

Analysis of transcriptomics and metabolomics data

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March 9, 2021

Parameters:

```
.years = 18  
.ver = 1  
.bins1 = 1  
.bins2 = 2  
.bins3 = 3  
.bins4 = 4  
.bins5 = 5  
.bins6 = 6  
.bins7 = 7
```

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1 Task

Analysis of transcriptomics and metabolomics data with canonical correlation.
Main ideas are from package **mixOmics**.

Final document in folder /reports:

```
> fileName(outputFile)
```

```
[1] "60_Trans-x-Meta_18-1-1-2-3-4-5-6-7"
```

2 Information from pISA

Data directory

```
> .inroot
```

```
[1] "../..input"
```

Results directory

```
> .oroot
```

```
[1] "../..output/60_Trans-x-Meta-18-1-1-2-3-4-5-6-7"
```

```
project:      _p_VinskaTrta
```

```
Investigation: _I_EnViRoS
```

```
Study:        _S_01_Integ
```

```
Assay:         _A_01_Desc-R
```

3 Data

Priporočljivo je najprej prebrati phenodata in feature-data, potem pa podatke. To omogoča izbor spremenljivk in vzorcev takoj za tem, ko podatke preberemo.

```
> cat(knit_child(file.path("../doc", "10a_Read-Data.Rnw"), quiet=TRUE))
```

Entered 10a-ReadData.Rnw

3.1 Phenodata

Two incomplete lines in original phenodata were manually completed. Sample ids (ID) were parsed in Excel to form factors of interest: variety, year and day.

```

> (pfn <- getDesc(.adesc
+      , "Phenodata"))
[1] "Phenodata_20201109.txt"
> dir(file.path(.iroot), pattern = pfn)
[1] "Phenodata_20201109.txt"
> phdata <- read.table(file.path(.iroot, pfn)
+      , header = TRUE
+      , sep = "\t"
+      , stringsAsFactors = FALSE
+      )
> rownames(phdata) <- phdata[,1]
> my.summary(phdata)

```

ID	Variety	Date	variety
C18_11d_WS1: 1	Cabernet Volos:134	21.06.2018: 20	C:134
C18_11d_WS2: 1	Fleurtaï :136	26.06.2019: 20	F:136
C18_11d_WS3: 1		7.08.2018 : 20	
C18_11d_WS4: 1		8.07.2019 : 20	
C18_11d_WW1: 1		1.08.2018 : 16	
C18_11d_WW2: 1		18.07.2019: 16	
(Other) :264		(Other) :158	

```

year      day      treat      rep      project.name
18:136    34      : 36      WS:135    1      :48      EnViros:270
19:134    49      : 24      WW:135    2      :48
          20      : 20                3      :48
          22      : 20                4      :48
          67      : 20                NA's:78
          44      : 16
          (Other):134

```

species	plant.name	tissue	health.status
Vitis vinifera:270	grapevine:270	: 78	water stress:134
		leaf:192	well watered:136

```

plant.number growth.location growth.conditions
:78          Udine:270          outside:270
R1:48
R2:48
R3:48
R4:48

```

several leaves ground together and stored at -80oC; an aliquot was received f


```

Metabolites.Order                               Metabolites.File.Name
2      : 2                                     : 80
3      : 2      1019Cabernet Volos_6_12_WS1.D: 1
4      : 2      1019Cabernet Volos_6_12_WS2.D: 1
5      : 2      1019Cabernet Volos_6_12_WW1.D: 1
6      : 2      1019Cabernet Volos_6_12_WW2.D: 1
(Other):180      1019Cabernet Volos_6_21_WS1.D: 1
NA's   : 80      (Other)                       :185

```

```

Metabolites.File Transcripts.ID
: 80 :190
Cabernet Volos08_07WS: 4 C1_S1 : 1
Cabernet Volos08_07WW: 4 C1_S2 : 1
Cabernet Volos08_08WS: 4 C1_S3 : 1
Cabernet Volos08_08WW: 4 C1_S4 : 1
Cabernet Volos18_07WS: 4 C1_W1 : 1
(Other) :170 (Other): 75

```

```
> dim(phdata)
```

```
[1] 270 21
```

```
> names(phdata)
```

```

[1] "ID" "Variety"
[3] "Date" "variety"
[5] "year" "day"
[7] "treat" "rep"
[9] "project.name" "species"
[11] "plant.name" "tissue"
[13] "health.status" "plant.number"
[15] "growth.location" "growth.conditions"
[17] "sampling" "Metabolites.Order"
[19] "Metabolites.File.Name" "Metabolites.File"
[21] "Transcripts.ID"

```

3.2 Declare factor types

In 2018, day 11 is present only for C and day 10 only for F. The two day points coincide and are changed to a common value 11.

```

> for(yr in levels(phdata$year)){
+ cat("\n\nYear:", yr, "\n")
+ print(
+ with(phdata[phdata$year==yr, ], ftable(treat, variety, factor(day)))
+ )
+ }
> phdata[phdata$day==10, "day"] <- 11

```

Factors

```

> factors <- colnames(phdata)[4:8]
> factors
[1] "variety" "year"      "day"        "treat"      "rep"
> for (varname in factors)
+ phdata[,varname] <- factor(phdata[,varname])
> #
> # Change the order of levels for treatment
> phdata$treat <- factor(phdata$treat, levels=c("WW","WS"))
> str(phdata[,factors])
'data.frame':      270 obs. of  5 variables:
 $ variety: Factor w/ 2 levels "C","F": 1 1 1 1 1 1 1 1 1 1 ...
 $ year   : Factor w/ 2 levels "18","19": 1 1 1 1 1 1 1 1 1 1 ...
 $ day    : Factor w/ 26 levels "3","11","14",...: 2 2 2 2 2 2 2 2 4 4 ...
 $ treat  : Factor w/ 2 levels "WW","WS": 2 2 2 2 1 1 1 1 2 2 ...
 $ rep    : Factor w/ 4 levels "1","2","3","4": 1 2 3 4 1 2 3 4 1 2 ...

```

Date of sampling might be useful as well

```

> library(lubridate)
Warning: package 'lubridate' was built under R version 4.0.3

```

Attaching package: 'lubridate'

The following objects are masked from 'package:BiocGenerics':

```
intersect, setdiff, union
```

The following objects are masked from 'package:base':

```
date, intersect, setdiff, union
```

```

> phdata$date <- as_date(phdata$Date, format="%d.%m.%Y")
> table(data.frame(phdata$date, "Count"=1))

```

	Count
phdata.date	1
2018-06-04	4
2018-06-11	8
2018-06-12	8
2018-06-15	4
2018-06-21	20
2018-06-29	4
2018-07-04	4
2018-07-05	16
2018-07-10	4
2018-07-16	4
2018-07-20	16
2018-07-23	4
2018-07-31	4
2018-08-01	16
2018-08-07	20
2019-06-18	4
2019-06-26	20
2019-07-08	20
2019-07-12	4
2019-07-17	4

```

2019-07-18 16
2019-07-21 8
2019-07-22 2
2019-07-23 8
2019-07-29 16
2019-07-30 4
2019-08-06 4
2019-08-08 16
2019-08-12 4
2019-08-19 4

```

3.3 Select samples

Check for assay specific sample selection column (assay name)

```

> .aName
[1] "_A_01_Desc-R"
> selectId <- substr(gsub("-", ".", .aName), 2, nchar(.aName))
> #selectId <- .vzorci
> selectId
[1] "A_01_Desc.R"
> #
> if(selectId %in% names(phdata)){
+ pdata <- phdata[!is.na(phdata[,selectId]),]
+ cat("Sample selection column found (", selectId, ")\n",
+ nrow(pdata), "samples will be used.\n")
+ } else {
+ pdata <- phdata
+ cat("No sample selection column found,\n",
+ nrow(pdata), "samples will be used.\n")
+ }
No sample selection column found,
 270 samples will be used.
> dim(pdata)
[1] 270  22

```

Table of sample conditions

```

> head(pdata[, factors])
      variety year day treat rep
C18_11d_WS1      C   18  11   WS   1
C18_11d_WS2      C   18  11   WS   2
C18_11d_WS3      C   18  11   WS   3
C18_11d_WS4      C   18  11   WS   4
C18_11d_WW1      C   18  11   WW   1
C18_11d_WW2      C   18  11   WW   2
> for(yr in levels(pdata$year)){
+ cat("\n\nYear: ", yr, "\n")
+ print(
+ with(pdata[pdata$year==yr, ], ftable(treat, variety, factor(day)))
+ )
+ }

```

Year: 18

		3	11	14	20	28	33	34	39	45	49	52	60	61	67
treat	variety														
WW	C	1	4	1	5	1	1	4	1	1	4	1	1	4	5
	F	1	4	1	5	1	1	4	1	1	4	1	1	4	5
WS	C	1	4	1	5	1	1	4	1	1	4	1	1	4	5
	F	1	4	1	5	1	1	4	1	1	4	1	1	4	5

Year: 19

		14	22	34	38	43	44	47	48	49	55	56	63	65	69	76
treat	variety															
WW	C	1	5	5	1	1	4	0	0	4	4	1	1	4	1	1
	F	1	5	5	1	1	4	4	1	0	4	1	1	4	1	1
WS	C	1	5	5	1	1	4	0	0	4	4	1	1	4	1	1
	F	1	5	5	1	1	4	4	1	0	4	1	1	4	1	1

> my.summary(pdata)

