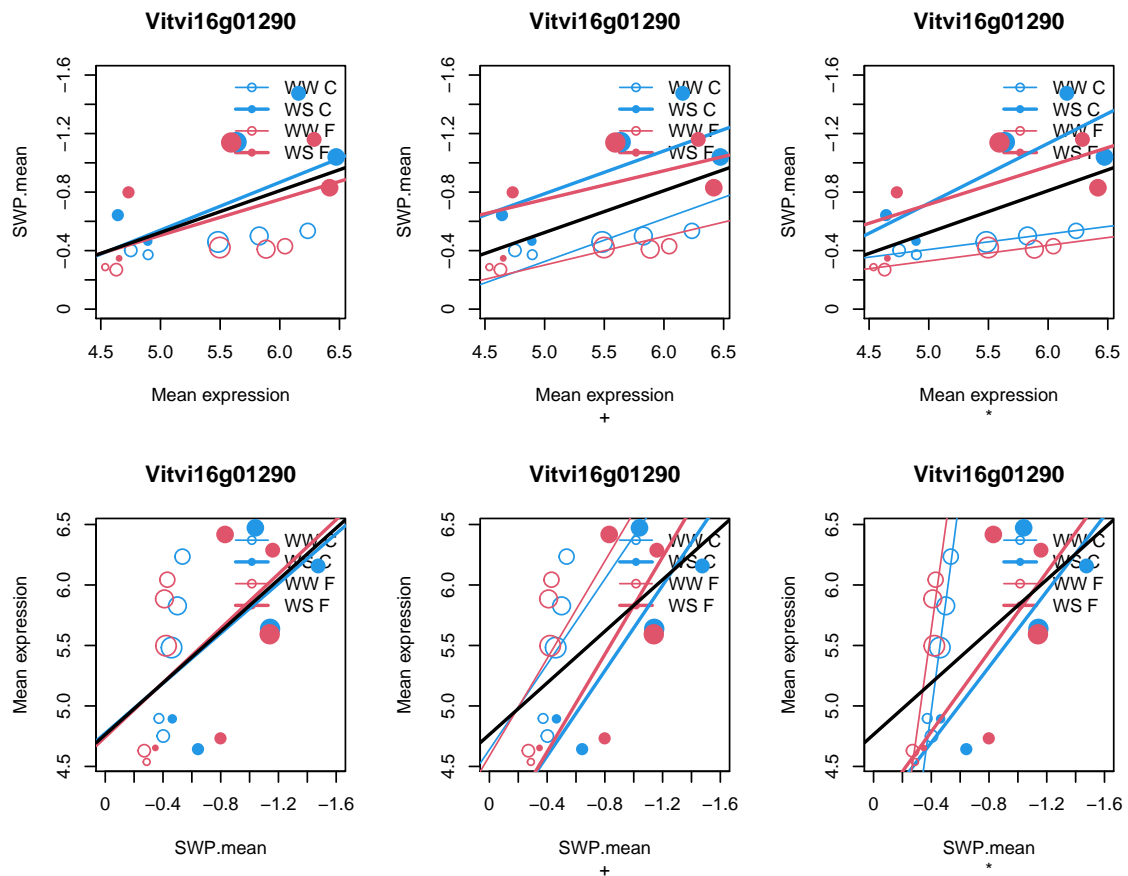
201606

Coefficient swp for Vitvi16g01290.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi16g01290	10.34352	5.463575	17.27953	5.619824e-29	1.070717e-25

B

Vitvi16g01290 50.76115



Correlation of all measurements (not just means):  $r = 0.8955$ ,  $r^2 = 0.8020$

Correlation of mean values (plots):  $r_2 = 0.5522$

## 5.2.9 Vitvi17g00695

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi17g00695

20.02.2001

stress.abiotic.heat

heat shock protein 101 |

Chr1:27936715-27939862 REVERSE LENGTH=911 |

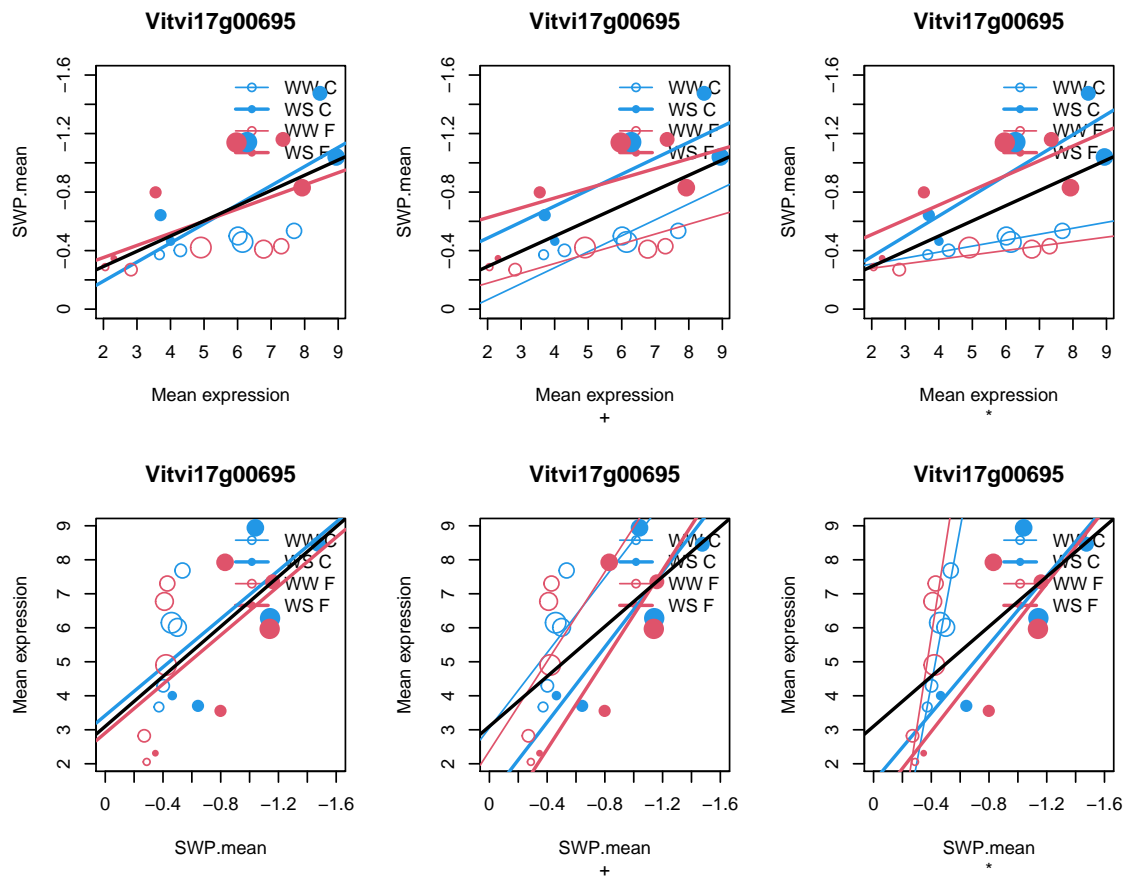
201606

Coefficient swp for Vitvi17g00695.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi17g00695	31.26828	5.508229	17.18961	7.853662e-29	1.330061e-25

B

Vitvi17g00695	50.48708
---------------	----------



Correlation of all measurements (not just means):  $r = 0.8863$ ,  $r^2 = 0.7856$

Correlation of mean values (plots):  $r_2 = 0.6175$

## 5.2.10 Vitvi11g01041

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi11g01041

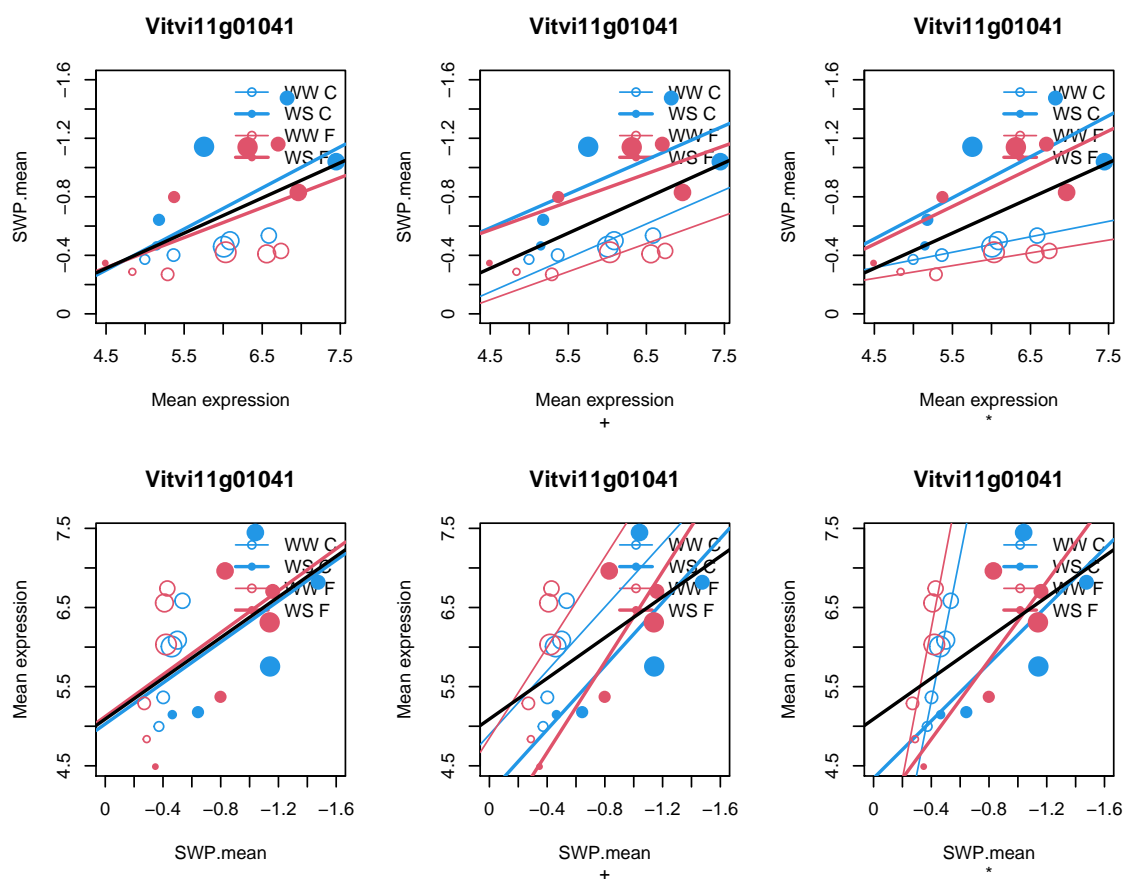
35.2

not assigned.unknown

Genomic DNA chromosome 5 P1 clone MVP2

Coefficient swp for Vitvi11g01041.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi11g01041	12.15265	5.934203	17.0483	1.331594e-28	2.017785e-25
					B
Vitvi11g01041	50.05386				



Correlation of all measurements (not just means):  $r = 0.8903$ ,  $r^2 = 0.7927$

Correlation of mean values (plots):  $r_2 = 0.5562$

### 5.2.11 Vitvi08g01757

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

```
Vitvi08g01757
```

```
35.1
```

```
not assigned.no ontology
```

```
Ahal domain-containing protein |
```

```
Chr3:3839289-3841303 FORWARD LENGTH=360 |
```

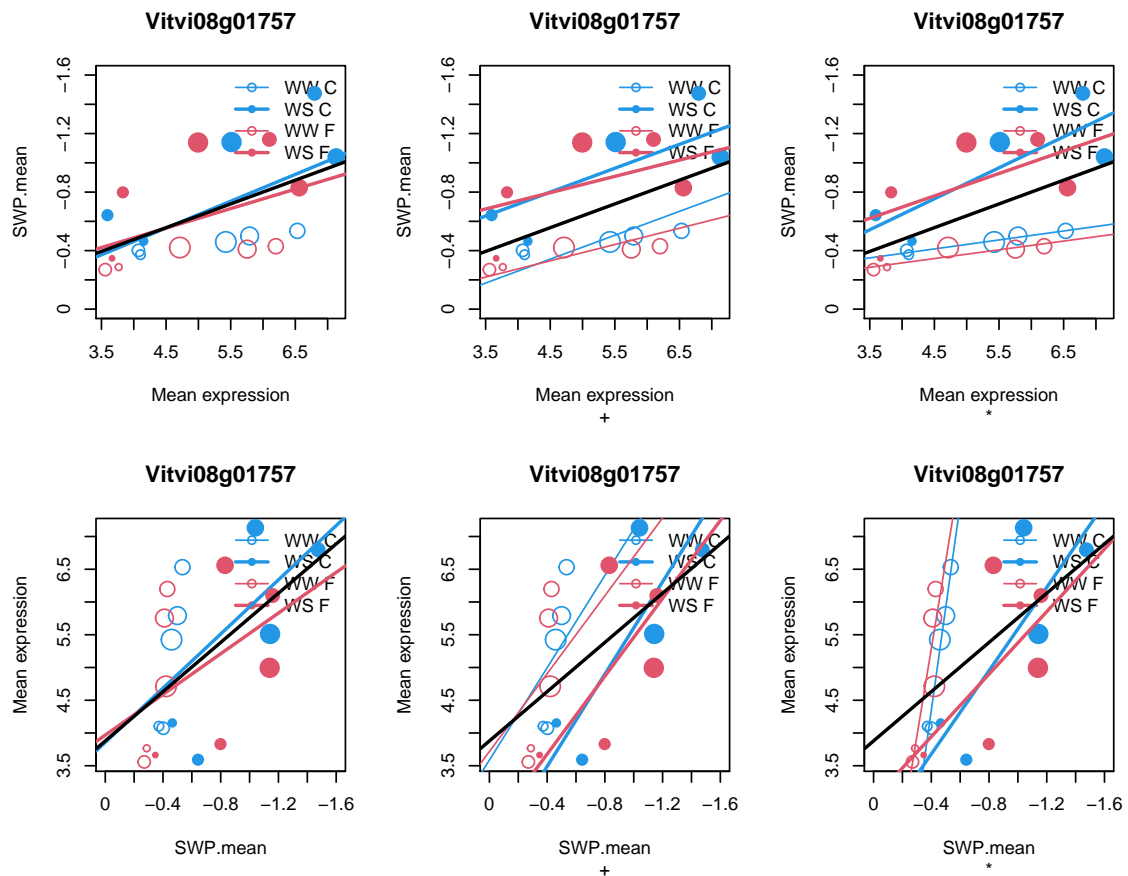
```
201606
```

Coefficient swp for Vitvi08g01757.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi08g01757	17.90111	5.112598	17.02443	1.456216e-28	2.017785e-25

B

Vitvi08g01757	49.98036
---------------	----------



Correlation of all measurements (not just means):  $r = 0.8865$ ,  $r^2 = 0.7859$