	ID		Variety		Date		variety
C18_11d_WS1:	1	Cabernet	Volos:134		21.06.2018:	20	C:134
C18_11d_WS2:	1	Fleurtaï	:136		26.06.2019:	20	F:136
C18_11d_WS3:	1				7.08.2018 :	20	
C18_11d_WS4:	1				8.07.2019 :	20	
C18_11d_WW1:	1				1.08.2018 :	16	
C18_11d_WW2:	1				18.07.2019:	16	
(Other)	:264				(Other)	:158	
year	day	treat	rep		project.name		
18:136	34	: 36	WW:135	1	:48	EnViros:270	
19:134	49	: 24	WS:135	2	:48		
	20	: 20		3	:48		
	22	: 20		4	:48		
	67	: 20		NA's:78			
	11	: 16					
	(Other)	:134					
species	plant.name	tissue	health.status				
Vitis vinifera:270	grapevine:270	: 78	water stress:134				
		leaf:192	well watered:136				

plant.number	growth.location	growth.conditions
:78	Udine:270	outside:270
R1:48		
R2:48		
R3:48		
R4:48		

several leaves ground together and stored at -80oC; an aliquot was received f

```

Metabolites.Order                      Metabolites.File.Name
2      : 2                               : 80
3      : 2      1019Cabernet Volos_6_12_WS1.D: 1
4      : 2      1019Cabernet Volos_6_12_WS2.D: 1
5      : 2      1019Cabernet Volos_6_12_WW1.D: 1
6      : 2      1019Cabernet Volos_6_12_WW2.D: 1
(Other):180      1019Cabernet Volos_6_21_WS1.D: 1
NA's   : 80      (Other)                    :185

      Metabolites.File Transcripts.ID      date
      : 80                :190      2018-06-21: 20
Cabernet Volos08_07WS: 4      C1_S1 : 1      2018-08-07: 20
Cabernet Volos08_07WW: 4      C1_S2 : 1      2019-06-26: 20
Cabernet Volos08_08WS: 4      C1_S3 : 1      2019-07-08: 20
Cabernet Volos08_08WW: 4      C1_S4 : 1      2018-07-05: 16
Cabernet Volos18_07WS: 4      C1_W1 : 1      2018-07-20: 16
(Other)                :170      (Other): 75      (Other) :158

```

For separate yearly analyses, it will be handy to have two, separate phenodata objects.

```

> dim(pdata)
[1] 270 22
> table(pdata$year)

 18 19
136 134
> pdata18 <- pdata[pdata$year==18,]
> dim(pdata18)
[1] 136 22
> pdata18[1:5,1:6]
      ID      Variety      Date variety year day
C18_11d_WS1 C18_11d_WS1 Cabernet Volos 12.06.2018      C    18 11
C18_11d_WS2 C18_11d_WS2 Cabernet Volos 12.06.2018      C    18 11
C18_11d_WS3 C18_11d_WS3 Cabernet Volos 12.06.2018      C    18 11
C18_11d_WS4 C18_11d_WS4 Cabernet Volos 12.06.2018      C    18 11
C18_11d_WW1 C18_11d_WW1 Cabernet Volos 12.06.2018      C    18 11
> #
> pdata19 <- pdata[pdata$year==19,]
> dim(pdata19)
[1] 134 22
> pdata19[1:5,1:6]
      ID      Variety      Date variety year day
C19_22d_WS1 C19_22d_WS1 Cabernet Volos 26.06.2019      C    19 22
C19_22d_WS2 C19_22d_WS2 Cabernet Volos 26.06.2019      C    19 22
C19_22d_WS3 C19_22d_WS3 Cabernet Volos 26.06.2019      C    19 22
C19_22d_WS4 C19_22d_WS4 Cabernet Volos 26.06.2019      C    19 22
C19_22d_WW1 C19_22d_WW1 Cabernet Volos 26.06.2019      C    19 22

```

3.4 Featuredata

```
> (ffn <- getDesc(.adesc, "Featuredata"))
[1] "Transcripts1819-02.txt"
> if(ffn=="") {
+ cat("No feature data declared in the assay metadata file.\\\\"")
+ fdata <- NULL
+ }

> fdata <- read.table(file.path(.iroot, ffn)
+ , sep = "\t"
+ , header = TRUE
+ , na.strings = c("", "-", "NA")
+ , stringsAsFactors = FALSE
+ , quote = "\"",
+ # , row.names = 1
+ )
> if(interactive()) my.summary(fdata)
> #"
```

Some genes are listed more than once. Is it OK? Column geneID can not be used as row names.

```
> tbl <- table(fdata$geneID)
> tbl[tbl>3]
named integer(0)
> tbl[tbl==3]

Vitvi01g01443 Vitvi03g00845 Vitvi04g02157 Vitvi05g01641 Vitvi06g01689
      3          3          3          3          3
Vitvi07g01861 Vitvi08g01514 Vitvi08g01692 Vitvi08g02189 Vitvi12g01199
      3          3          3          3          3
Vitvi16g00114 Vitvi17g01011 Vitvi19g00061 Vitvi19g01931 Vitvi19g01936
      3          3          3          3          3
Vitvi19g01964
      3

> tbl[tbl==2]

Vitvi00g00711 Vitvi00g00822 Vitvi00g00983 Vitvi00g01149 Vitvi00g01320
      2          2          2          2          2
Vitvi00g01329 Vitvi00g01996 Vitvi00g01999 Vitvi00g02041 Vitvi01g00036
      2          2          2          2          2
Vitvi01g00137 Vitvi01g00165 Vitvi01g00274 Vitvi01g00301 Vitvi01g00348
      2          2          2          2          2
Vitvi01g00373 Vitvi01g00397 Vitvi01g00398 Vitvi01g00417 Vitvi01g00478
      2          2          2          2          2
Vitvi01g00553 Vitvi01g00625 Vitvi01g00681 Vitvi01g00696 Vitvi01g00793
      2          2          2          2          2
Vitvi01g00814 Vitvi01g00828 Vitvi01g00861 Vitvi01g00885 Vitvi01g00993
      2          2          2          2          2
Vitvi01g01351 Vitvi01g01436 Vitvi01g01632 Vitvi01g01645 Vitvi01g01671
      2          2          2          2          2
```

Vitvi01g01773	Vitvi01g01814	Vitvi01g01956	Vitvi01g01982	Vitvi01g02258
2	2	2	2	2
Vitvi01g02269	Vitvi02g00007	Vitvi02g00023	Vitvi02g00085	Vitvi02g00187
2	2	2	2	2
Vitvi02g00220	Vitvi02g00222	Vitvi02g00278	Vitvi02g00380	Vitvi02g00387
2	2	2	2	2
Vitvi02g00485	Vitvi02g00561	Vitvi02g00739	Vitvi02g01143	Vitvi02g01273
2	2	2	2	2
Vitvi02g01365	Vitvi02g01416	Vitvi02g01458	Vitvi02g01751	Vitvi03g00042
2	2	2	2	2
Vitvi03g00059	Vitvi03g00097	Vitvi03g00121	Vitvi03g00167	Vitvi03g00273
2	2	2	2	2
Vitvi03g00278	Vitvi03g00375	Vitvi03g00432	Vitvi03g00442	Vitvi03g00464
2	2	2	2	2
Vitvi03g00521	Vitvi03g00532	Vitvi03g00537	Vitvi03g00541	Vitvi03g00757
2	2	2	2	2
Vitvi03g00785	Vitvi03g00804	Vitvi03g00855	Vitvi03g00900	Vitvi03g00936
2	2	2	2	2
Vitvi03g01059	Vitvi03g01228	Vitvi03g01320	Vitvi04g00092	Vitvi04g00112
2	2	2	2	2
Vitvi04g00113	Vitvi04g00127	Vitvi04g00199	Vitvi04g00205	Vitvi04g00227
2	2	2	2	2
Vitvi04g00279	Vitvi04g00314	Vitvi04g00371	Vitvi04g00450	Vitvi04g00483
2	2	2	2	2
Vitvi04g00520	Vitvi04g00888	Vitvi04g01149	Vitvi04g01257	Vitvi04g01359
2	2	2	2	2
Vitvi04g01462	Vitvi04g01469	Vitvi04g01474	Vitvi04g01560	Vitvi04g01624
2	2	2	2	2
Vitvi04g01633	Vitvi04g01668	Vitvi04g01671	Vitvi04g01694	Vitvi04g01695
2	2	2	2	2
Vitvi04g01709	Vitvi04g01746	Vitvi04g01763	Vitvi04g02022	Vitvi04g02037
2	2	2	2	2
Vitvi04g02038	Vitvi04g02099	Vitvi04g02114	Vitvi04g02185	Vitvi05g00115
2	2	2	2	2
Vitvi05g00132	Vitvi05g00144	Vitvi05g00210	Vitvi05g00239	Vitvi05g00390
2	2	2	2	2
Vitvi05g00409	Vitvi05g00418	Vitvi05g00436	Vitvi05g00443	Vitvi05g00503
2	2	2	2	2
Vitvi05g00541	Vitvi05g00556	Vitvi05g00652	Vitvi05g00737	Vitvi05g00790
2	2	2	2	2
Vitvi05g00953	Vitvi05g00991	Vitvi05g01068	Vitvi05g01130	Vitvi05g01255
2	2	2	2	2
Vitvi05g01369	Vitvi05g01562	Vitvi05g01570	Vitvi05g01603	Vitvi05g01634
2	2	2	2	2
Vitvi05g02135	Vitvi06g00022	Vitvi06g00089	Vitvi06g00200	Vitvi06g00201
2	2	2	2	2
Vitvi06g00212	Vitvi06g00213	Vitvi06g00215	Vitvi06g00243	Vitvi06g00259
2	2	2	2	2
Vitvi06g00319	Vitvi06g00337	Vitvi06g00356	Vitvi06g00365	Vitvi06g00414
2	2	2	2	2
Vitvi06g00439	Vitvi06g00443	Vitvi06g00454	Vitvi06g00472	Vitvi06g00512
2	2	2	2	2
Vitvi06g00525	Vitvi06g00533	Vitvi06g00652	Vitvi06g00668	Vitvi06g00701