Correlation of mean values (plots):  $r_2 = 0.5523$

## 5.2.12 Vitvi01g01512

```
> cat.feature(fdata[fdata$geneID %in% varname, 2:5])
```

Vitvi01g01512

20.02.2001

stress.abiotic.heat

heat shock protein 21 |

Chr4:13819048-13819895 REVERSE LENGTH=227 |

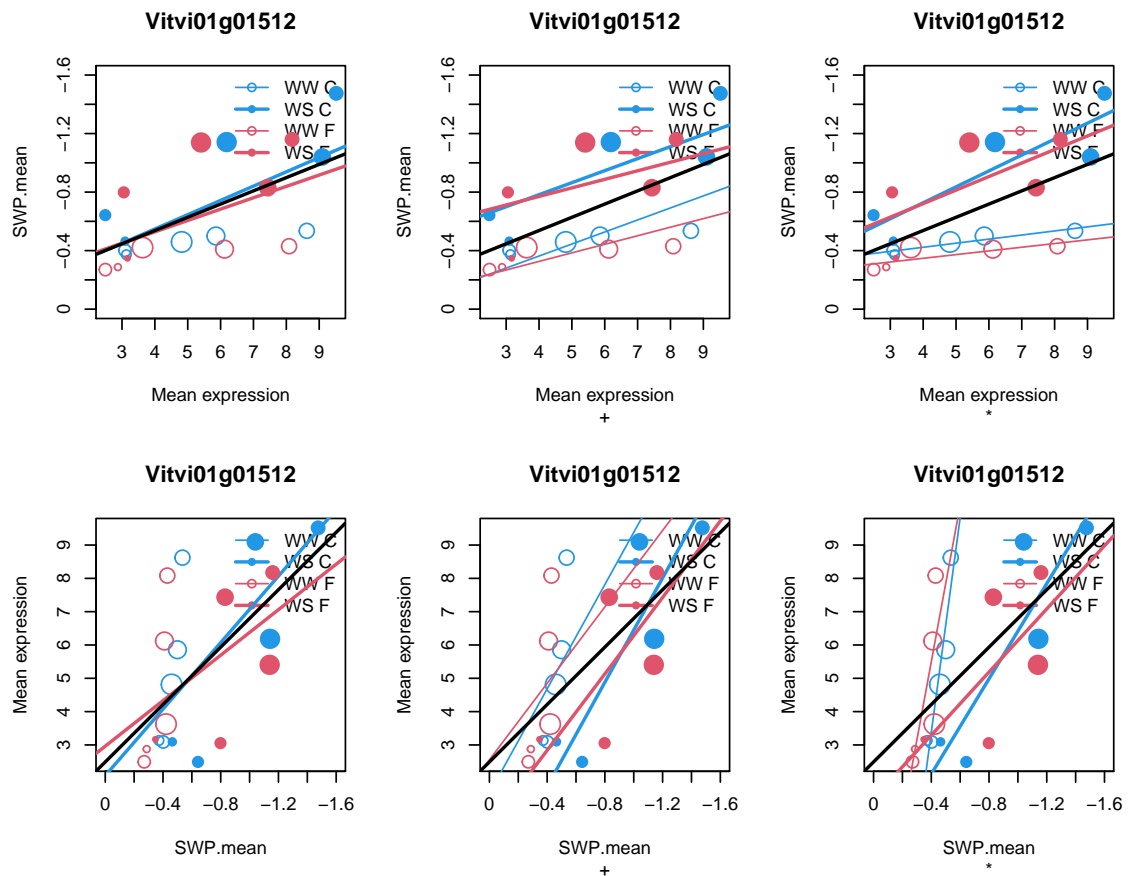
201606

Coefficient swp for Vitvi01g01512.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi01g01512	36.17683	5.31657	16.82069	3.133275e-28	3.979781e-25

B

Vitvi01g01512 49.3496



Correlation of all measurements (not just means):  $r = 0.8819$ ,  $r^2 = 0.7777$

Correlation of mean values (plots):  $r_2 = 0.6241$

### 5.2.13 Vitvi02g00991

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

```
Vitvi02g00991
```

```
35.2
```

```
not assigned.unknown
```

```
UDP-Glycosyltransferase superfamily protein |
```

```
Chr2:15417618-15419108 REVERSE LENGTH=496 |
```

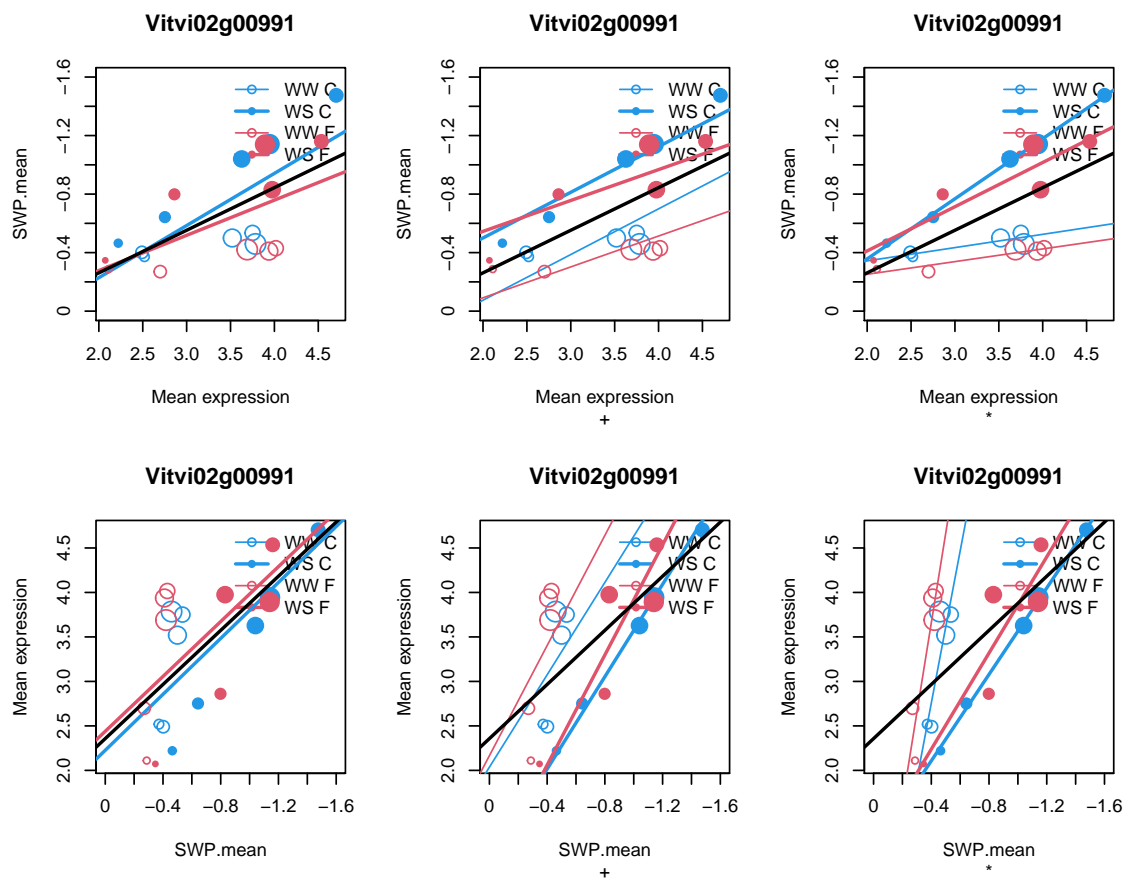
```
201606
```

Coefficient swp for Vitvi02g00991.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi02g00991	12.05033	3.35527	16.64172	6.168548e-28	7.232385e-25

B

```
Vitvi02g00991 48.79022
```



Correlation of all measurements (not just means):  $r = 0.8856$ ,  $r^2 = 0.7843$

Correlation of mean values (plots):  $r_2 = 0.6655$



## 5.2.14 Vitvi08g00645

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi08g00645

29.03.1999

protein.targeting.unknown

Haloacid dehalogenase-like hydrolase (HAD) superfamily protein |

Chr3:20760797-20762892 REVERSE LENGTH=305 |

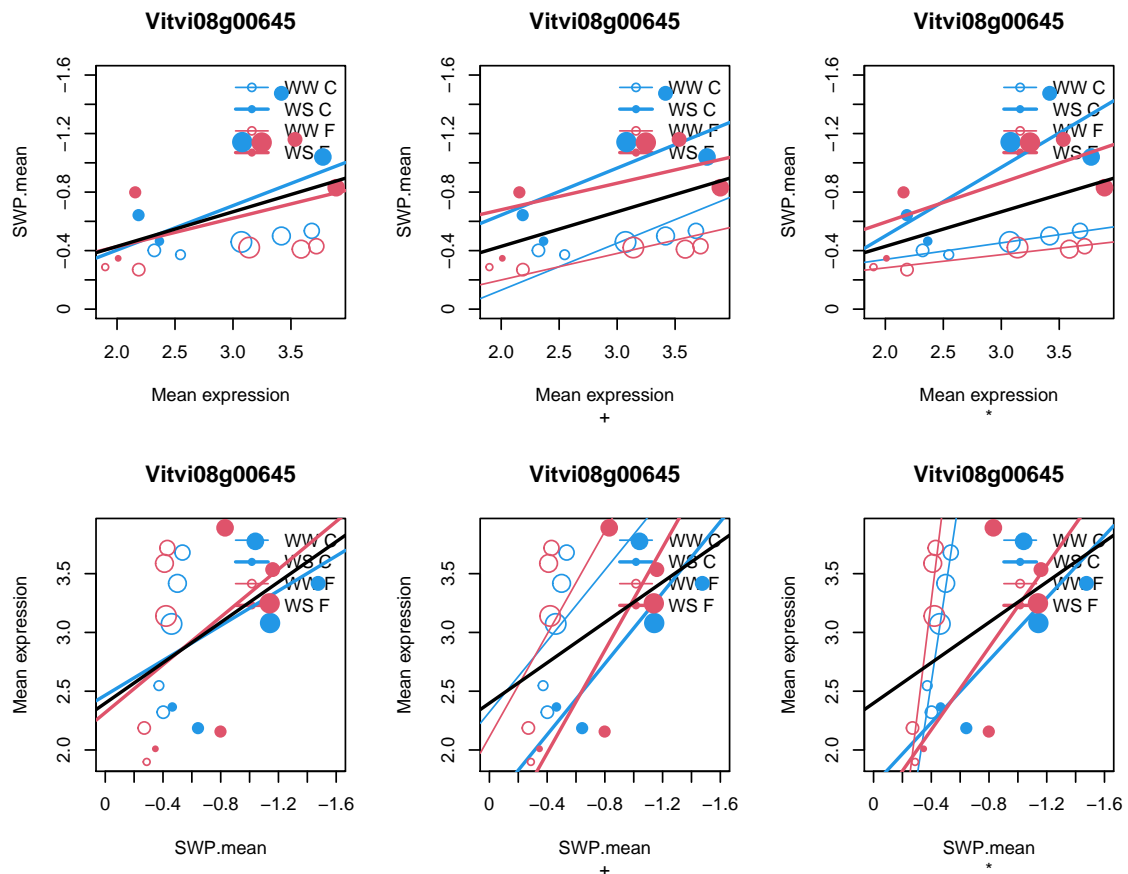
201606

Coefficient swp for Vitvi08g00645.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi08g00645	10.12433	2.961665	16.61784	6.754009e-28	7.353186e-25

B

Vitvi08g00645 48.71522



Correlation of all measurements (not just means):  $r = 0.8881$ ,  $r^2 = 0.7887$

Correlation of mean values (plots):  $r_2 = 0.4506$

## 5.2.15 Vitvi18g02066

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi18g02066

20.02.2001

stress.abiotic.heat

chloroplast heat shock protein 70-1 |

Chr4:12590094-12593437 FORWARD LENGTH=718 |

201606

Vitvi18g02066

29.6

protein.folding

chloroplast heat shock protein 70-1 |

Chr4:12590094-12593437 FORWARD LENGTH=718 |

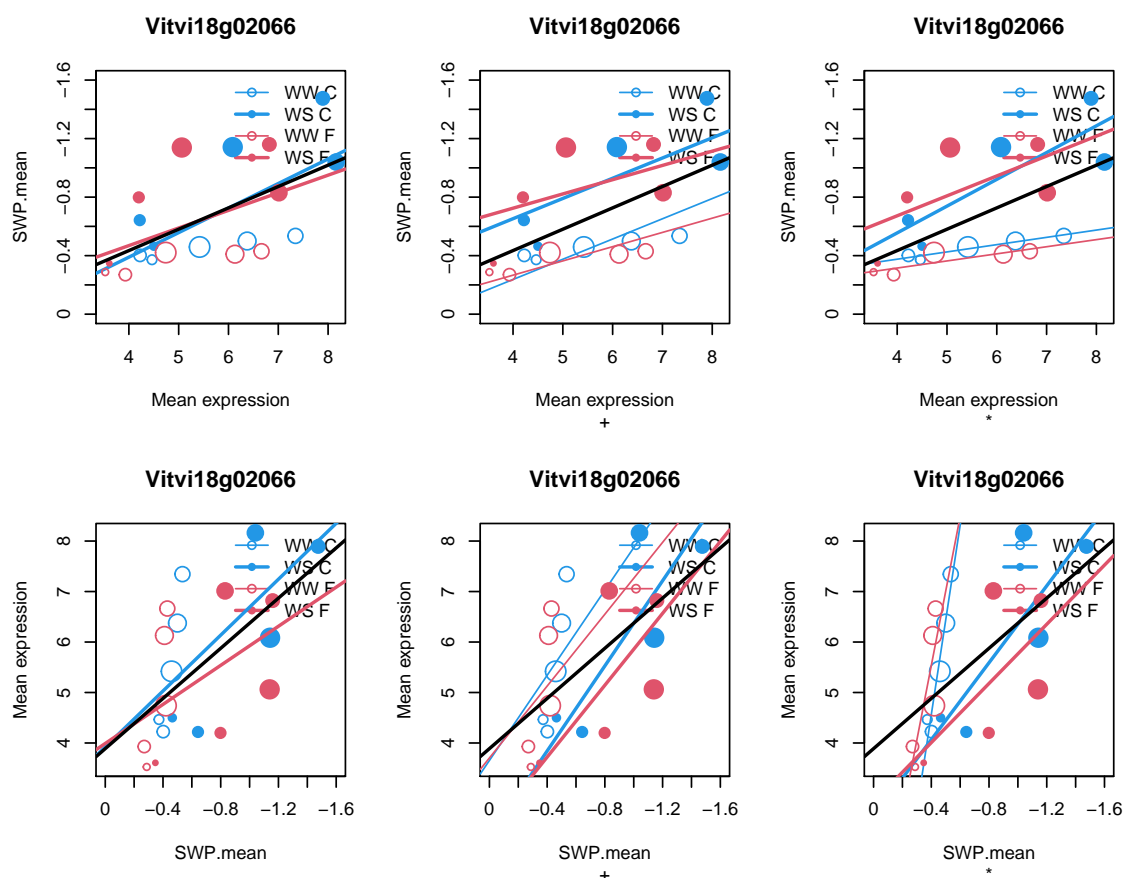
201606

Coefficient swp for Vitvi18g02066.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi18g02066	21.43313	5.518923	16.41996	1.435959e-27	1.459126e-24

B

Vitvi18g02066	48.09019
---------------	----------



Correlation of all measurements (not just means):  $r = 0.8784$ ,  $r^2 = 0.7715$

Correlation of mean values (plots):  $r_2 = 0.6021$

## 5.2.16 Vitvi01g01459

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi01g01459

17.01.2003

hormone metabolism.abscisic acid.induced-regulated-responsive-activate

GRAM domain family protein |

Chr5:4207081-4207812 FORWARD LENGTH=213 |

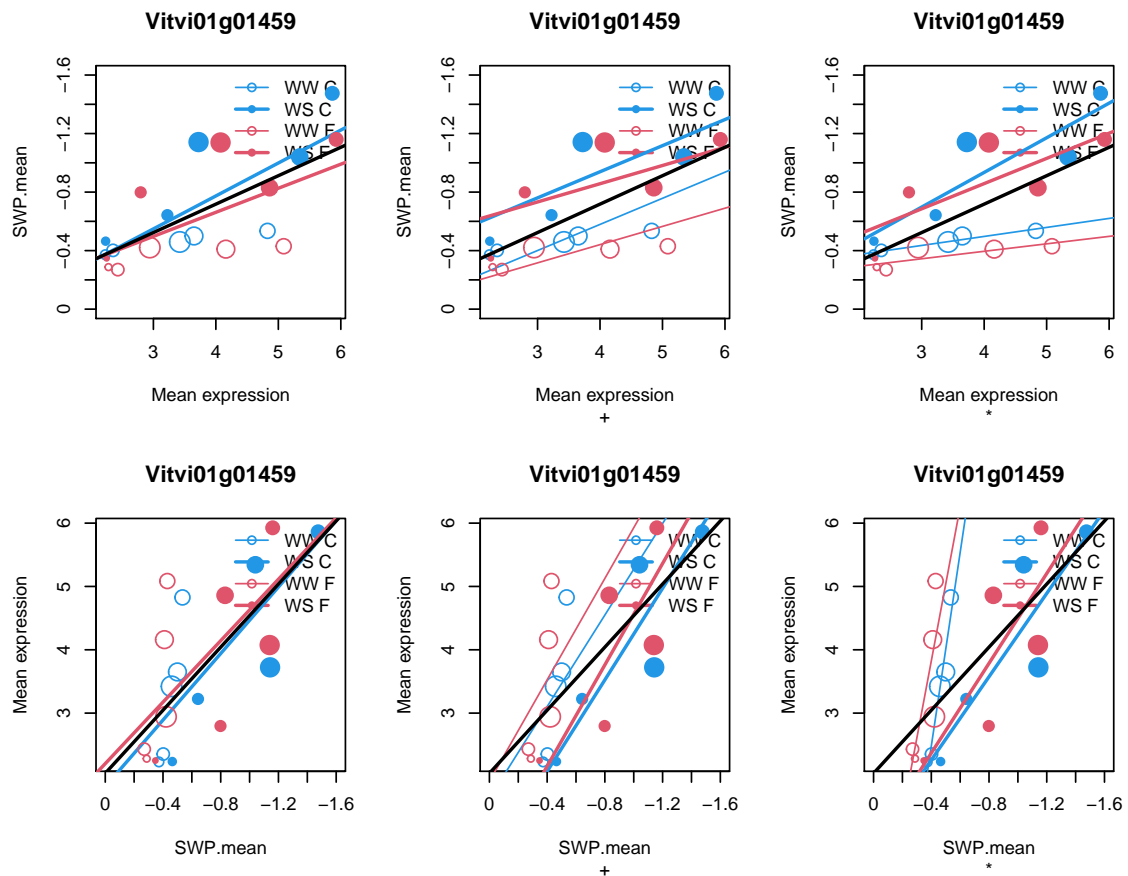
201606

Coefficient swp for Vitvi01g01459.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi01g01459	18.91439	3.683872	16.32173	2.091898e-27	1.992795e-24