2	2	2	2	2
Vitvi06g00789	Vitvi06g01242	Vitvi06g01279	Vitvi06g01469	Vitvi06g01486
2	2	2	2	2
Vitvi06g01604	Vitvi06g01690	Vitvi06g01693	Vitvi06g01718	Vitvi06g01739
2	2	2	2	2
Vitvi06g01836	Vitvi07g00067	Vitvi07g00078	Vitvi07g00080	Vitvi07g00205
2	2	2	2	2
Vitvi07g00316	Vitvi07g00406	Vitvi07g00416	Vitvi07g00464	Vitvi07g00594
2	2	2	2	2
Vitvi07g00630	Vitvi07g00636	Vitvi07g00708	Vitvi07g00908	Vitvi07g00916
2	2	2	2	2
Vitvi07g01028	Vitvi07g01205	Vitvi07g01306	Vitvi07g01340	Vitvi07g01364
2	2	2	2	2
Vitvi07g01402	Vitvi07g01441	Vitvi07g01524	Vitvi07g01569	Vitvi07g01737
2	2	2	2	2
Vitvi07g01749	Vitvi07g01817	Vitvi07g01903	Vitvi07g01937	Vitvi07g01973
2	2	2	2	2
Vitvi07g01983	Vitvi07g01996	Vitvi07g02049	Vitvi07g02102	Vitvi07g02119
2	2	2	2	2
Vitvi07g02123	Vitvi07g02135	Vitvi07g02302	Vitvi07g02394	Vitvi07g02412
2	2	2	2	2
Vitvi07g02447	Vitvi07g02491	Vitvi07g02523	Vitvi07g02528	Vitvi07g02557
2	2	2	2	2
Vitvi07g02631	Vitvi07g02863	Vitvi07g03034	Vitvi07g03040	Vitvi07g03070
2	2	2	2	2
Vitvi07g03110	Vitvi08g00215	Vitvi08g00703	Vitvi08g00740	Vitvi08g00744
2	2	2	2	2
Vitvi08g00799	Vitvi08g00853	Vitvi08g00935	Vitvi08g00961	Vitvi08g00981
2	2	2	2	2
Vitvi08g00983	Vitvi08g00999	Vitvi08g01042	Vitvi08g01082	Vitvi08g01108
2	2	2	2	2
Vitvi08g01206	Vitvi08g01241	Vitvi08g01243	Vitvi08g01302	Vitvi08g01346
2	2	2	2	2
Vitvi08g01376	Vitvi08g01395	Vitvi08g01417	Vitvi08g01506	Vitvi08g01513
2	2	2	2	2
Vitvi08g01519	Vitvi08g01766	Vitvi08g01833	Vitvi08g01864	Vitvi08g01884
2	2	2	2	2
Vitvi08g01931	Vitvi08g01959	Vitvi08g02249	Vitvi08g02367	Vitvi08g02413
2	2	2	2	2
Vitvi09g00067	Vitvi09g00156	Vitvi09g00233	Vitvi09g00239	Vitvi09g00243
2	2	2	2	2
Vitvi09g00249	Vitvi09g00253	Vitvi09g00258	Vitvi09g00277	Vitvi09g00292
2	2	2	2	2
Vitvi09g00297	Vitvi09g00446	Vitvi09g00490	Vitvi09g00653	Vitvi09g00675
2	2	2	2	2
Vitvi09g00816	Vitvi09g00861	Vitvi09g01320	Vitvi09g01500	Vitvi09g01555
2	2	2	2	2
Vitvi09g01556	Vitvi09g01557	Vitvi09g01559	Vitvi09g01930	Vitvi09g01933
2	2	2	2	2
Vitvi09g02012	Vitvi10g00078	Vitvi10g00087	Vitvi10g00125	Vitvi10g00154
2	2	2	2	2
Vitvi10g00219	Vitvi10g00290	Vitvi10g00303	Vitvi10g00316	Vitvi10g00317
2	2	2	2	2

Vitvi10g00336	Vitvi10g00365	Vitvi10g00457	Vitvi10g00554	Vitvi10g00582
2	2	2	2	2
Vitvi10g00605	Vitvi10g00635	Vitvi10g00662	Vitvi10g00663	Vitvi10g00784
2	2	2	2	2
Vitvi10g01029	Vitvi10g01098	Vitvi10g02139	Vitvi10g02141	Vitvi10g02143
2	2	2	2	2
Vitvi10g02151	Vitvi10g02152	Vitvi10g02155	Vitvi10g02157	Vitvi10g02322
2	2	2	2	2
Vitvi10g02324	Vitvi10g02368	Vitvi10g02370	Vitvi10g02408	Vitvi11g00001
2	2	2	2	2
Vitvi11g00068	Vitvi11g00102	Vitvi11g00136	Vitvi11g00137	Vitvi11g00198
2	2	2	2	2
Vitvi11g00270	Vitvi11g00339	Vitvi11g00360	Vitvi11g00480	Vitvi11g00517
2	2	2	2	2
Vitvi11g00597	Vitvi11g00614	Vitvi11g00692	Vitvi11g00724	Vitvi11g00812
2	2	2	2	2
Vitvi11g00865	Vitvi11g01141	Vitvi11g01176	Vitvi12g00021	Vitvi12g00032
2	2	2	2	2
Vitvi12g00089	Vitvi12g00121	Vitvi12g00157	Vitvi12g00220	Vitvi12g00229
2	2	2	2	2
Vitvi12g00284	Vitvi12g00596	Vitvi12g00598	Vitvi12g00606	Vitvi12g00615
2	2	2	2	2
Vitvi12g00628	Vitvi12g00646	Vitvi12g00689	Vitvi12g00691	Vitvi12g00694
2	2	2	2	2
Vitvi12g00837	Vitvi12g01277	Vitvi12g01467	Vitvi12g01780	Vitvi12g01840
2	2	2	2	2
Vitvi12g01863	Vitvi12g01897	Vitvi12g01952	Vitvi12g02074	Vitvi12g02434
2	2	2	2	2
Vitvi12g02435	Vitvi12g02440	Vitvi13g00019	Vitvi13g00031	Vitvi13g00053
2	2	2	2	2
Vitvi13g00095	Vitvi13g00096	Vitvi13g00111	Vitvi13g00149	Vitvi13g00288
2	2	2	2	2
Vitvi13g00328	Vitvi13g00332	Vitvi13g00334	Vitvi13g00344	Vitvi13g00510
2	2	2	2	2
Vitvi13g00627	Vitvi13g00628	Vitvi13g00636	Vitvi13g00637	Vitvi13g00700
2	2	2	2	2
Vitvi13g00744	Vitvi13g01070	Vitvi13g01077	Vitvi13g01233	Vitvi13g01272
2	2	2	2	2
Vitvi13g01327	Vitvi13g01364	Vitvi13g01839	Vitvi13g01854	Vitvi13g02272
2	2	2	2	2
Vitvi14g00020	Vitvi14g00025	Vitvi14g00053	Vitvi14g00160	Vitvi14g00185
2	2	2	2	2
Vitvi14g00237	Vitvi14g00281	Vitvi14g00327	Vitvi14g00380	Vitvi14g00383
2	2	2	2	2
Vitvi14g00397	Vitvi14g00404	Vitvi14g00439	Vitvi14g00484	Vitvi14g00535
2	2	2	2	2
Vitvi14g00562	Vitvi14g00627	Vitvi14g00654	Vitvi14g00658	Vitvi14g00763
2	2	2	2	2
Vitvi14g00841	Vitvi14g01061	Vitvi14g01128	Vitvi14g01280	Vitvi14g01294
2	2	2	2	2
Vitvi14g01321	Vitvi14g01472	Vitvi14g01520	Vitvi14g01596	Vitvi14g01678
2	2	2	2	2
Vitvi14g01746	Vitvi14g01758	Vitvi14g01809	Vitvi14g01983	Vitvi14g02002