B

Vitvi01g01459 47.77767



Correlation of all measurements (not just means):  $r = 0.8777, r^2 = 0.7704$

Correlation of mean values (plots):  $r_2 = 0.6956$

## 5.2.17 Vitvi19g01760

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

```
Vitvi19g01760
```

```
35.2
```

```
not assigned.unknown
```

```
HSP20-like chaperones superfamily protein |
```

```
Chr1:19980510-19980983 FORWARD LENGTH=157 |
```

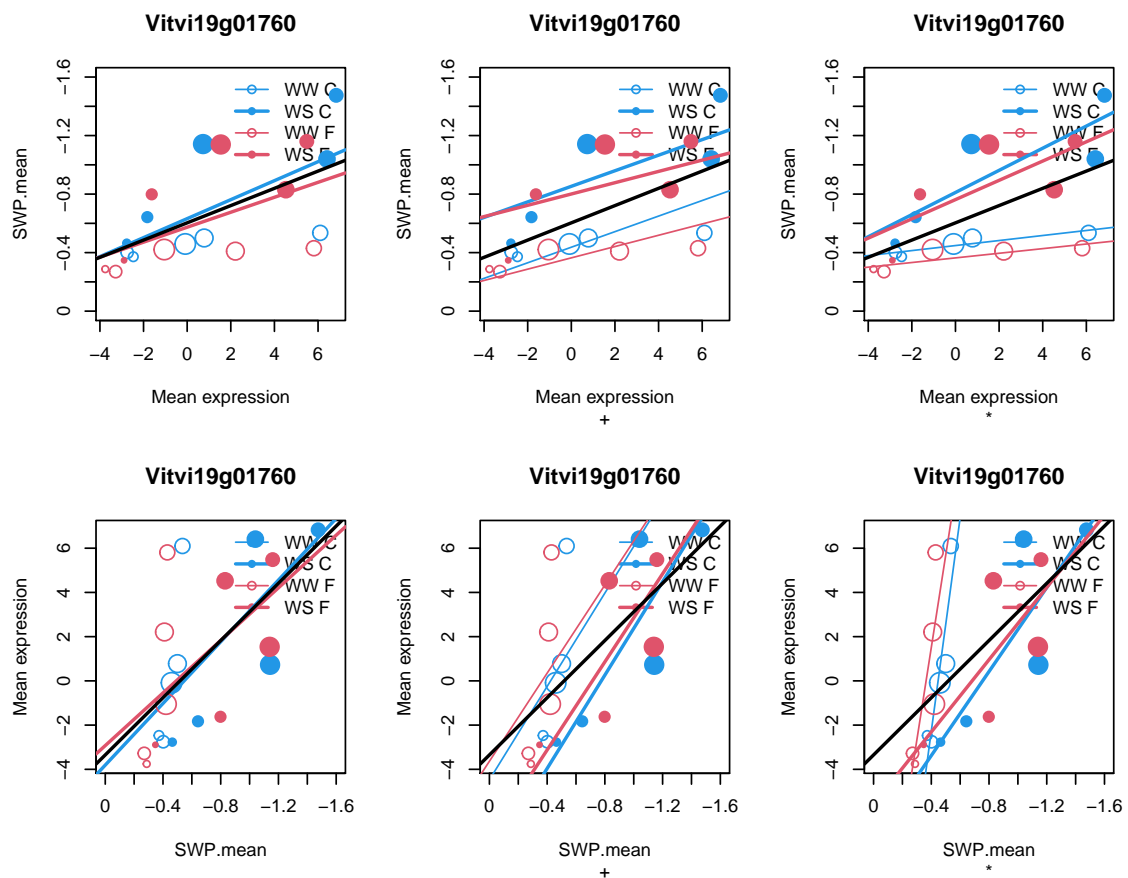
```
201606
```

Coefficient swp for Vitvi19g01760.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi19g01760	55.69011	0.8952506	16.20667	3.255542e-27	2.918881e-24

B

```
Vitvi19g01760 47.40966
```



Correlation of all measurements (not just means):  $r = 0.8739$ ,  $r^2 = 0.7638$

Correlation of mean values (plots):  $r_2 = 0.6174$

## 5.2.18 Vitvi14g02026

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi14g02026

20.02.2001

stress.abiotic.heat

casein lytic proteinase B3 |

Chr5:5014399-5018255 REVERSE LENGTH=968 |

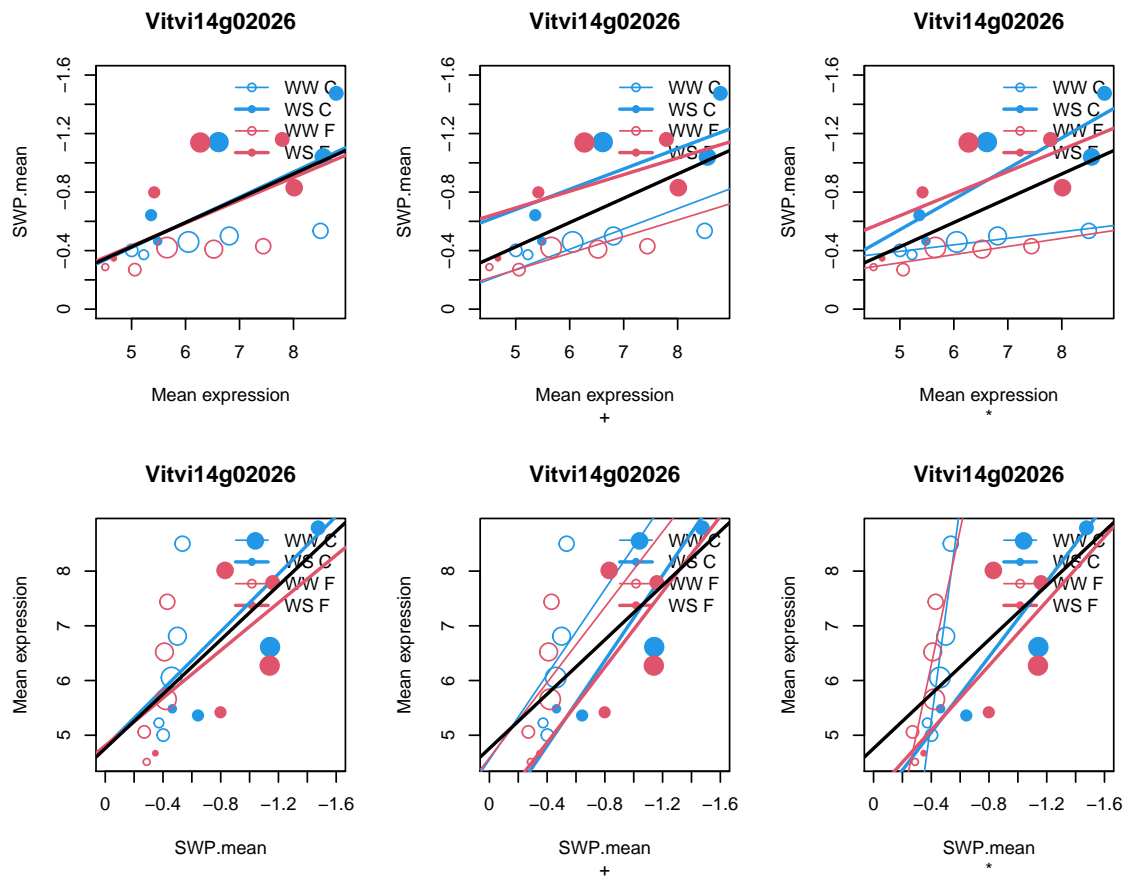
201606

Coefficient swp for Vitvi14g02026.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi14g02026	20.05851	6.387659	16.17746	3.643339e-27	3.044873e-24

B

Vitvi14g02026 47.31591



Correlation of all measurements (not just means):  $r = 0.8756$ ,  $r^2 = 0.7666$

Correlation of mean values (plots):  $r_2 = 0.6419$

## 5.2.19 Vitvi05g00218

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi05g00218

34.9

transport.metabolite transporters at the mitochondrial membrane

Mitochondrial substrate carrier family protein |

Chr5:5087822-5089677 FORWARD LENGTH=316 |

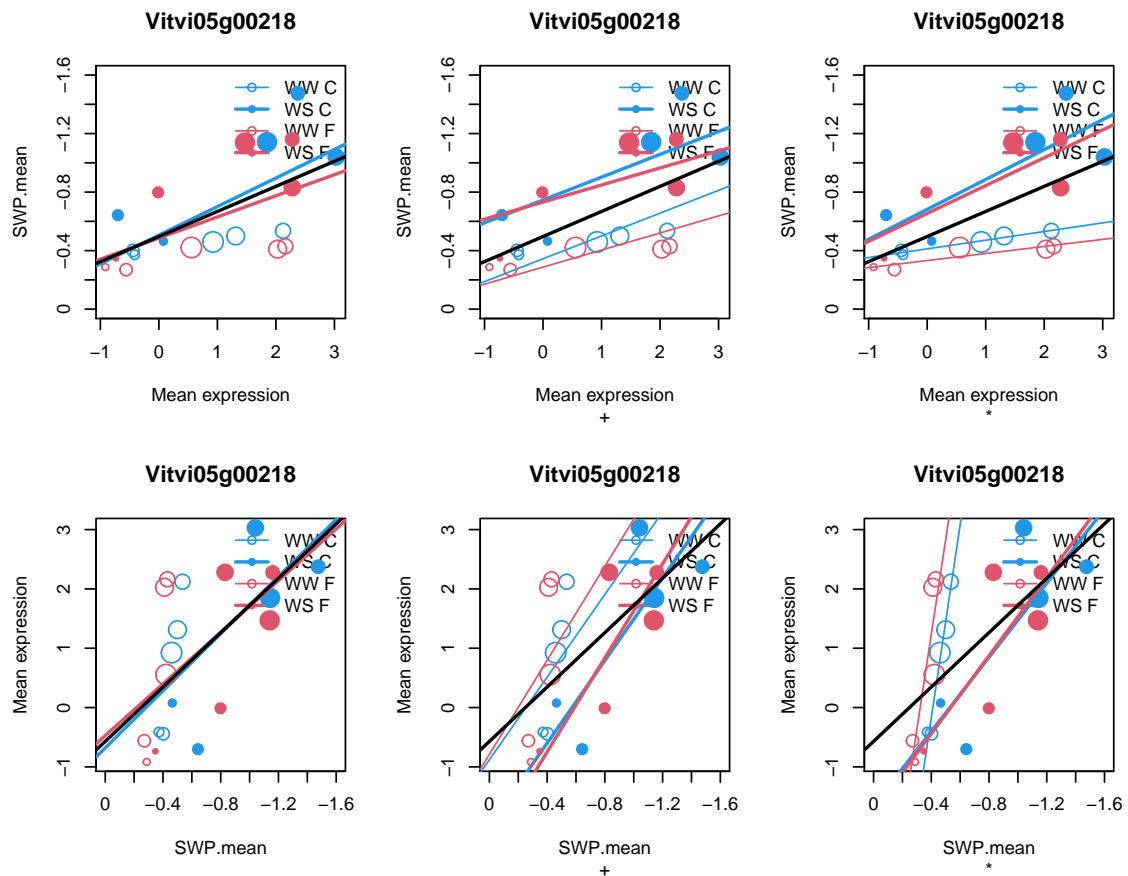
201606

Coefficient swp for Vitvi05g00218.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi05g00218	19.21392	0.933593	16.16684	3.795603e-27	3.044873e-24

B

Vitvi05g00218 47.2818



Correlation of all measurements (not just means):  $r = 0.8756, r^2 = 0.7668$

Correlation of mean values (plots):  $r_2 = 0.6269$

## 5.2.20 Vitvi02g01827

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi02g01827

31.05.2002

cell.cell death.apoptosis

BAX inhibitor 1 |

Chr5:19136071-19137585 FORWARD LENGTH=247 |

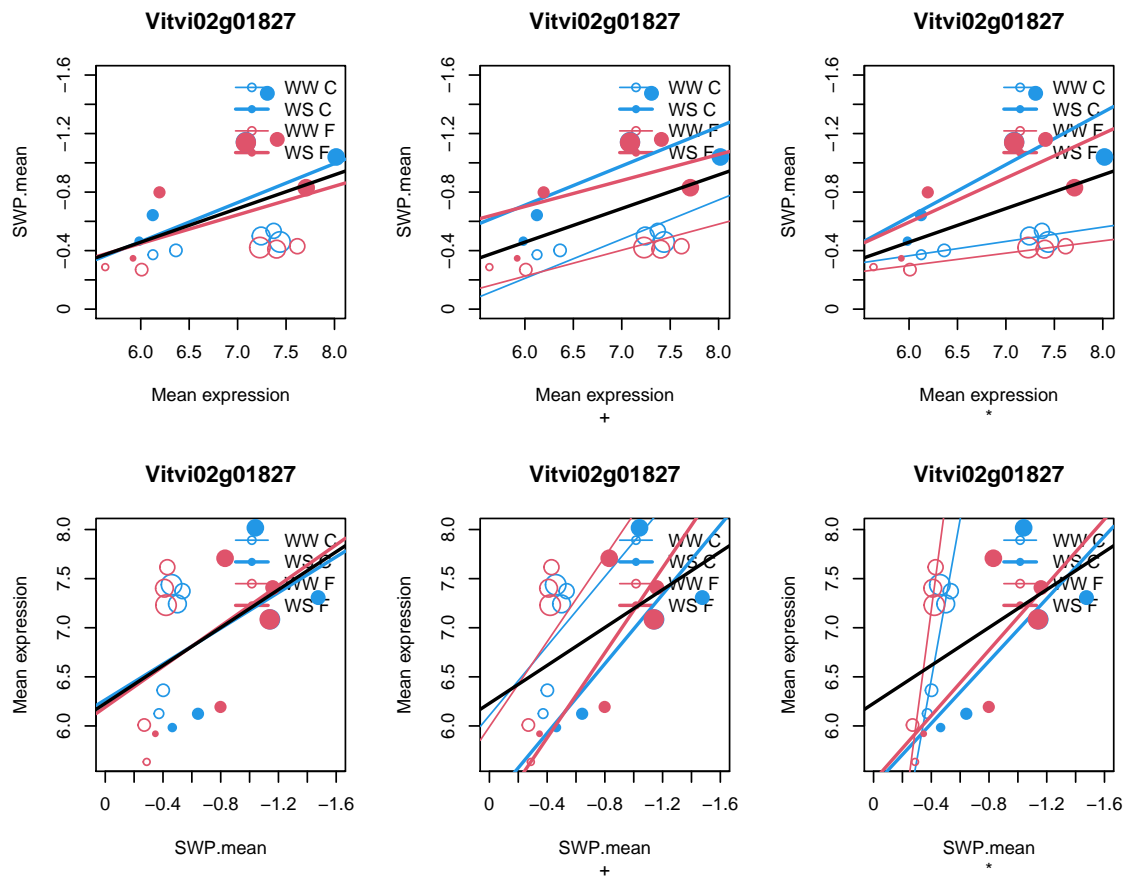
201606

Coefficient swp for Vitvi02g01827.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi02g01827	10.64761	6.863265	16.12778	4.413194e-27	3.363295e-24

B

Vitvi02g01827	47.15613
---------------	----------



Correlation of all measurements (not just means):  $r = 0.8809$ ,  $r^2 = 0.7760$

Correlation of mean values (plots):  $r_2 = 0.4711$

### 5.3 Bottom 20 genes

Plots of situation for top 20 genes

```
> out <- ""
> (cname <- colnames(coefs)[2])
[1] "swp"
> varnames <- rownames(tail(tt, topn))
> varname <- varnames[1]
> for(varname in varnames)
+ out <- paste(out, knit_child(file.path("../doc/", "40aa_one-gene-plots.Rnw")),
```



### 5.3.1 Vitvi13g00182

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi13g00182

30.2.7

signalling.receptor.kinases.leucine rich repeat VII

Leucine-rich repeat protein kinase family protein |

Chr3:20899403-20902390 REVERSE LENGTH=964 |

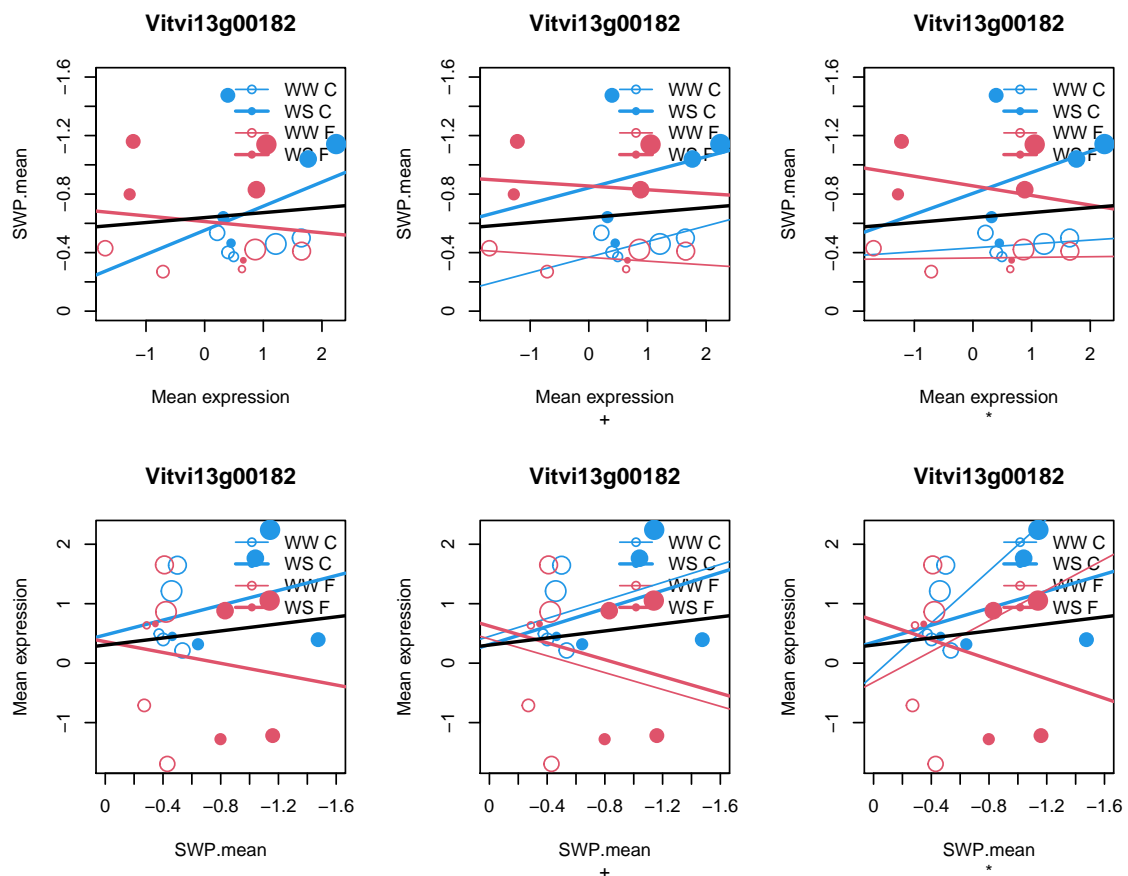
201606

Coefficient swp for Vitvi13g00182.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi13g00182	0.01401526	0.4989287	0.006841771	0.9945578	0.9957991

B

Vitvi13g00182 -6.117765



Correlation of all measurements (not just means):  $r = 8e - 04, r^2 = 6e - 07$

Correlation of mean values (plots):  $r_2 = 0.1003$

### 5.3.2 Vitvi07g02020

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi07g02020

11.9.4.5

lipid metabolism.lipid degradation.beta-oxidation.acyl-CoA thioesterase

Thioesterase superfamily protein |

Chr1:1147888-1148352 REVERSE LENGTH=127 |

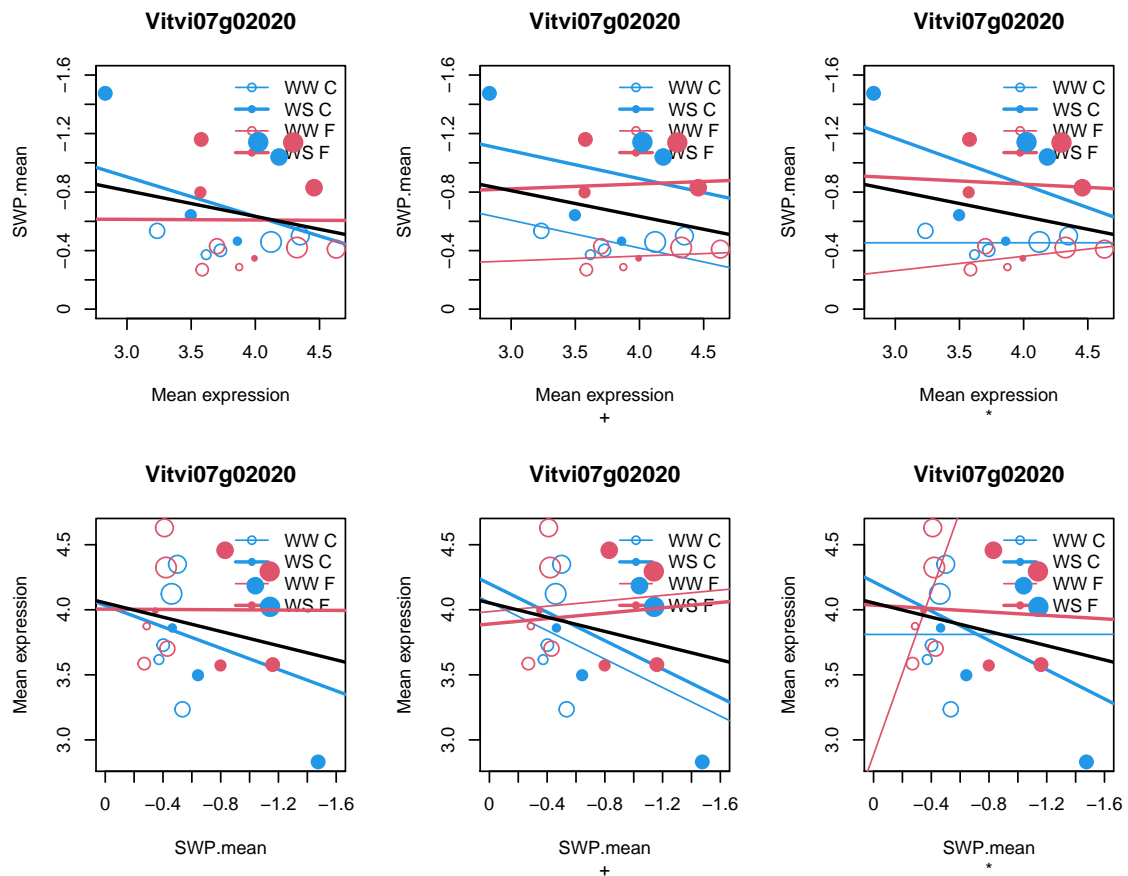
201606

Coefficient swp for Vitvi07g02020.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi07g02020	0.005691716	3.873045	0.006632351	0.9947244	0.9959005

B

Vitvi07g02020 -6.117766



Correlation of all measurements (not just means):  $r = 8e - 04, r^2 = 6e - 07$

Correlation of mean values (plots):  $r_2 = -0.2195$

### 5.3.3 Vitvi17g00137

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi17g00137

23.04.2001

nucleotide metabolism.phosphotransfer and pyrophosphatases.adenylate k

P-loop containing nucleoside triphosphate hydrolases superfamily prote

Chr3:293981-295357 REVERSE LENGTH=263 |

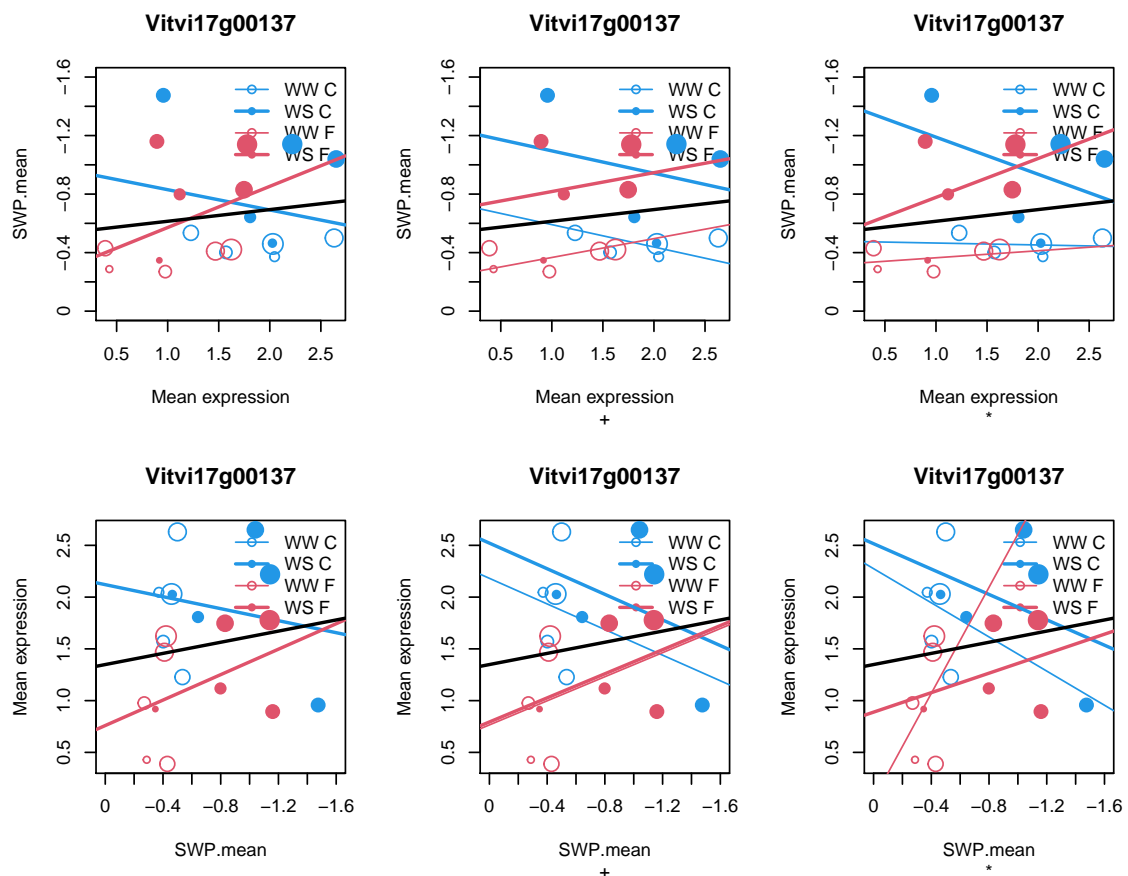
201606

Coefficient swp for Vitvi17g00137.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi17g00137	0.008198947	1.525163	0.006493677	0.9948347	0.9959455

B

Vitvi17g00137 -6.117767



Correlation of all measurements (not just means):  $r = 7e - 04, r^2 = 5e - 07$

Correlation of mean values (plots):  $r_2 = 0.1466$

### 5.3.4 Vitvi16g00501

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi16g00501

30.2.11

signalling.receptor.kinases.leucine rich repeat XI

Leucine-rich repeat protein kinase family protein |

Chr5:24724541-24727842 REVERSE LENGTH=1041 |

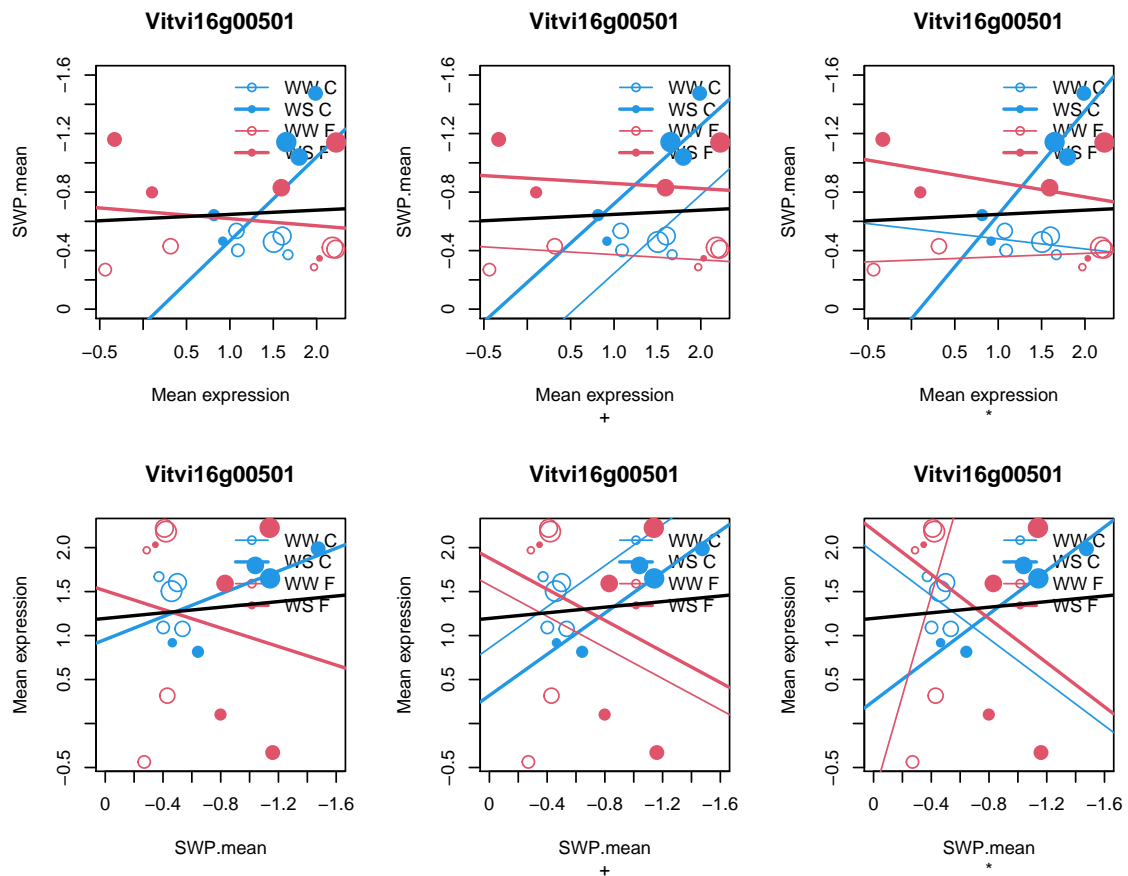
201606

Coefficient swp for Vitvi16g00501.

```
logFC AveExpr t P.Value adj.P.Val
Vitvi16g00501 0.01244711 1.299418 0.006334405 0.9949614 0.9960069
```