2	2	2	2	2
Vitvi14g02027	Vitvi14g02478	Vitvi14g02644	Vitvi14g02869	Vitvi15g00225
2	2	2	2	2
Vitvi15g00526	Vitvi15g00563	Vitvi15g00690	Vitvi15g00827	Vitvi15g00840
2	2	2	2	2
Vitvi15g00878	Vitvi15g00908	Vitvi15g00974	Vitvi15g01077	Vitvi15g01403
2	2	2	2	2
Vitvi15g01468	Vitvi15g01491	Vitvi15g01537	Vitvi15g01539	Vitvi16g00018
2	2	2	2	2
Vitvi16g00048	Vitvi16g00095	Vitvi16g00461	Vitvi16g00796	Vitvi16g00857
2	2	2	2	2
Vitvi16g00935	Vitvi16g00949	Vitvi16g00982	Vitvi16g01025	Vitvi16g01083
2	2	2	2	2
Vitvi16g01089	Vitvi16g01211	Vitvi16g01323	Vitvi16g01326	Vitvi16g01348
2	2	2	2	2
Vitvi16g01358	Vitvi16g01371	Vitvi16g01375	Vitvi16g01391	Vitvi17g00038
2	2	2	2	2
Vitvi17g00161	Vitvi17g00210	Vitvi17g00222	Vitvi17g00225	Vitvi17g00240
2	2	2	2	2
Vitvi17g00253	Vitvi17g00263	Vitvi17g00277	Vitvi17g00287	Vitvi17g00299
2	2	2	2	2
Vitvi17g00428	Vitvi17g00468	Vitvi17g00519	Vitvi17g00548	Vitvi17g00603
2	2	2	2	2
Vitvi17g00614	Vitvi17g00869	Vitvi17g00905	Vitvi17g01148	Vitvi17g01183
2	2	2	2	2
Vitvi17g01348	Vitvi17g01399	Vitvi17g01407	Vitvi18g00056	Vitvi18g00132
2	2	2	2	2
Vitvi18g00177	Vitvi18g00234	Vitvi18g00310	Vitvi18g00358	Vitvi18g00444
2	2	2	2	2
Vitvi18g00507	Vitvi18g00517	Vitvi18g00534	Vitvi18g00579	Vitvi18g00743
2	2	2	2	2
Vitvi18g00762	Vitvi18g00784	Vitvi18g00838	Vitvi18g00909	Vitvi18g00960
2	2	2	2	2
Vitvi18g00966	Vitvi18g01002	Vitvi18g01004	Vitvi18g01019	Vitvi18g01095
2	2	2	2	2
Vitvi18g01103	Vitvi18g01223	Vitvi18g01226	Vitvi18g01242	Vitvi18g01299
2	2	2	2	2
Vitvi18g01309	Vitvi18g01430	Vitvi18g01436	Vitvi18g01634	Vitvi18g01677
2	2	2	2	2
Vitvi18g02066	Vitvi18g02228	Vitvi18g02237	Vitvi18g02378	Vitvi18g03244
2	2	2	2	2
Vitvi19g00015	Vitvi19g00020	Vitvi19g00094	Vitvi19g00138	Vitvi19g00155
2	2	2	2	2
Vitvi19g00197	Vitvi19g00226	Vitvi19g00337	Vitvi19g00339	Vitvi19g00341
2	2	2	2	2
Vitvi19g00353	Vitvi19g00421	Vitvi19g00531	Vitvi19g00539	Vitvi19g00552
2	2	2	2	2
Vitvi19g00602	Vitvi19g00612	Vitvi19g00621	Vitvi19g00675	Vitvi19g00706
2	2	2	2	2
Vitvi19g00745	Vitvi19g00782	Vitvi19g00904	Vitvi19g00928	Vitvi19g01352
2	2	2	2	2
Vitvi19g01464	Vitvi19g01552	Vitvi19g01690	Vitvi19g01692	Vitvi19g01725
2	2	2	2	2

```

Vitvi19g01864 Vitvi19g01933 Vitvi19g01937 Vitvi19g01940 Vitvi19g01952
                2                2                2                2                2
Vitvi19g01957 Vitvi19g01959 Vitvi19g01963 Vitvi19g01965 Vitvi19g01966
                2                2                2                2                2
Vitvi19g01969 Vitvi19g01970 Vitvi19g02001 Vitvi19g02292
                2                2                2                2

```

A glimpse into feature data (or NULL if nonexistent)

```
> fdata[1:6,]
```

```

YEAR      geneID BINCODE      NAME
1 2018 Vitvi15g01736    26.9 misc.glutathione S transferases
2 2018 Vitvi07g02832    35.2          not assigned.unknown
3 2018 Vitvi07g02830    35.2          not assigned.unknown
4 2018 Vitvi07g02812    35.2          not assigned.unknown
5 2018 Vitvi07g02811    35.2          not assigned.unknown
6 2018 Vitvi09g02033    35.2          not assigned.unknown

```

```

1          glutathione S-transferase tau 7 | Chr2:12618111-12618871
2 Disease resistance protein (TIR-NBS-LRR class) family | Chr4:7197325-7201393
3
4
5

```

```
6 Disease resistance protein (CC-NBS-LRR class) family | Chr1:4145374-4147680
```

```
> cat(gsub("\\| ", " |\\n\\|\\t", fdata$DESCRIPTION[1:6]), sep="\n\n")
```

```

glutathione S-transferase tau 7 |
      Chr2:12618111-12618871 REVERSE LENGTH=227 |
      201606

```

```

Disease resistance protein (TIR-NBS-LRR class) family |
      Chr4:7197325-7201393 REVERSE LENGTH=1219 |
      201606

```

NA

Tir-nbs-rrr, resistance protein

NA

```

Disease resistance protein (CC-NBS-LRR class) family |
      Chr1:4145374-4147680 FORWARD LENGTH=768 |
      201606

```

```
> fd <- (apply(fdata[1:6, c(2:5)], 1, paste, collapse="\n\t"))
```

```
> cat(gsub("\\| ", " |\\n\\|\\t", fd[1:6]), sep="\n\n")
```

```

Vitvi15g01736
      26.9
      misc.glutathione S transferases
      glutathione S-transferase tau 7 |
      Chr2:12618111-12618871 REVERSE LENGTH=227 |
      201606

```

```

Vitvi07g02832
      35.2

```

```
not assigned.unknown
Disease resistance protein (TIR-NBS-LRR class) family |
Chr4:7197325-7201393 REVERSE LENGTH=1219 |
201606
```

```
Vitvi07g02830
35.2
not assigned.unknown
NA
```

```
Vitvi07g02812
35.2
not assigned.unknown
Tir-nbs-lrr, resistance protein
```

```
Vitvi07g02811
35.2
not assigned.unknown
NA
```

```
Vitvi09g02033
35.2
not assigned.unknown
Disease resistance protein (CC-NBS-LRR class) family |
Chr1:4145374-4147680 FORWARD LENGTH=768 |
201606
```

```
> cat.feature <- function(x,...) {
+ #x <- t(apply(x,1, FUN=function(x) paste(names(x),x,sep=":\t") ))
+ #cat(x)
+ fd <- (apply(x,1,paste,collapse="\n\t"))
+ cat(gsub(" \\| ", " |\\n\\|\\t", fd), sep="\n\n")
+ }
```

```
> cat.feature(fdata[1,2:5])
```

```
Vitvi15g01736
26.9
misc.glutathione S transferases
glutathione S-transferase tau 7 |
Chr2:12618111-12618871 REVERSE LENGTH=227 |
201606
```

```
> cat.feature(fdata[1:2,2:5])
```

```
Vitvi15g01736
26.9
misc.glutathione S transferases
glutathione S-transferase tau 7 |
Chr2:12618111-12618871 REVERSE LENGTH=227 |
201606
```

```
Vitvi07g02832
35.2
not assigned.unknown
Disease resistance protein (TIR-NBS-LRR class) family |
Chr4:7197325-7201393 REVERSE LENGTH=1219 |
201606
```

3.5 Transcripts measurements

Normalised transcript measurements for each year are available in two separate files. The first few lines contain factor values and are commented out (by a hash). The first effective line is the table header line with sample ids.

3.5.1 Normalized values

Data for year 2018:

```
> (t18fn <- getDesc(.adesc, "Transcript data 18"))
[1] "/input/Transcripts 2018.txt"
> td <- read.table(file.path(.aroot, t18fn),
+   header = TRUE,
+   sep = "\t",
+   row.names = 1
+   )
> head(td)
```

	C18_11d_WS1	C18_11d_WS2	C18_11d_WS3	C18_11d_WS4
Vitvi15g01736	4.6646023	4.109897	4.4482203	4.013955
Vitvi07g02832	-2.4786650	-1.601359	-1.4274193	-2.066419
Vitvi07g02830	-1.8506337	-2.218030	-1.0648492	-1.580992
Vitvi07g02812	-0.5805446	-1.266940	-0.3278837	-1.019113
Vitvi07g02811	2.2762225	1.823447	1.9550503	1.648742
Vitvi09g02033	-1.6901691	-2.218030	-2.6498117	-1.449747
	C18_11d_WW1	C18_11d_WW2	C18_11d_WW3	C18_11d_WW4
Vitvi15g01736	4.717536	4.154237	4.475261	4.466465
Vitvi07g02832	-2.608485	-1.801183	-1.886474	-3.583384
Vitvi07g02830	-1.544354	-1.405255	-1.755229	-1.381750
Vitvi07g02812	-0.858463	-0.434401	-1.234397	-1.047331
Vitvi07g02811	2.086902	2.105707	2.134837	1.908469
Vitvi09g02033	-2.608485	-1.961648	-3.108866	-4.320350
	C18_34d_WS1	C18_34d_WS2	C18_34d_WS3	C18_34d_WS4
Vitvi15g01736	4.400333	4.641164	4.4191231	4.534438
Vitvi07g02832	-5.683146	-2.452000	-4.4274120	-1.876949
Vitvi07g02830	-2.513221	-2.452000	-4.4274120	-2.263972
Vitvi07g02812	-2.875791	-2.693008	-2.5529429	-3.157057
Vitvi07g02811	1.414886	1.409803	0.3970164	1.273993
Vitvi09g02033	-2.513221	-3.345084	-2.1054839	-1.716484
	C18_34d_WW1	C18_34d_WW2	C18_34d_WW3	C18_34d_WW4
Vitvi15g01736	4.924801	5.085690	5.3397350	5.263629
Vitvi07g02832	-2.827859	-3.130016	-4.6322874	-3.196388
Vitvi07g02830	-3.190429	-1.545054	-2.1297871	-3.196388
Vitvi07g02812	-2.827859	-1.182484	-3.4098950	-2.303303
Vitvi07g02811	1.517916	1.357250	0.9626591	1.344985
Vitvi09g02033	-4.412821	-3.130016	-2.3103594	-4.418780
	C18_67d_WS1	C18_67d_WS2	C18_67d_WS3	C18_67d_WS4
Vitvi15g01736	2.05871733	2.2895026	2.9234830	2.575657
Vitvi07g02832	-5.85417200	-4.4786818	-4.4659691	-2.468738
Vitvi07g02830	-2.15373228	-3.2562893	-3.7290035	-2.709746
Vitvi07g02812	-1.21031581	-2.3632045	-2.3504919	-2.999252
Vitvi07g02811	0.02847105	0.5362686	0.9824914	1.231702
Vitvi09g02033	-4.26920950	-3.7417162	-2.5915000	-3.847249