B

```
Vitvi16g00501 -6.117768
```



Correlation of all measurements (not just means):  $r = 7e - 04, r^2 = 5e - 07$

Correlation of mean values (plots):  $r_2 = 0.0679$

### 5.3.5 Vitvi02g00007

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi02g00007

35.2

not assigned.unknown

RNA-binding (RRM/RBD/RNP motifs) family protein |

Chr4:16953404-16955127 REVERSE LENGTH=187 |

201606

Vitvi02g00007

27.4

RNA.RNA binding

RNA-binding (RRM/RBD/RNP motifs) family protein |

Chr4:16953404-16955127 REVERSE LENGTH=187 |

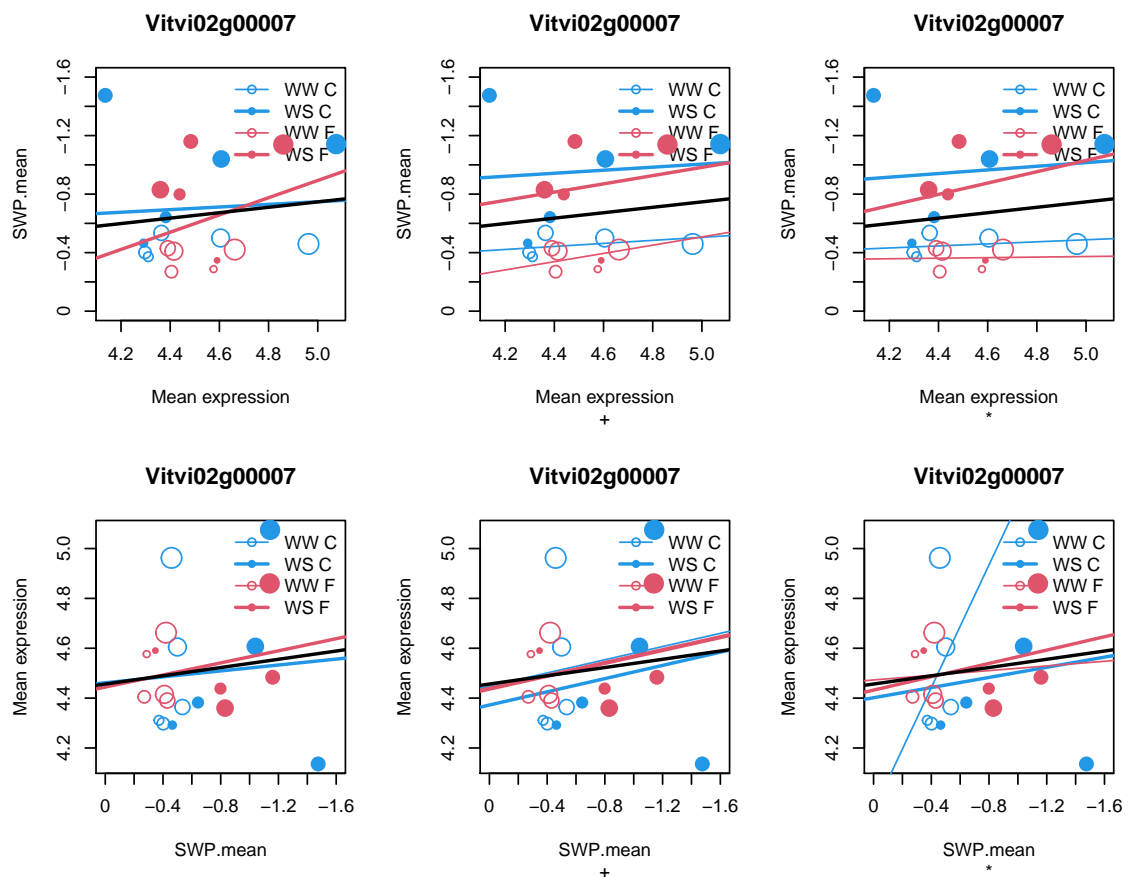
201606

Coefficient swp for Vitvi02g00007.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi02g00007	0.003011828	4.510734	0.006025826	0.9952068	0.9961872

B

Vitvi02g00007 -6.11777



Correlation of all measurements (not just means):  $r = 7e - 04$ ,  $r^2 = 5e - 07$

Correlation of mean values (plots):  $r_2 = 0.1238$

### 5.3.6 Vitvi03g00062

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi03g00062

27.1

RNA.processing

ribosome-binding factor A family protein |

Chr4:16570019-16571274 REVERSE LENGTH=215 |

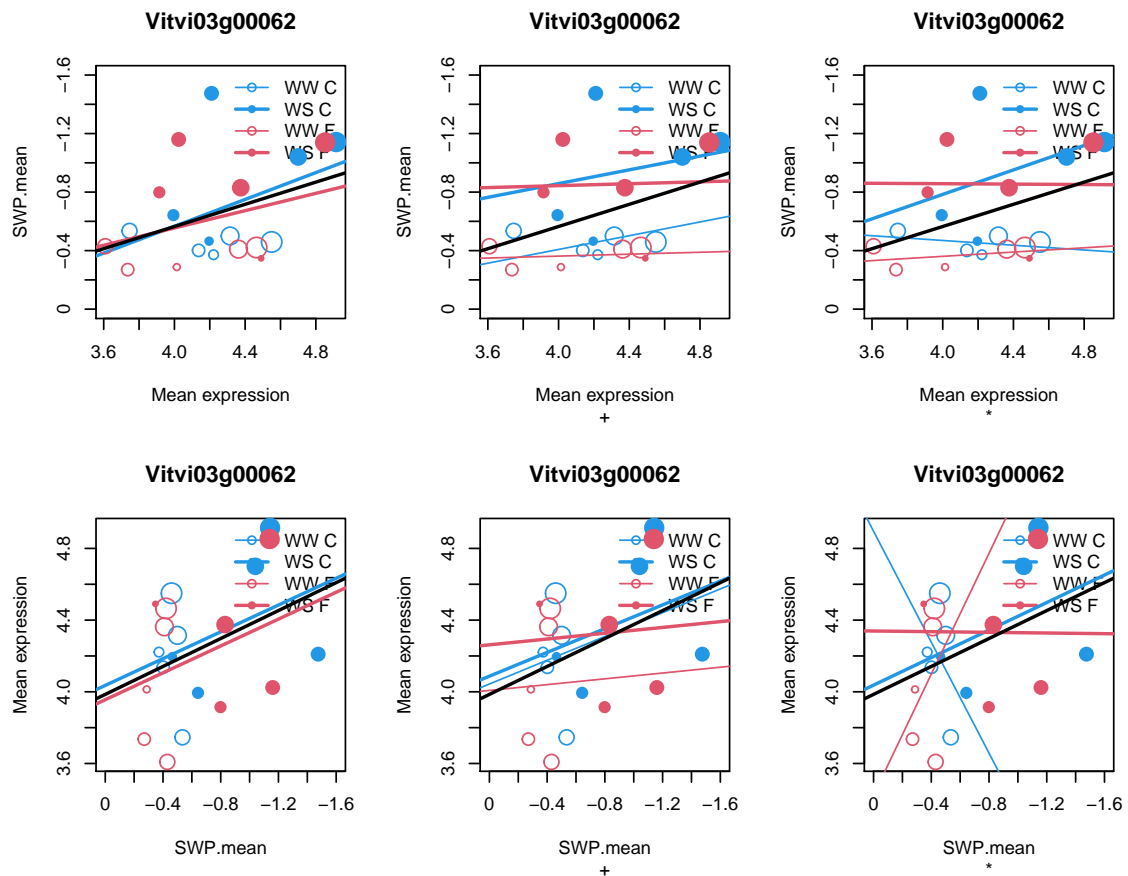
201606

Coefficient swp for Vitvi03g00062.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi03g00062	-0.004168606	4.241324	-0.005574307	0.995566	0.9964813

B

Vitvi03g00062 -6.117772



Correlation of all measurements (not just means):  $r = -6e - 04$ ,  $r^2 = 4e - 07$

Correlation of mean values (plots):  $r_2 = 0.3844$

### 5.3.7 Vitvi17g01515

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi17g01515

35.2

not assigned.unknown

alpha/beta-Hydrolases superfamily protein |

Chr3:17755553-17757692 REVERSE LENGTH=623 |

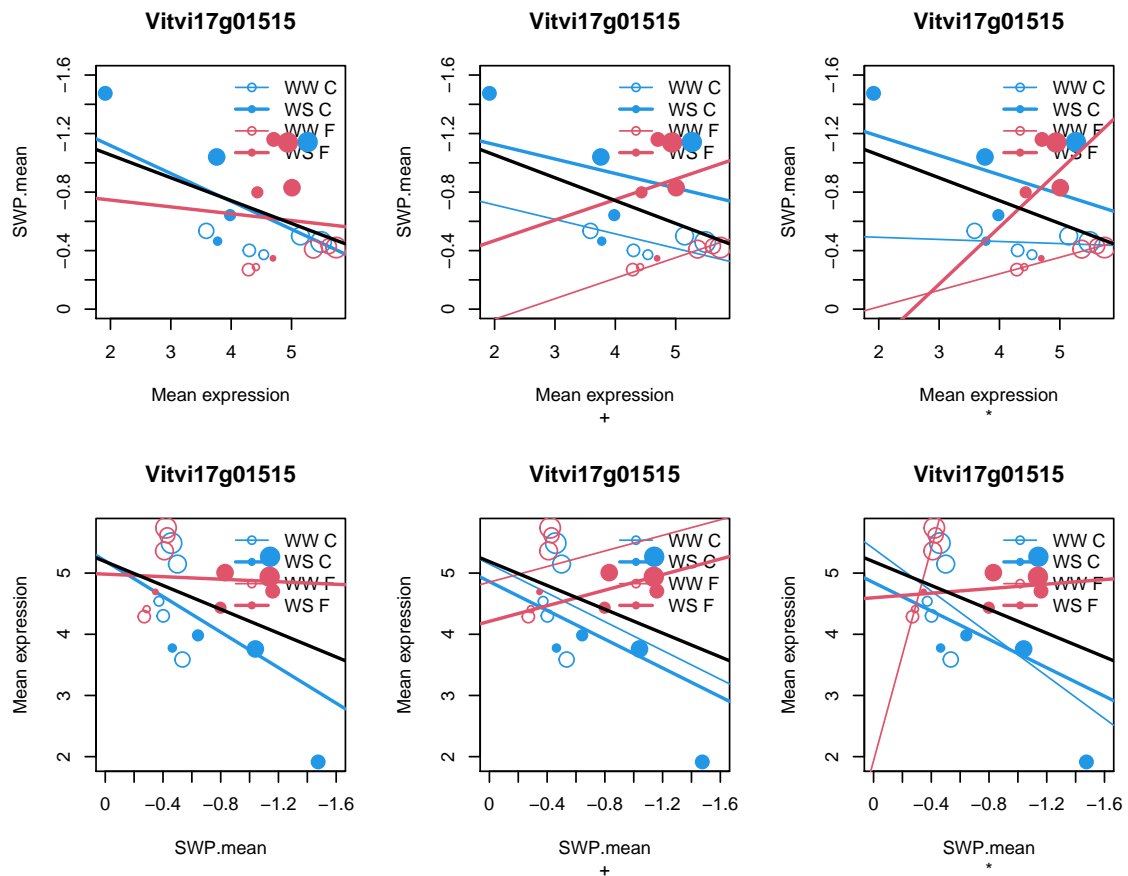
201606

Coefficient swp for Vitvi17g01515.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi17g01515	0.008828815	4.548098	0.005108763	0.9959363	0.9967865

B

Vitvi17g01515 -6.117775



Correlation of all measurements (not just means):  $r = 6e - 04, r^2 = 3e - 07$

Correlation of mean values (plots):  $r_2 = -0.3898$

### 5.3.8 Vitvi18g00098

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi18g00098

27.1

RNA.processing

Small nuclear ribonucleoprotein family protein |

Chr1:7128979-7130371 FORWARD LENGTH=131 |

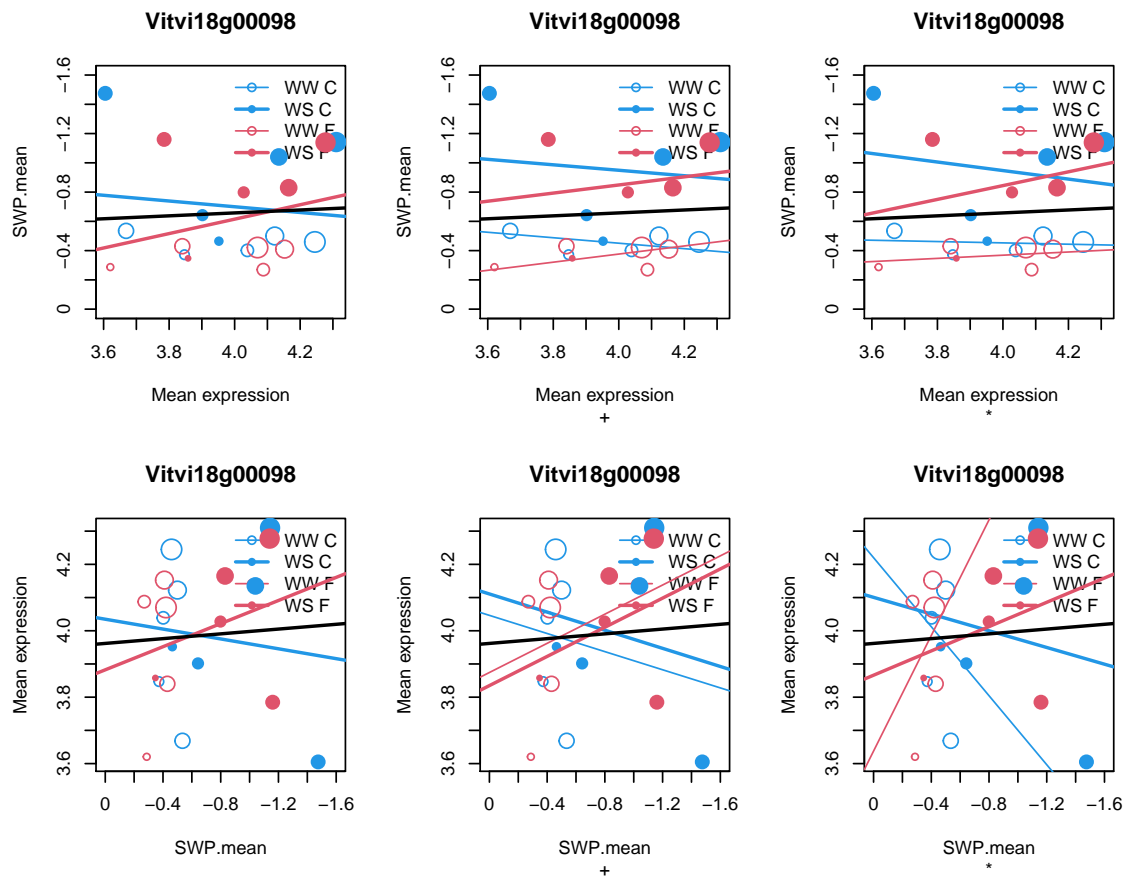
201606

Coefficient swp for Vitvi18g00098.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi18g00098	0.002265333	3.985465	0.004627403	0.9963192	0.9971042

B

Vitvi18g00098 -6.11777



Correlation of all measurements (not just means):  $r = 6e - 04, r^2 = 3e - 07$

Correlation of mean values (plots):  $r_2 = 0.0599$



### 5.3.9 Vitvi06g00091

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

```
Vitvi06g00091
```

```
35.2
```

```
not assigned.unknown
```

```
transmembrane protein%2C putative (Protein of unknown function%2C DUF5
```

```
Chr5:6714533-6715837 REVERSE LENGTH=181 |
```

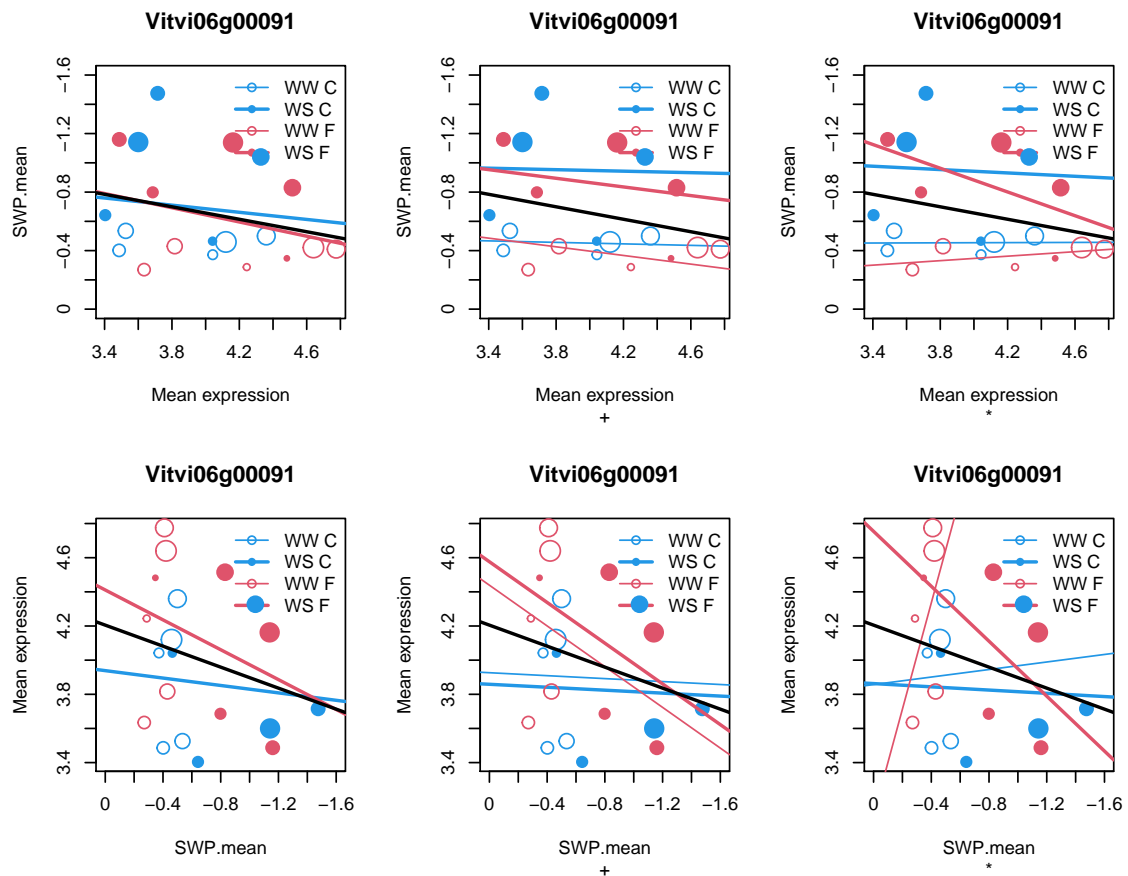
```
201606
```

Coefficient swp for Vitvi06g00091.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi06g00091	0.003007602	4.003266	0.003585786	0.9971477	0.9978679

B

```
Vitvi06g00091 -6.117781
```



Correlation of all measurements (not just means):  $r = 4e - 04, r^2 = 2e - 07$

Correlation of mean values (plots):  $r_2 = -0.2560$

### 5.3.10 Vitvi14g02707

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi14g02707

20.1.2.1

stress.biotic.receptors.CC-NBS-LRR

NB-ARC domain-containing disease resistance protein |

Chr4:13633953-13636712 REVERSE LENGTH=919 |

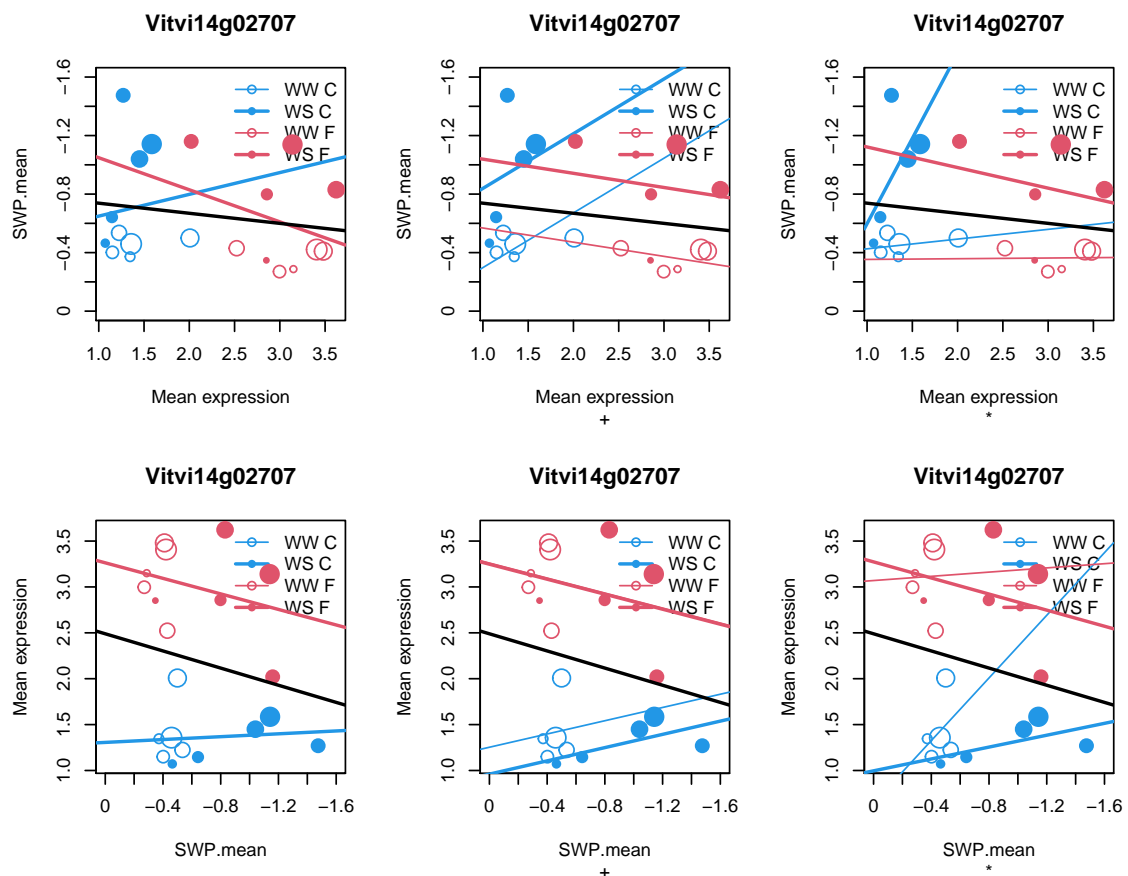
201606

Coefficient swp for Vitvi14g02707.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi14g02707	0.005784772	2.182838	0.003387544	0.9973054	0.9978996

B

Vitvi14g02707 -6.117782



Correlation of all measurements (not just means):  $r = 4e - 04, r^2 = 1e - 07$

Correlation of mean values (plots):  $r_2 = -0.1795$

### 5.3.11 Vitvi12g02492

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi12g02492

30.2.12

signalling.receptor.kinases.leucine.rich.repeat.XII

receptor.like.protein.22 |

Chr2:13853897-13855666 REVERSE LENGTH=589 |

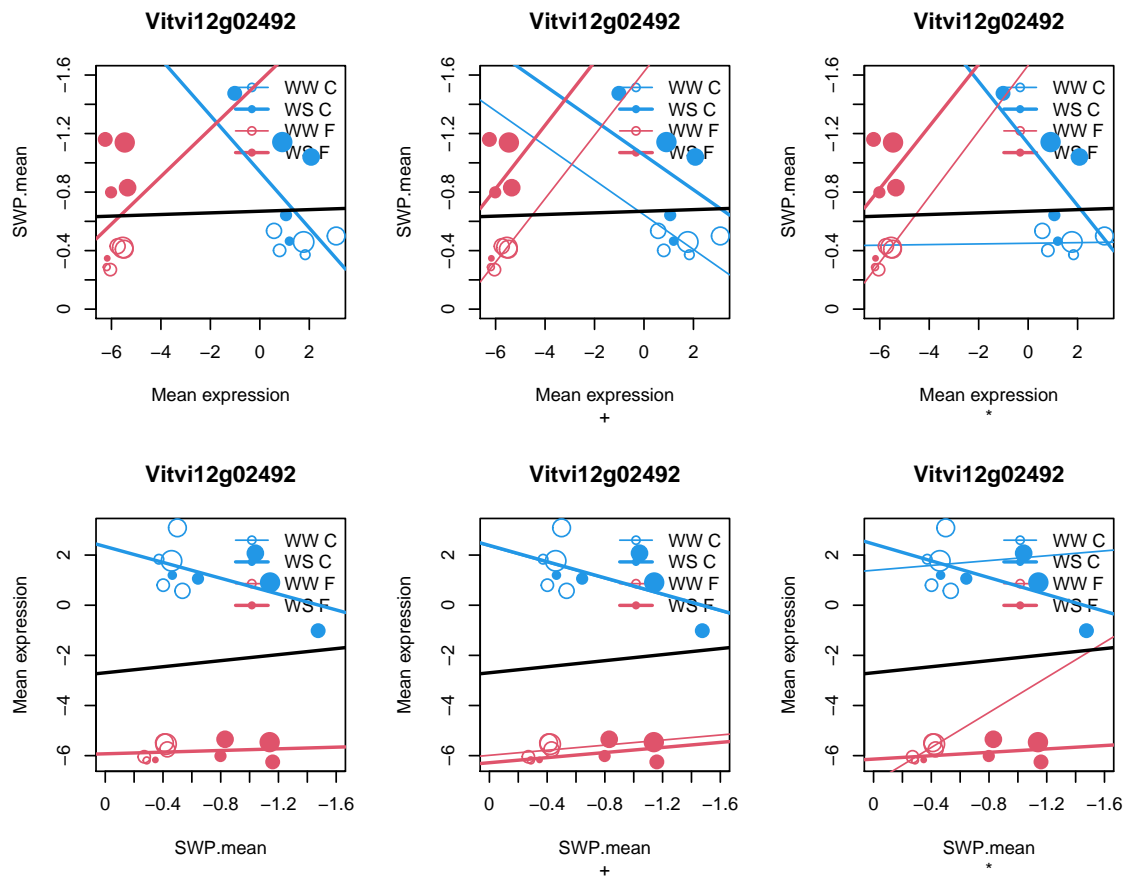
201606

Coefficient swp for Vitvi12g02492.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi12g02492	-0.02233725	-2.298563	-0.003381299	0.9973104	0.9978996

B

Vitvi12g02492 -6.117782



Correlation of all measurements (not just means):  $r = -4e - 04, r^2 = 1e - 07$

Correlation of mean values (plots):  $r_2 = 0.0579$

### 5.3.12 Vitvi16g01192

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi16g01192

28.1

DNA.synthesis/chromatin structure

P-loop containing nucleoside triphosphate hydrolases superfamily prote

Chr1:24477043-24480728 REVERSE LENGTH=1050 |

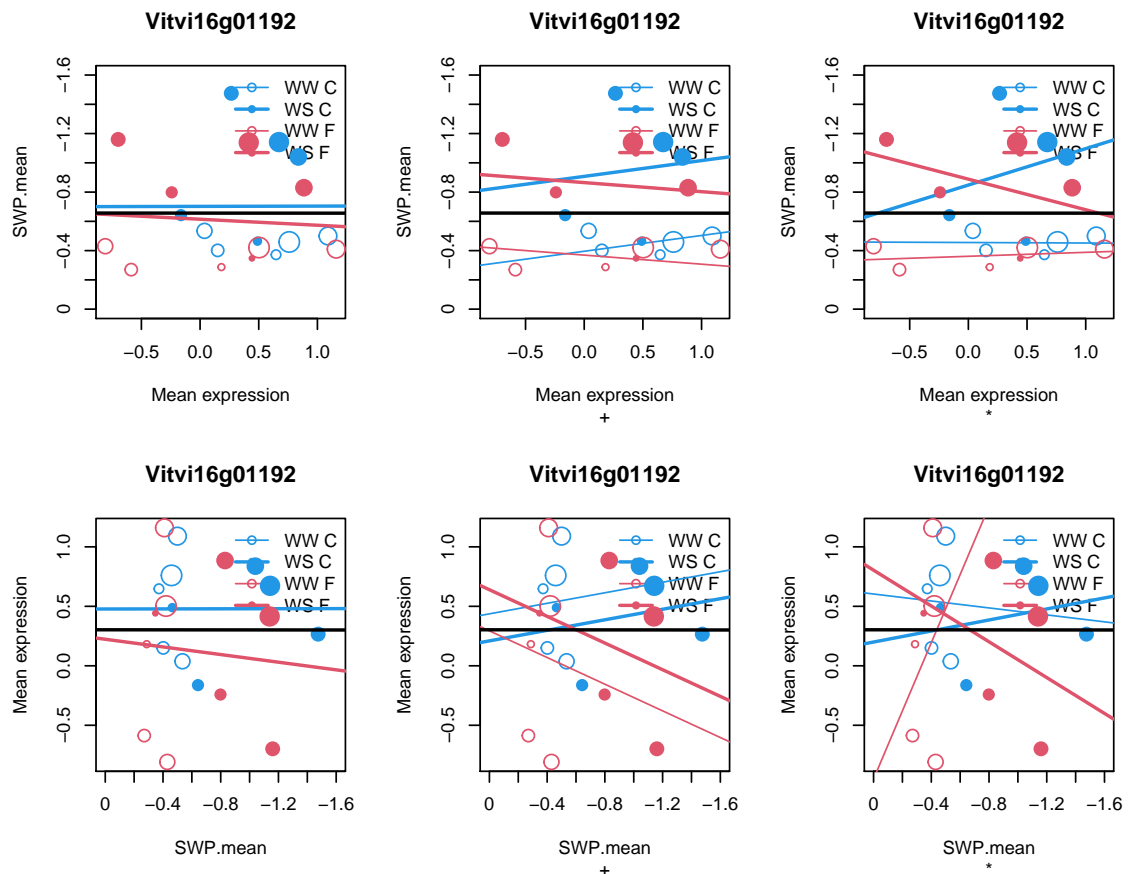
201606

Coefficient swp for Vitvi16g01192.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi16g01192	-0.003194491	0.3020401	-0.002557562	0.9979656	0.9984897

B

Vitvi16g01192 -6.117784



Correlation of all measurements (not just means):  $r = -3e - 04$ ,  $r^2 = 8e - 08$

Correlation of mean values (plots):  $r_2 = -6e - 04$

### 5.3.13 Vitvi14g00080

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi14g00080

35.2

not assigned.unknown

Polynucleotidyl transferase%2C ribonuclease H-like superfamily protein

Chr3:3946267-3946959 REVERSE LENGTH=230 |

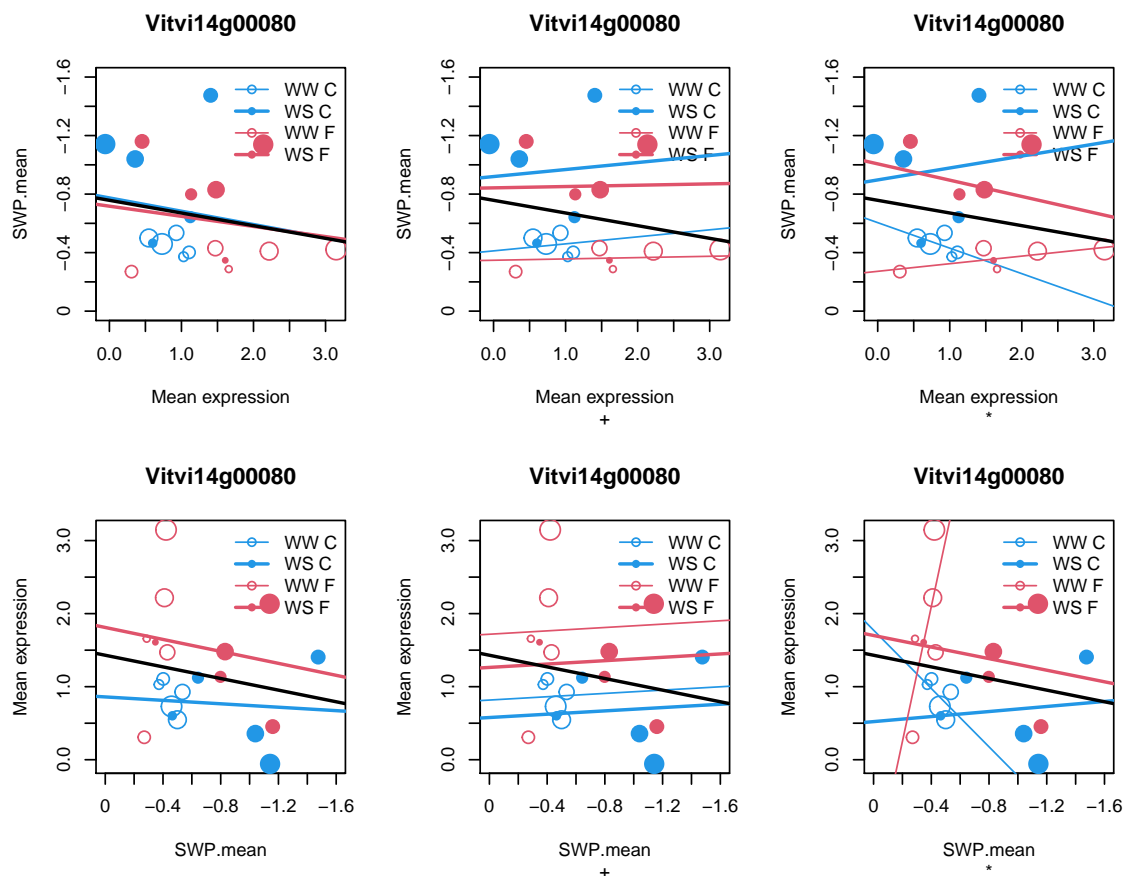
201606

Coefficient swp for Vitvi14g00080.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi14g00080	0.005137988	1.169004	0.002383192	0.9981043	0.9985629