	C18_67d_WW1	C18_67d_WW2	C18_67d_WW3	C18_67d_WW4
Vitvi15g01736	3.891121	3.040087	4.216897	3.763086
Vitvi07g02832	-2.726982	-4.610965	-2.436913	-3.355855
Vitvi07g02830	-2.726982	-3.873999	-1.493496	-2.075747
Vitvi07g02812	-1.672534	-3.388572	-2.230462	-2.462770
Vitvi07g02811	1.436764	1.552266	1.811015	1.352490
Vitvi09g02033	-2.520531	-2.736495	-3.815424	-2.993285
F18_10d_WS1	F18_10d_WS2	F18_10d_WS3	F18_10d_WS4	
Vitvi15g01736	4.8591409	4.8669435	4.9805904	4.9453953
Vitvi07g02832	2.8881443	2.7032394	2.2757651	2.0553154
Vitvi07g02830	2.8426117	2.4130023	2.2135368	2.0169691
Vitvi07g02812	0.8222302	0.9363895	0.7209384	0.3104447
Vitvi07g02811	3.5853480	3.5007015	3.2965509	3.2141610
Vitvi09g02033	1.0793880	1.0850336	0.6285743	0.8579324
F18_10d_WW1	F18_10d_WW2	F18_10d_WW3	F18_10d_WW4	
Vitvi15g01736	5.705006	4.619238	4.808548	5.276124
Vitvi07g02832	2.747538	3.349854	2.537311	2.621336
Vitvi07g02830	2.287722	3.464855	1.989824	2.321031
Vitvi07g02812	1.073767	1.384521	1.472076	1.016877
Vitvi07g02811	3.818074	4.000053	3.198197	3.501954
Vitvi09g02033	1.073767	1.566041	1.237985	1.268416
F18_34d_WS1	F18_34d_WS2	F18_34d_WS3	F18_34d_WS4	
Vitvi15g01736	4.0647184	3.8504580	5.1418322	4.6269091
Vitvi07g02832	2.2592790	2.1761360	2.3686660	1.2127920
Vitvi07g02830	2.1186455	1.9359657	2.6356368	1.5129655
Vitvi07g02812	0.4316833	-0.6220298	-0.5985972	-0.2055206
Vitvi07g02811	2.9490282	2.7595613	3.1300210	2.4602029
Vitvi09g02033	1.5459405	1.3699329	1.4288835	1.4477479
F18_34d_WW1	F18_34d_WW2	F18_34d_WW3	F18_34d_WW4	
Vitvi15g01736	4.0961725	4.80222138	4.3291524	4.4419027
Vitvi07g02832	1.8816574	1.82887526	1.7895023	1.6490200
Vitvi07g02830	2.1414534	1.62149541	2.1153419	2.1541121
Vitvi07g02812	0.1031253	-0.09471163	0.3053805	0.2855497
Vitvi07g02811	3.0025984	2.73213947	2.8750439	3.0458304
Vitvi09g02033	1.7460934	1.45938991	2.0072243	1.2444445
F18_67d_WS1	F18_67d_WS2	F18_67d_WS3	F18_67d_WS4	
Vitvi15g01736	1.5831276	1.7408584	2.97840555	4.1664059
Vitvi07g02832	1.3093668	1.9241330	2.01456521	2.9813855
Vitvi07g02830	0.8382242	1.4203666	2.18501915	2.8731326
Vitvi07g02812	0.1974739	0.4558154	-0.03878421	-0.0111843
Vitvi07g02811	1.8823168	2.5341865	2.65708760	2.9461961
Vitvi09g02033	1.6066824	1.5040245	1.25520700	0.9986635
F18_67d_WW1	F18_67d_WW2	F18_67d_WW3	F18_67d_WW4	
Vitvi15g01736	3.5580430	3.569471	4.4112089	4.3597440
Vitvi07g02832	2.5563386	2.180583	2.6637430	2.7538173
Vitvi07g02830	2.0265816	2.154972	2.7552617	2.3663511
Vitvi07g02812	0.6773148	0.106805	-0.2413317	-0.1063251
Vitvi07g02811	3.0678390	2.949349	2.9761917	2.9625700
Vitvi09g02033	1.7205680	1.541742	1.6527997	1.3320450

> dim(td)

[1] 16005 48

> t18 <- td

```
> (t19fn <- getDesc(.adesc, "Transcript data 19"))
```

```
[1] "/input/Transcripts 2019.txt"
```

```
> td <- read.table(file.path(.aroot, t19fn),  
+ header = TRUE,  
+ sep = "\t",  
+ row.names = 1  
+ )  
> head(td)
```

	C19_22d_WS1	C19_22d_WS2	C19_22d_WS3	C19_22d_WS4
Vitvi15g01736	3.18	3.50	3.19	3.61
Vitvi07g02832	-5.05	-3.02	-0.80	-0.02
Vitvi07g02830	-5.05	-5.35	-2.39	-3.99
Vitvi07g02812	-1.35	-0.95	-0.15	0.26
Vitvi07g02811	2.09	2.29	2.34	2.63
Vitvi09g02033	-5.05	-3.76	-2.39	-2.76
	C19_22d_WW1	C19_22d_WW2	C19_22d_WW3	C19_22d_WW4
Vitvi15g01736	5.42	5.35	4.68	4.92
Vitvi07g02832	-1.36	-2.10	-1.15	-3.81
Vitvi07g02830	-2.28	-1.81	-2.07	-2.59
Vitvi07g02812	-1.36	0.16	-0.29	-0.75
Vitvi07g02811	2.85	3.16	3.09	2.89
Vitvi09g02033	-5.45	-3.68	-1.78	-3.81
	C19_44d_WS1	C19_44d_WS2	C19_44d_WS3	C19_44d_WS4
Vitvi15g01736	5.13	5.19	5.71	5.29
Vitvi07g02832	-3.13	-1.28	-3.18	-2.29
Vitvi07g02830	-3.87	-5.53	-5.50	-2.29
Vitvi07g02812	-0.10	-1.01	-0.29	-0.04
Vitvi07g02811	2.11	1.92	2.40	2.29
Vitvi09g02033	-3.87	-3.95	-2.69	-2.00
	C19_44d_WW1	C19_44d_WW2	C19_44d_WW3	C19_44d_WW4
Vitvi15g01736	6.19	5.62	5.62	6.19
Vitvi07g02832	-5.54	-5.31	-3.73	-1.81
Vitvi07g02830	-3.22	-5.31	-3.73	-2.05
Vitvi07g02812	-0.59	-0.45	-1.07	-0.55
Vitvi07g02811	2.44	2.13	2.08	2.26
Vitvi09g02033	-3.22	-3.72	-3.73	-3.18
	F19_22d_WS1	F19_22d_WS2	F19_22d_WS3	F19_22d_WS4
Vitvi15g01736	3.03	3.93	4.24	2.85
Vitvi07g02832	3.35	3.23	3.02	2.98
Vitvi07g02830	2.28	2.39	1.75	1.35
Vitvi07g02812	2.19	1.04	0.89	1.56
Vitvi07g02811	3.84	3.72	3.01	3.43
Vitvi09g02033	2.70	1.61	1.94	2.47
	F19_22d_WW1	F19_22d_WW2	F19_22d_WW3	F19_22d_WW4
Vitvi15g01736	5.00	4.97	4.47	5.51
Vitvi07g02832	3.18	3.04	2.81	3.05
Vitvi07g02830	2.59	2.07	1.81	2.20
Vitvi07g02812	1.72	1.08	1.42	1.35
Vitvi07g02811	3.79	3.43	3.24	3.33
Vitvi09g02033	2.24	1.90	2.08	1.25
	F19_44d_WS1	F19_44d_WS2	F19_44d_WS3	F19_44d_WS4
Vitvi15g01736	5.84	5.83	4.56	5.23

Vitvi07g02832	2.54	3.14	3.01	2.16
Vitvi07g02830	1.13	2.25	2.18	1.90
Vitvi07g02812	0.99	0.84	0.85	0.22
Vitvi07g02811	2.91	3.10	3.18	2.61
Vitvi09g02033	1.21	1.67	2.52	2.21
	F19_44d_WW1	F19_44d_WW2	F19_44d_WW3	F19_44d_WW4
Vitvi15g01736	5.44	6.13	5.96	6.16
Vitvi07g02832	3.84	3.05	3.89	4.02
Vitvi07g02830	2.74	2.79	2.90	2.88
Vitvi07g02812	1.03	0.93	1.49	1.29
Vitvi07g02811	3.32	2.89	3.35	3.26
Vitvi09g02033	1.99	1.14	1.65	1.29

```
> dim(td)
```

```
[1] 15562 32
```

```
> t19 <- td
```

```
> dim(t18)-dim(t19)
```

```
[1] 443 16
```

Data for year 2018 have more genes than data for year 2019.

```
> allNames <- unique(c(rownames(t18), rownames(t19)))
```

```
> length(allNames)
```

```
[1] 16325
```

```
> sum(allNames %in% rownames(t18))
```

```
[1] 16005
```

```
> sum(allNames %in% rownames(t19))
```

```
[1] 15562
```

```
> tbl <- table(gene.in.18=allNames %in% rownames(t18),
```

```
+ gene.in.19=allNames %in% rownames(t19))
```

```
> tbl
```

	gene.in.19	
gene.in.18	FALSE	TRUE
FALSE	0	320
TRUE	763	15242

We have complete yearly data on 15242 genes. At the meeting on Nov. 19 we decided to use only the genes in the intersection. This will allow us to use the combined set of data.