B

Vitvi14g00080 -6.117785



Correlation of all measurements (not just means):  $r = 3e - 04, r^2 = 7e - 08$

Correlation of mean values (plots):  $r_2 = -0.1857$

### 5.3.14 Vitvi18g02570

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi18g02570

35.2

not assigned.unknown

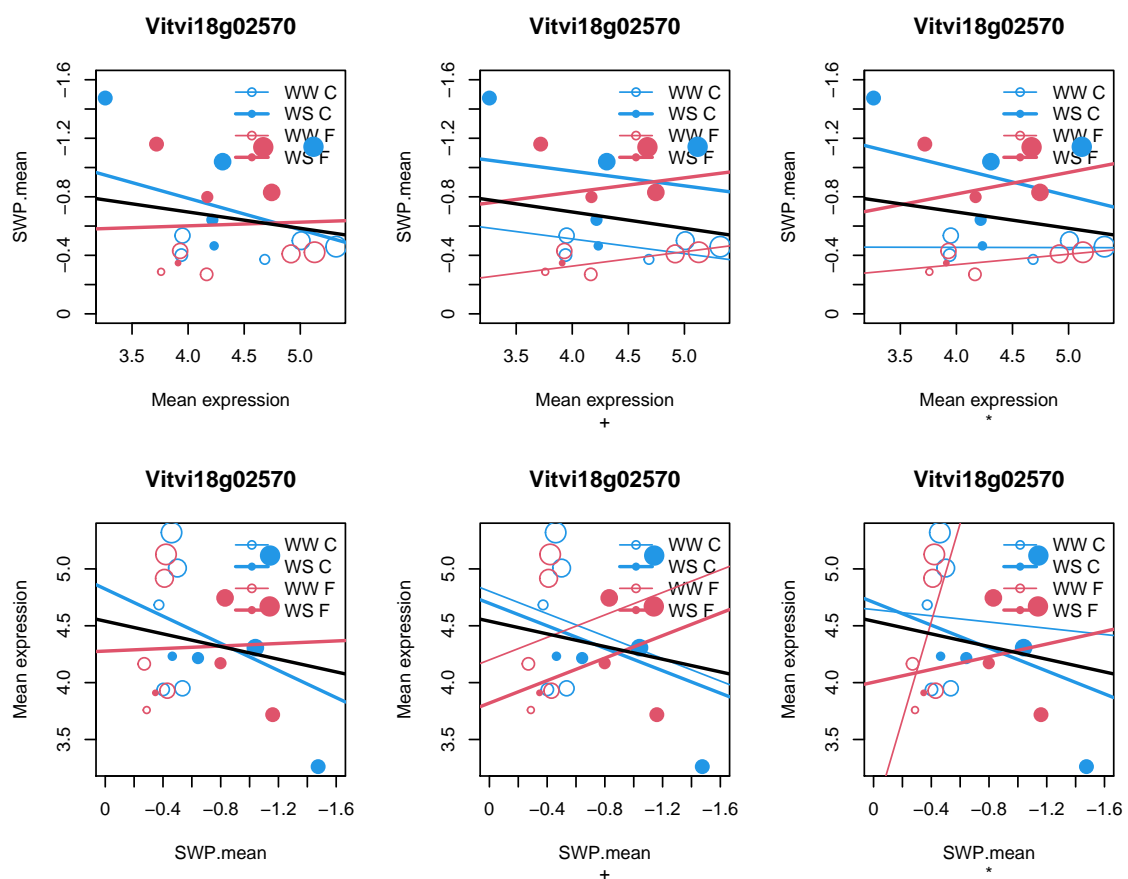
Glucan 1 3-beta-glucosidase IPR001547 Glycoside hydrolase, family 5

Coefficient swp for Vitvi18g02570.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi18g02570	0.002364193	4.35735	0.002093285	0.9983349	0.9987281

B

Vitvi18g02570	-6.117785
---------------	-----------



Correlation of all measurements (not just means):  $r = 2e - 04$ ,  $r^2 = 6e - 08$

Correlation of mean values (plots):  $r_2 = -0.1764$

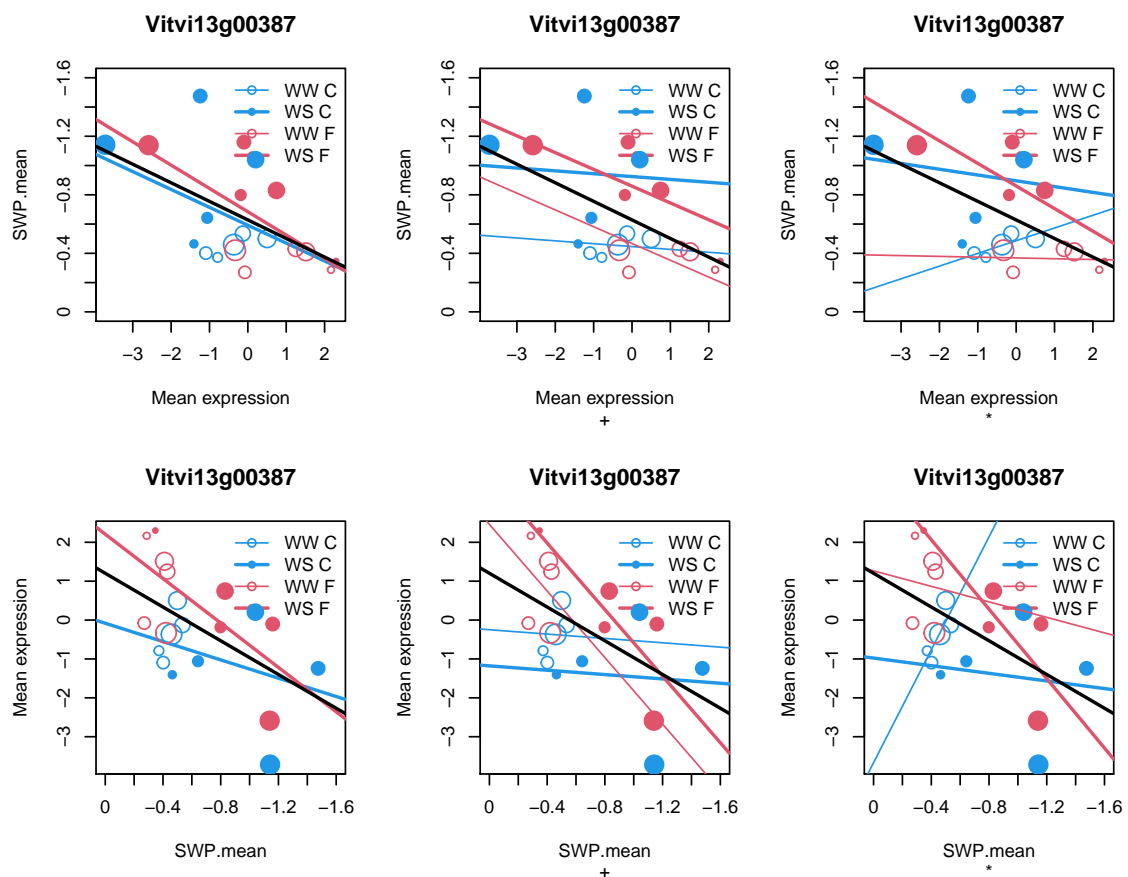
### 5.3.15 Vitvi13g00387

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

```
Vitvi13g00387
  35.2
not assigned.unknown
NA
```

Coefficient swp for Vitvi13g00387.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi13g00387	-0.00537456	-0.2205684	-0.001790019	0.9985761	0.9988888
	B				
Vitvi13g00387	-6.117786				



Correlation of all measurements (not just means):  $r = -2e - 04$ ,  $r^2 = 4e - 08$   
 Correlation of mean values (plots):  $r_2 = -0.5247$

### 5.3.16 Vitvi05g00637

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi05g00637

31.1

cell.organisation

Ankyrin repeat family protein |

Chr4:1554420-1556571 FORWARD LENGTH=629 |

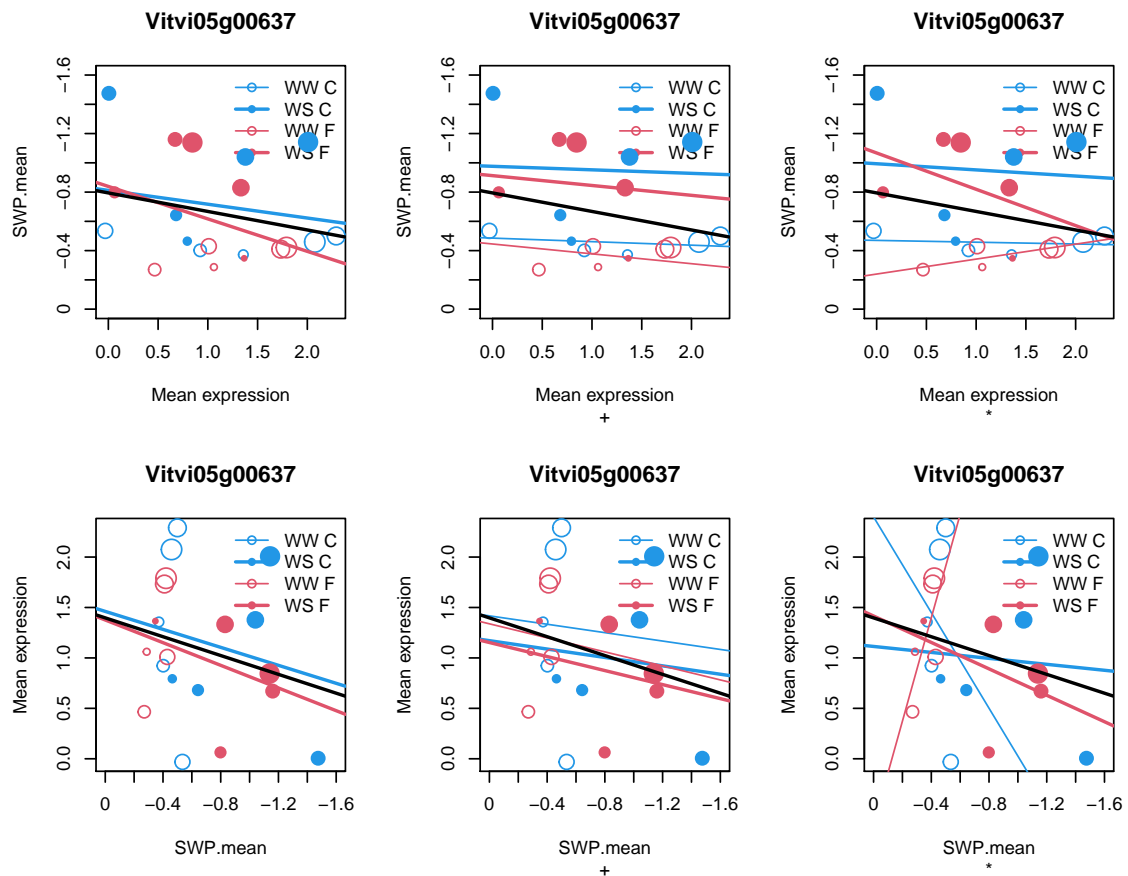
201606

Coefficient swp for Vitvi05g00637.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi05g00637	0.002396486	1.090674	0.001721486	0.9986307	0.9988888

B

Vitvi05g00637 -6.117786



Correlation of all measurements (not just means):  $r = 2e - 04, r^2 = 4e - 08$

Correlation of mean values (plots):  $r_2 = -0.2430$



### 5.3.17 Vitvi19g00151

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi19g00151

35.2

not assigned.unknown

TAF RNA polymerase I subunit A |

Chr1:19840486-19842692 REVERSE LENGTH=474 |

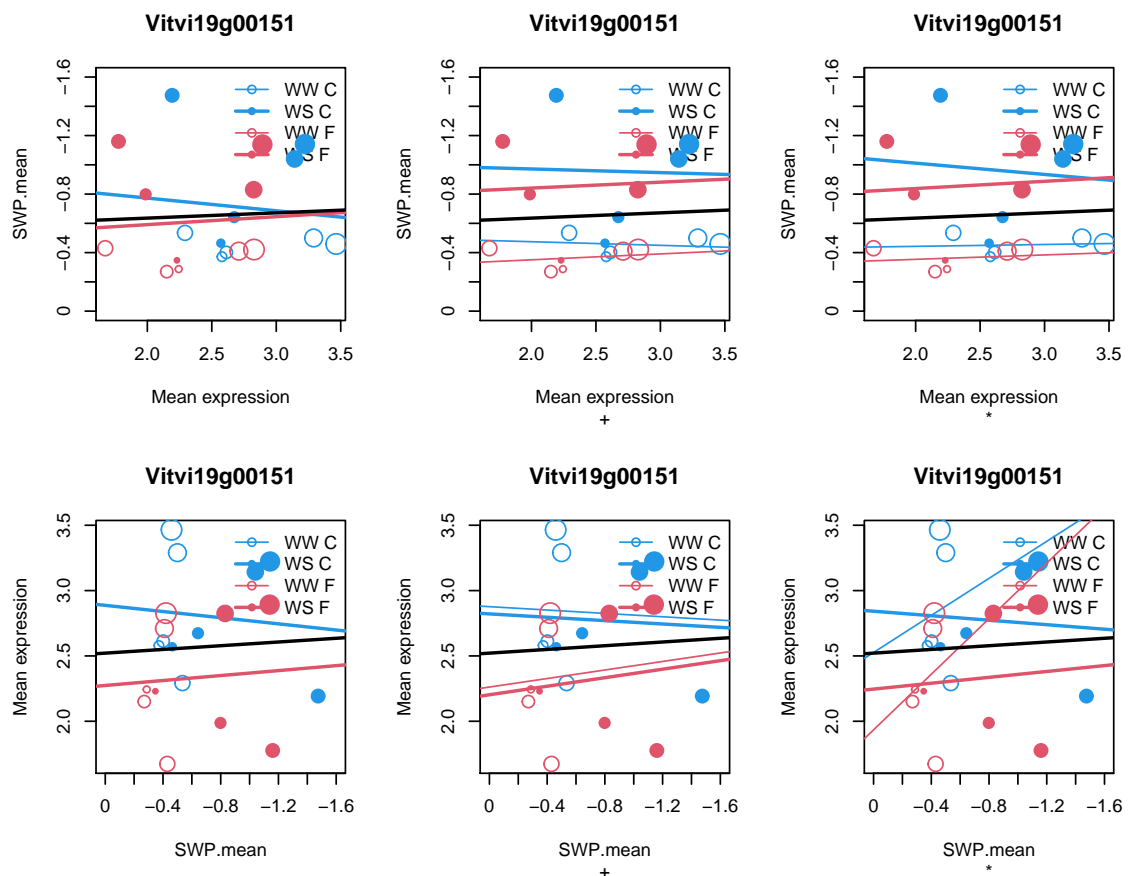
201606

Coefficient swp for Vitvi19g00151.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi19g00151	0.001547361	2.567847	0.001644123	0.9986922	0.9988888