3.5.2 Keep genes in the intersection

```
> gene.keep <- (allNames %in% rownames(t18)) & (allNames %in% rownames(t19))
> sum(gene.keep)
[1] 15242
> t18 <- t18[allNames[gene.keep],]
> t19 <- t19[allNames[gene.keep],]
> all(rownames(t18) == rownames(t19))
[1] TRUE
> dim(t18)
[1] 15242    48
> dim(t19)
[1] 15242    32
```

Order rows of yearly phenodata according to the sequence of samples (columns) in the measurements.

Year 2018

```
> dim(pdata18)
[1] 136  22
> dim(t18)
[1] 15242    48
> pdata18 <- pdata18[colnames(t18),]
> dim(pdata18)
[1] 48 22
> dim(t18)
[1] 15242    48
> pdata18[,4:8]
```

	variety	year	day	treat	rep
C18_11d_WS1	C	18	11	WS	1
C18_11d_WS2	C	18	11	WS	2
C18_11d_WS3	C	18	11	WS	3
C18_11d_WS4	C	18	11	WS	4
C18_11d_WW1	C	18	11	WW	1
C18_11d_WW2	C	18	11	WW	2
C18_11d_WW3	C	18	11	WW	3
C18_11d_WW4	C	18	11	WW	4
C18_34d_WS1	C	18	34	WS	1
C18_34d_WS2	C	18	34	WS	2
C18_34d_WS3	C	18	34	WS	3
C18_34d_WS4	C	18	34	WS	4
C18_34d_WW1	C	18	34	WW	1
C18_34d_WW2	C	18	34	WW	2
C18_34d_WW3	C	18	34	WW	3
C18_34d_WW4	C	18	34	WW	4
C18_67d_WS1	C	18	67	WS	1
C18_67d_WS2	C	18	67	WS	2
C18_67d_WS3	C	18	67	WS	3
C18_67d_WS4	C	18	67	WS	4
C18_67d_WW1	C	18	67	WW	1
C18_67d_WW2	C	18	67	WW	2

```

C18_67d_WW3      C    18  67   WW    3
C18_67d_WW4      C    18  67   WW    4
F18_10d_WS1      F    18  11   WS    1
F18_10d_WS2      F    18  11   WS    2
F18_10d_WS3      F    18  11   WS    3
F18_10d_WS4      F    18  11   WS    4
F18_10d_WW1      F    18  11   WW    1
F18_10d_WW2      F    18  11   WW    2
F18_10d_WW3      F    18  11   WW    3
F18_10d_WW4      F    18  11   WW    4
F18_34d_WS1      F    18  34   WS    1
F18_34d_WS2      F    18  34   WS    2
F18_34d_WS3      F    18  34   WS    3
F18_34d_WS4      F    18  34   WS    4
F18_34d_WW1      F    18  34   WW    1
F18_34d_WW2      F    18  34   WW    2
F18_34d_WW3      F    18  34   WW    3
F18_34d_WW4      F    18  34   WW    4
F18_67d_WS1      F    18  67   WS    1
F18_67d_WS2      F    18  67   WS    2
F18_67d_WS3      F    18  67   WS    3
F18_67d_WS4      F    18  67   WS    4
F18_67d_WW1      F    18  67   WW    1
F18_67d_WW2      F    18  67   WW    2
F18_67d_WW3      F    18  67   WW    3
F18_67d_WW4      F    18  67   WW    4

```

```
> all(rownames(pdata18)==colnames(t18))
```

```
[1] TRUE
```

Get rid of unused factors

```
> str(pdata18[, factors])
```

```
'data.frame':      48 obs. of  5 variables:
 $ variety: Factor w/ 2 levels "C","F": 1 1 1 1 1 1 1 1 1 1 ...
 $ year   : Factor w/ 2 levels "18","19": 1 1 1 1 1 1 1 1 1 1 ...
 $ day    : Factor w/ 26 levels "3","11","14",...: 2 2 2 2 2 2 2 2 8 8 ...
 $ treat  : Factor w/ 2 levels "WW","WS": 2 2 2 2 1 1 1 1 2 2 ...
 $ rep    : Factor w/ 4 levels "1","2","3","4": 1 2 3 4 1 2 3 4 1 2 ...
```

```
> for (var in factors) pdata18[,var] <- factor(pdata18[,var])
```

```
> str(pdata18[, factors])
```

```
'data.frame':      48 obs. of  5 variables:
 $ variety: Factor w/ 2 levels "C","F": 1 1 1 1 1 1 1 1 1 1 ...
 $ year   : Factor w/ 1 level "18": 1 1 1 1 1 1 1 1 1 1 ...
 $ day    : Factor w/ 3 levels "11","34","67": 1 1 1 1 1 1 1 1 2 2 ...
 $ treat  : Factor w/ 2 levels "WW","WS": 2 2 2 2 1 1 1 1 2 2 ...
 $ rep    : Factor w/ 4 levels "1","2","3","4": 1 2 3 4 1 2 3 4 1 2 ...
```

Year 2019

```
> dim(pdata19)
```

```
[1] 134  22
```

```
> dim(t19)
```

```
[1] 15242  32
```

```

> pdata19 <- pdata19[colnames(t19),]
> dim(pdata19)
[1] 32 22
> dim(t19)
[1] 15242 32
> pdata19[,4:9]
      variety year day treat rep project.name
C19_22d_WS1      C   19  22   WS    1     EnViros
C19_22d_WS2      C   19  22   WS    2     EnViros
C19_22d_WS3      C   19  22   WS    3     EnViros
C19_22d_WS4      C   19  22   WS    4     EnViros
C19_22d_WW1      C   19  22   WW    1     EnViros
C19_22d_WW2      C   19  22   WW    2     EnViros
C19_22d_WW3      C   19  22   WW    3     EnViros
C19_22d_WW4      C   19  22   WW    4     EnViros
C19_44d_WS1      C   19  44   WS    1     EnViros
C19_44d_WS2      C   19  44   WS    2     EnViros
C19_44d_WS3      C   19  44   WS    3     EnViros
C19_44d_WS4      C   19  44   WS    4     EnViros
C19_44d_WW1      C   19  44   WW    1     EnViros
C19_44d_WW2      C   19  44   WW    2     EnViros
C19_44d_WW3      C   19  44   WW    3     EnViros
C19_44d_WW4      C   19  44   WW    4     EnViros
F19_22d_WS1      F   19  22   WS    1     EnViros
F19_22d_WS2      F   19  22   WS    2     EnViros
F19_22d_WS3      F   19  22   WS    3     EnViros
F19_22d_WS4      F   19  22   WS    4     EnViros
F19_22d_WW1      F   19  22   WW    1     EnViros
F19_22d_WW2      F   19  22   WW    2     EnViros
F19_22d_WW3      F   19  22   WW    3     EnViros
F19_22d_WW4      F   19  22   WW    4     EnViros
F19_44d_WS1      F   19  44   WS    1     EnViros
F19_44d_WS2      F   19  44   WS    2     EnViros
F19_44d_WS3      F   19  44   WS    3     EnViros
F19_44d_WS4      F   19  44   WS    4     EnViros
F19_44d_WW1      F   19  44   WW    1     EnViros
F19_44d_WW2      F   19  44   WW    2     EnViros
F19_44d_WW3      F   19  44   WW    3     EnViros
F19_44d_WW4      F   19  44   WW    4     EnViros

```

```

> all(rownames(pdata19)==colnames(t19))
[1] TRUE

```

Get rid of unused factors levels

```

> str(pdata19[,factors])
'data.frame':      32 obs. of  5 variables:
 $ variety: Factor w/ 2 levels "C","F": 1 1 1 1 1 1 1 1 1 1 ...
 $ year   : Factor w/ 2 levels "18","19": 2 2 2 2 2 2 2 2 2 2 ...
 $ day    : Factor w/ 26 levels "3","11","14",...: 5 5 5 5 5 5 5 5 12 12 ...
 $ treat  : Factor w/ 2 levels "WW","WS": 2 2 2 2 1 1 1 1 2 2 ...
 $ rep    : Factor w/ 4 levels "1","2","3","4": 1 2 3 4 1 2 3 4 1 2 ...

```

```

> for (var in factors) pdata19[,var] <- factor(pdata19[,var])
> str(pdata19[,factors])

'data.frame':      32 obs. of  5 variables:
 $ variety: Factor w/ 2 levels "C","F": 1 1 1 1 1 1 1 1 1 1 ...
 $ year   : Factor w/ 1 level "19": 1 1 1 1 1 1 1 1 1 1 ...
 $ day    : Factor w/ 2 levels "22","44": 1 1 1 1 1 1 1 1 2 2 ...
 $ treat  : Factor w/ 2 levels "WW","WS": 2 2 2 2 1 1 1 1 2 2 ...
 $ rep    : Factor w/ 4 levels "1","2","3","4": 1 2 3 4 1 2 3 4 1 2 ...

> t1819 <- data.frame(t18, t19)
> pdata1819 <- rbind(pdata18, pdata19)
> str(pdata1819[,factors])

'data.frame':      80 obs. of  5 variables:
 $ variety: Factor w/ 2 levels "C","F": 1 1 1 1 1 1 1 1 1 1 ...
 $ year   : Factor w/ 2 levels "18","19": 1 1 1 1 1 1 1 1 1 1 ...
 $ day    : Factor w/ 5 levels "11","34","67",...: 1 1 1 1 1 1 1 1 2 2 ...
 $ treat  : Factor w/ 2 levels "WW","WS": 2 2 2 2 1 1 1 1 2 2 ...
 $ rep    : Factor w/ 4 levels "1","2","3","4": 1 2 3 4 1 2 3 4 1 2 ...

> catln("Sample names ",
+ c("different", "OK")[1+all(rownames(pdata1819)==colnames(t1819))])
Sample names  OK

```

3.6 Metabolite measurements

Normalised transcript measurements for each year are available in two separate files. The first few lines contains variable names

```
> (mfn18 <- getMeta(.adesc, "Metabolite data 18"))
[1] "/input/Metabolites_2018_raw_data.txt"
> (mfn19 <- getMeta(.adesc, "Metabolite data 19"))
[1] "/input/Metabolites_2019_raw_data.txt"
```

Read the metabolomics data

```
> m18 <- read.table(file.path(.aroot, mfn18), sep="\t", header=TRUE)
> dim(m18)
[1] 96 60
> m19 <- read.table(file.path(.aroot, mfn19), sep="\t", header=TRUE)
> dim(m19)
[1] 96 60
```

Check consistency of column names

```
> ok <- all(colnames(m18)==colnames(m19))
> if(ok) {catln("Column names match")} else {
+   catln("Column names do not match")}
```

Column names match

```
> cbind(names18=colnames(m18), names19=colnames(m19))
      names18                names19
[1,] "ID"                    "ID"
[2,] "Variety"                "Variety"
[3,] "Date"                   "Date"
[4,] "Treat"                  "Treat"
[5,] "Rep"                    "Rep"
[6,] "Alanine"                "Alanine"
[7,] "Arabinose"              "Arabinose"
[8,] "Ascorbic.acid"          "Ascorbic.acid"
[9,] "Aspartic.acid"          "Aspartic.acid"
[10,] "GABA"                   "GABA"
[11,] "Trans.Caffeic.acid"     "Trans.Caffeic.acid"
[12,] "Catechin"              "Catechin"
[13,] "Citric.acid"           "Citric.acid"
[14,] "Erythronic.acid"       "Erythronic.acid"
[15,] "Ethanolamine"          "Ethanolamine"
[16,] "Fructose"              "Fructose"
[17,] "Fructose.6.phosphate"   "Fructose.6.phosphate"
[18,] "Fumaric.acid"          "Fumaric.acid"
[19,] "Galactinol"            "Galactinol"
[20,] "Galactose"             "Galactose"
[21,] "Gallic.acid"           "Gallic.acid"
[22,] "Gluconic.acid"         "Gluconic.acid"
[23,] "Glucopyranose..H2O."    "Glucopyranose..H2O."
[24,] "Glucose"               "Glucose"
[25,] "Glutamic.acid"         "Glutamic.acid"
[26,] "Glyceric.acid"         "Glyceric.acid"
```

```

[27,] "Glycine"           "Glycine"
[28,] "Hydroquinone"    "Hydroquinone"
[29,] "Myo.Inostol"     "Myo.Inostol"
[30,] "Isoleucine"     "Isoleucine"
[31,] "Leucine"        "Leucine"
[32,] "Lyxonic.acid"   "Lyxonic.acid"
[33,] "Maleic.acid"    "Maleic.acid"
[34,] "Malic.acid"     "Malic.acid"
[35,] "Malonic.acid"   "Malonic.acid"
[36,] "Mannose.6.phosphate" "Mannose.6.phosphate"
[37,] "Melibiose"      "Melibiose"
[38,] "Phenylalanine"  "Phenylalanine"
[39,] "Phosphoric.acid" "Phosphoric.acid"
[40,] "Proline"        "Proline"
[41,] "Putrescine"     "Putrescine"
[42,] "Pyroglutamic.acid" "Pyroglutamic.acid"
[43,] "Quinic.acid"   "Quinic.acid"
[44,] "X3.caffeoylquinic.acid" "X3.caffeoylquinic.acid"
[45,] "Raffinose"     "Raffinose"
[46,] "Rhamnose"      "Rhamnose"
[47,] "Ribonic.acid"  "Ribonic.acid"
[48,] "Ribose"        "Ribose"
[49,] "Serine"        "Serine"
[50,] "Shikimic.acid" "Shikimic.acid"
[51,] "Succinic.acid" "Succinic.acid"
[52,] "Sucrose"       "Sucrose"
[53,] "Tartaric.acid" "Tartaric.acid"
[54,] "Threitol"      "Threitol"
[55,] "Threonic.acid" "Threonic.acid"
[56,] "Threonolactone" "Threonolactone"
[57,] "Threonine"     "Threonine"
[58,] "Uracil"        "Uracil"
[59,] "Valine"        "Valine"
[60,] "Xylose"        "Xylose"

```

Use first column as row names

```

> rownames(m18) <- m18[,1]
> rownames(m19) <- m19[,1]

```

Delete samples with missing values

```

> x <- m18
> num_id <- 6:ncol(x)
> dim(x)
[1] 96 60
> x <- x[apply(!is.na(x[,num_id]),1,all),]
> dim(x)
[1] 96 60
> m18 <- x

```

```

> x <- m19
> num_id <- 6:ncol(x)
> dim(x)
[1] 96 60
> x <- x[apply(!is.na(x[, num_id]), 1, all), ]
> dim(x)
[1] 94 60
> m19 <- x

```

Both years together

```

> m1819 <- rbind(m18, m19)
> dim(m1819)
[1] 190 60
> colnames(m1819)[1:7]
[1] "ID" "Variety" "Date" "Treat" "Rep"
[6] "Alanine" "Arabinose"
> rownames(m1819) <- m1819[, 1]
> head(rownames(m1819))
[1] "C18_11d_WS1" "C18_11d_WS2" "C18_11d_WS3" "C18_11d_WS4"
[5] "C18_11d_WW1" "C18_11d_WW2"
> length(rownames(m1819))
[1] 190

```

At some point, we will not need the first five descriptive columns.
Intersection of transcriptomics and metabolomics samples

```

> dim(m1819)
[1] 190 60
> dim(t1819)
[1] 15242 80
> head(colnames(t1819))
[1] "C18_11d_WS1" "C18_11d_WS2" "C18_11d_WS3" "C18_11d_WS4"
[5] "C18_11d_WW1" "C18_11d_WW2"
> head(rownames(m1819))
[1] "C18_11d_WS1" "C18_11d_WS2" "C18_11d_WS3" "C18_11d_WS4"
[5] "C18_11d_WW1" "C18_11d_WW2"
> sum(colnames(t1819) %in% rownames(m1819))
[1] 80
> sum(rownames(m1819) %in% colnames(t1819))
[1] 80
> all(sort(colnames(t1819)[(colnames(t1819) %in% rownames(m1819))]) ==
+ sort(rownames(m1819)[(rownames(m1819) %in% colnames(t1819))]))
[1] TRUE

> m_num_id <- 6:ncol(m1819)
> colnames(m1819)[-m_num_id]
[1] "ID" "Variety" "Date" "Treat" "Rep"

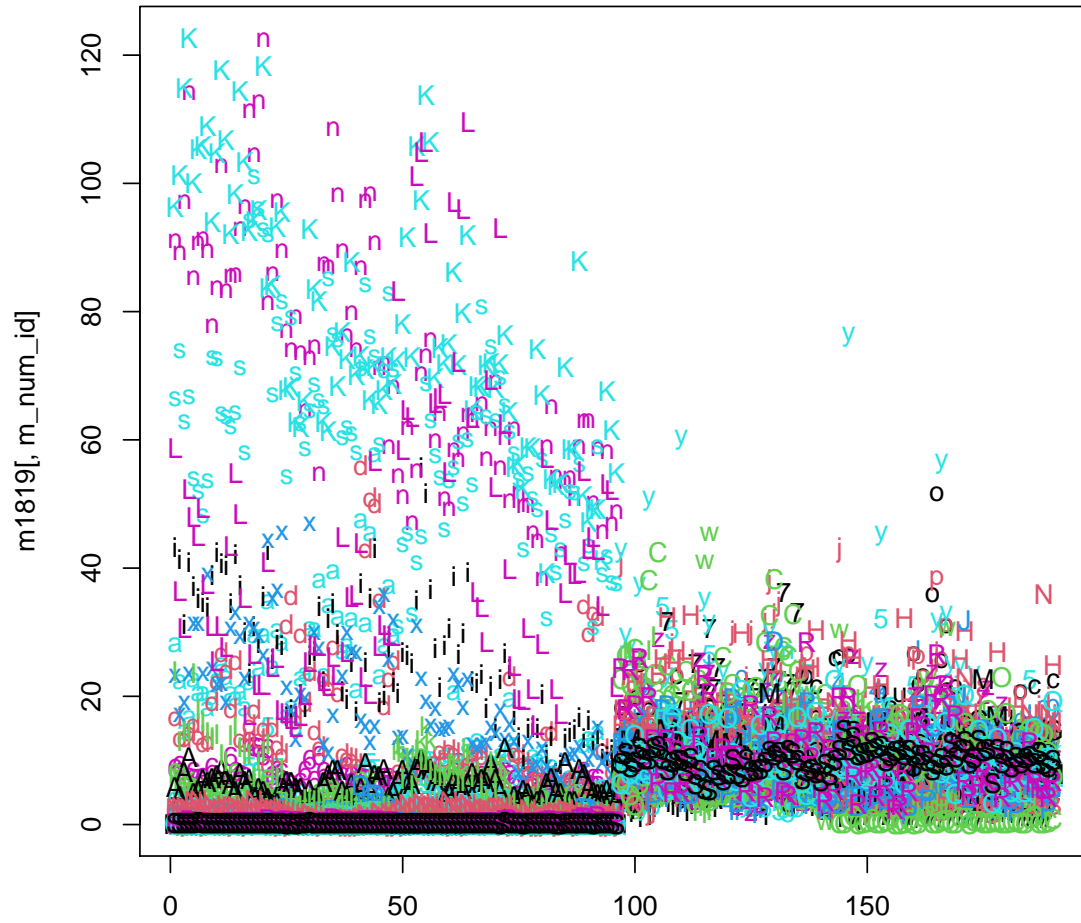
```