B

Vitvi19g00151 -6.117786



Correlation of all measurements (not just means):  $r = 2e - 04, r^2 = 3e - 08$

Correlation of mean values (plots):  $r_2 = 0.0506$

### 5.3.18 Vitvi16g00190

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi16g00190

27.01.2019

RNA.processing.ribonucleases

exoribonuclease 4 |

Chr1:20350697-20356650 FORWARD LENGTH=889 |

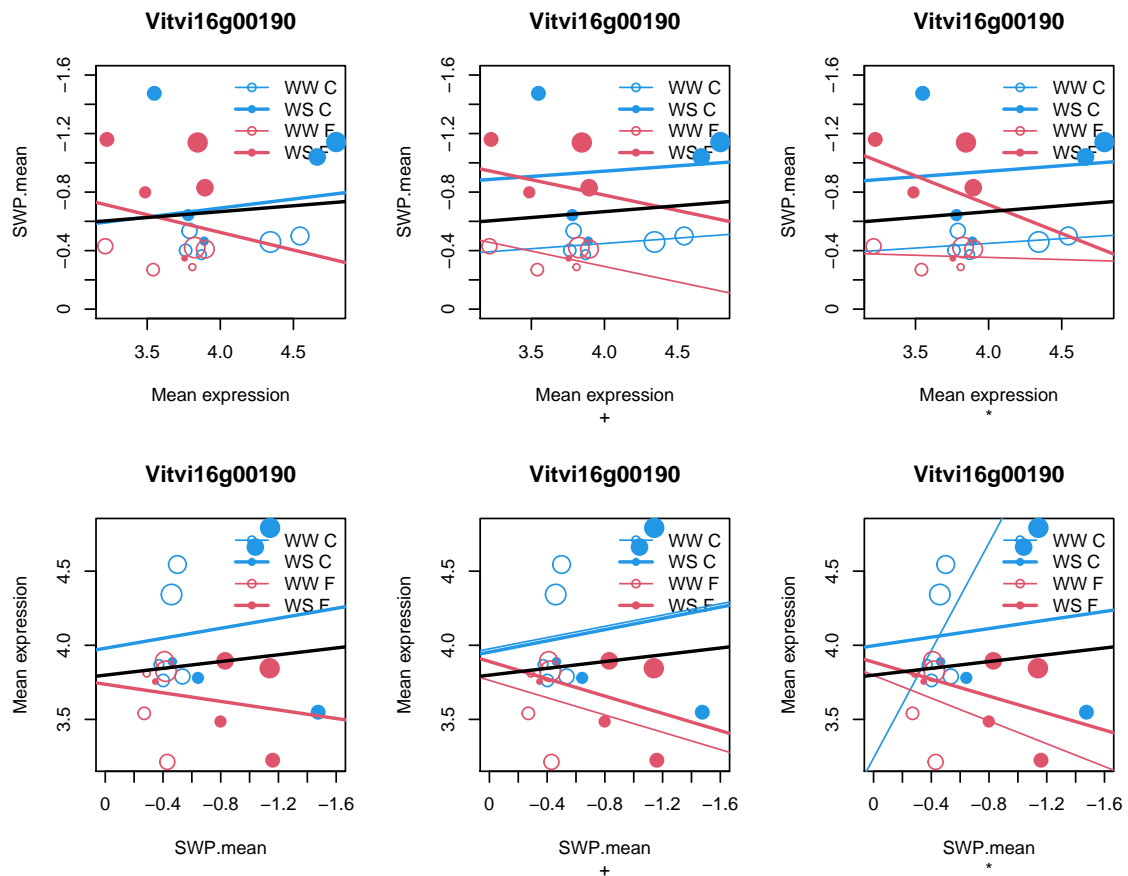
201606

Coefficient swp for Vitvi16g00190.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi16g00190	-0.000811887	3.873747	-0.0009974645	0.9992066	0.9993377

B

Vitvi16g00190 -6.117787



Correlation of all measurements (not just means):  $r = -1e - 04$ ,  $r^2 = 1e - 08$

Correlation of mean values (plots):  $r_2 = 0.0960$

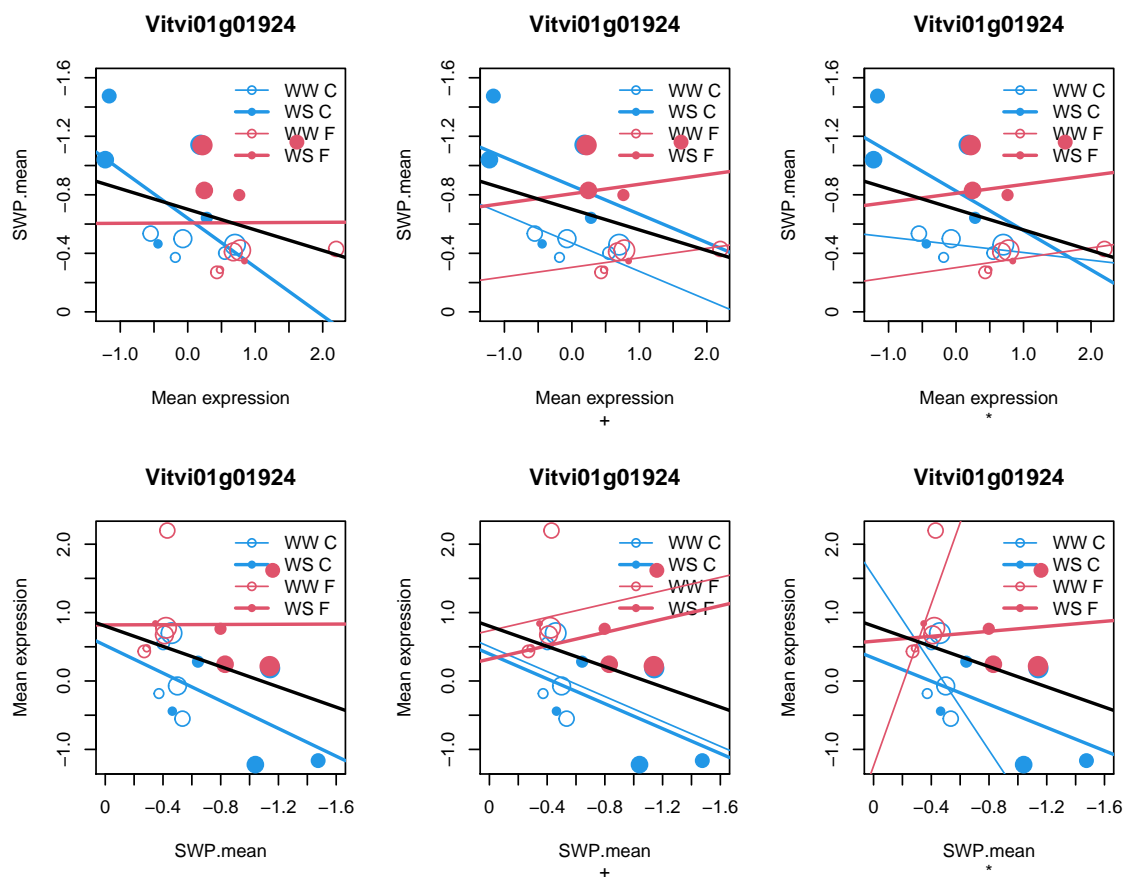
### 5.3.19 Vitvi01g01924

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

```
Vitvi01g01924
35.2
not assigned.unknown
Protein IDA
```

Coefficient swp for Vitvi01g01924.

	logFC	AveExpr	t	P.Value
Vitvi01g01924	-0.001325135	0.3165117	-0.0007298412	0.9994195
	adj.P.Val	B		
Vitvi01g01924	0.999485	-6.117787		



Correlation of all measurements (not just means):  $r = -8e - 05$ ,  $r^2 = 7e - 09$   
 Correlation of mean values (plots):  $r_2 = -0.3230$

### 5.3.20 Vitvi17g00289

```
> cat.feature(fdata[fdata$geneID %in% varname,2:5])
```

Vitvi17g00289

30.2.20

signalling.receptor.kinases.wheat LRK10 like

Serine/Threonine kinase family catalytic domain protein |

Chr1:6327463-6329935 FORWARD LENGTH=654 |

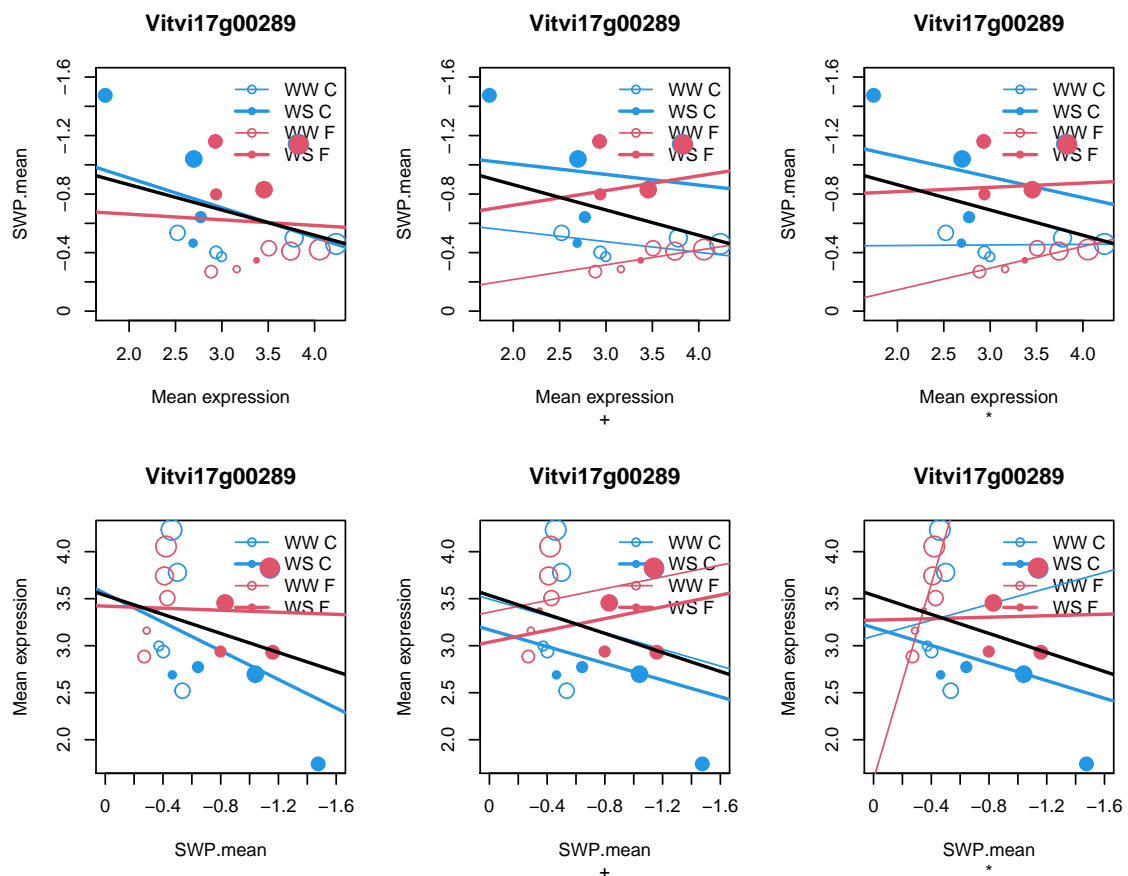
201606

Coefficient swp for Vitvi17g00289.

	logFC	AveExpr	t	P.Value	adj.P.Val
Vitvi17g00289	-0.0004491722	3.203332	-0.000382496	0.9996957	0.9996957

B

Vitvi17g00289 -6.117787



Correlation of all measurements (not just means):  $r = -4e - 05$ ,  $r^2 = 2e - 09$

Correlation of mean values (plots):  $r_2 = -0.2958$

# A Metadata files

## A.1 Project metadata

Table 1: Project metadata

Item	Value
project:	_p_VinskaTrta
Short Name:	VinskaTrta
Title:	Vine related research
Description:	*
pISA projects path:	D:/DEJAVNOSTI/OMIKE/pISA-projects
Local pISA-tree organisation:	NIB
pISA project creation date:	2020-11-13
pISA project creator:	AB
Project funding code:	*
Project coordinator:	Marusa Pompe Novak
Project partners:	*
Project start date:	*
Project end date:	*
Principal investigator:	*
License:	CC BY 4.0
Sharing permission:	Private
Upload to FAIRDOMHub:	Yes

## A.2 Investigation metadata

Table 2: Investigation metadata

Item	Value
Investigation:	_I_EnViRoS
Short Name:	EnViRoS
Title:	Integration of data from different platforms
Description:	*
Phenodata:	./phenodata_20201113.txt
pISA Investigation creation date:	2020-11-13
pISA Investigation creator:	AB
Principal investigator:	*
License:	CC BY 4.0
Sharing permission:	Private
Upload to FAIRDOMHub:	Yes

## A.3 Study metadata

Table 3: Study metadata

Item	Value
Study:	_S_01_Integ
Short Name:	01_Integ
Title:	*
Description:	*
Raw Data:	
pISA Study creation date:	2020-11-13
pISA Study creator:	AB
Principal investigator:	*
License:	CC BY 4.0
Sharing permission:	Private
Upload to FAIRDOMHub:	Yes

## A.4 Assay metadata

Table 4: Assay metadata

Item	Value
Assay:	_A_01_Desc-R
Short Name:	01_Desc-R
Assay Class:	DRY
Assay Type:	R
Title:	Data overview and descriptive statistical presentation
Description:	First we will organize the data and perform preliminary data analyses to overview what we have.
pISA Assay creation date:	2020-11-13
pISA Assay creator:	Andrej Blejec
Analyst:	Andrej Blejec
Phenodata:	Phenodata_20201109.txt
Featuredata:	Transcripts1819-unique.txt
Transcript data 18:	/input/Transcripts 2018.txt
Transcript data 19:	/input/Transcripts 2019.txt
Water potential data:	/input/Stem water potential 2018 2019.txt

## B SessionInfo

Windows 10 x64 (build 19041)

- R version 4.0.2 (2020-06-22), x86\_64-w64-mingw32
- Locale: LC\_COLLATE=Slovenian\_Slovenia.1250, LC\_CTYPE=Slovenian\_Slovenia.1250, LC\_MONETARY=Slovenian\_Slovenia.1250, LC\_NUMERIC=C, LC\_TIME=Slovenian\_Slovenia.1250
- Running under: Windows 10 x64 (build 19041)
- Matrix products: default
- Base packages: base, datasets, graphics, grDevices, methods, parallel, stats, utils
- Other packages: amisc 0.1.0, Biobase 2.50.0, BiocGenerics 0.36.0, Formula 1.2-4, ggplot2 3.3.2, Hmisc 4.4-1, knitr 1.30, lattice 0.20-41, limma 3.46.0, lubridate 1.7.9.2, reshape2 1.4.4, survival 3.2-7, xtable 1.8-4
- Loaded via a namespace (and not attached): backports 1.2.0, base64enc 0.1-3, checkmate 2.0.0, cluster 2.1.0, colorspace 1.4-1, compiler 4.0.2, crayon 1.3.4, data.table 1.13.2, digest 0.6.27, dplyr 1.0.2, ellipsis 0.3.1, evaluate 0.14, foreign 0.8-80, generics 0.1.0, glue 1.4.2, grid 4.0.2, gridExtra 2.3, gtable 0.3.0, highr 0.8, htmlTable 2.1.0, htmltools 0.5.0, htmlwidgets 1.5.2, jpeg 0.1-8.1, latticeExtra 0.6-29, lifecycle 0.2.0, magrittr 1.5, Matrix 1.2-18, munsell 0.5.0, nnet 7.3-14, pillar 1.4.6, pkgconfig 2.0.3, plyr 1.8.6, png 0.1-7, purrr 0.3.4, R6 2.5.0, RColorBrewer 1.1-2, Rcpp 1.0.5, rlang 0.4.8, rpart 4.1-15, rstudioapi 0.12, scales 1.1.1, splines 4.0.2, stringi 1.5.3, stringr 1.4.0, tibble 3.0.4, tidyr 1.1.0, tools 4.0.2, vctrs 0.3.4, withr 2.3.0, xfun 0.19

Analysis project path: [\[link\]](#)

Local

D:

DEJAVNOSTI

OMIKE

pISA-projects

\_p\_VinskaTrta

\_I\_EnViRoS

\_S\_01\_Integ

\_A\_01\_Desc-R

other

Network

O:

DEJAVNOSTI

OMIKE

pISA-projects

\_p\_VinskaTrta

\_I\_EnViRoS

\_S\_01\_Integ

\_A\_01\_Desc-R

other

Main file : [../doc/40\\_Expression-water-stress.Rnw](#)

Main file : [../scripts/40\\_Expression-water-stress.Rnw](#)

Project file: [\[link\]](#)

## View as vignette

Source files can be viewed by pasting this code to R console:

```
projectName <-"other"
mainFile <-"40_Expression-water-stress"
commandArgs()
library(tkWidgets)
openPDF(file.path(dirname(getwd()),"doc",
paste(mainFile,"PDF",sep=".")))
viewVignette("viewVignette", projectName,
file.path("../doc",paste(mainFile,"Rnw",sep=".")))
```