We end up with 80 samples

Phenodata for metabolomics data will be prepared when needed from the total phenodata. Selection of needed metabolomics data will be finalized in data analysis scripts.

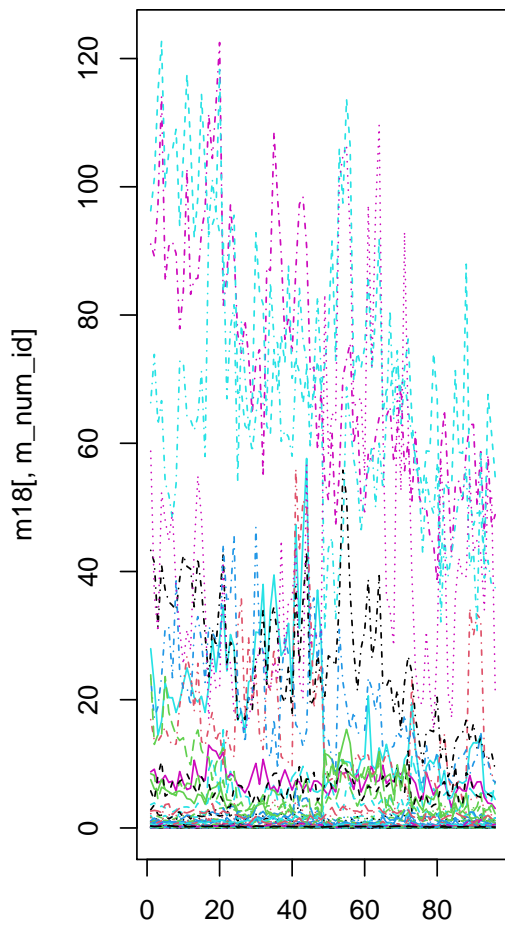
3.7 Visualize metabolites

```
> matplot(m1819[,m_num_id])
```

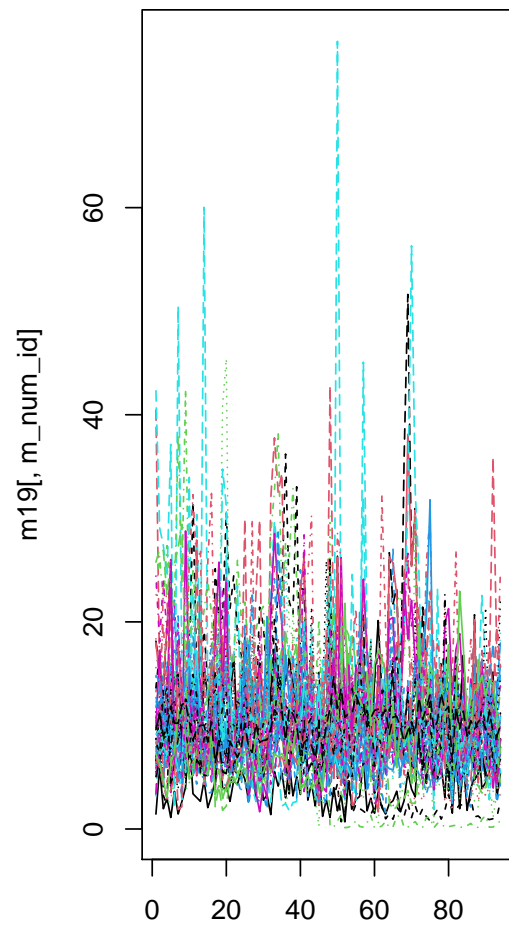


```
> par(mfrow=c(1,2))  
> matplot(m18[,m_num_id], main="2018", type="l")  
> matplot(m19[,m_num_id], main="2019", type="l")
```


2018



2019



Median standardization of samples

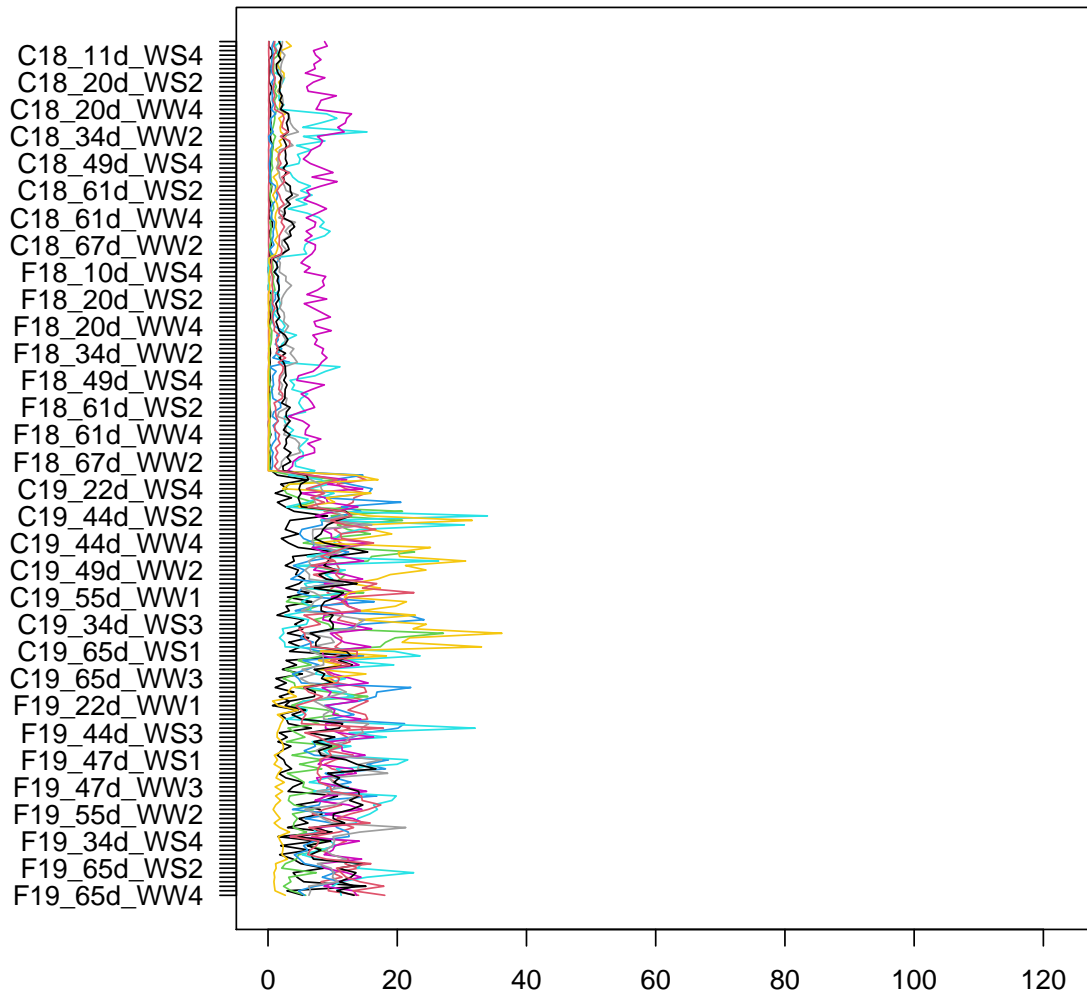
```
> my.parallel <- function(x, m=1:ncol(x), ...){  
+ m <- m[m<ncol(x)]  
+ par(mar=c(4, 8, 3, 1))  
+ xlim <- range(x, na.rm=TRUE)  
+ n1 <- nrow(x)  
+ plot(0, 0, xlim=xlim, ylim=c(n1, 1), type="n", axes=FALSE, ann=FALSE)  
+ title(paste(range(m), collapse=":"))  
+ axis(1)  
+ axis(2, at=1:n1, labels=rownames(x), las=2)  
+ box()  
+ for(i in m)  
+   lines(x[,i], 1:n1, col=i)  
+  
+ }  
> #  
> #my.parallel(x)
```

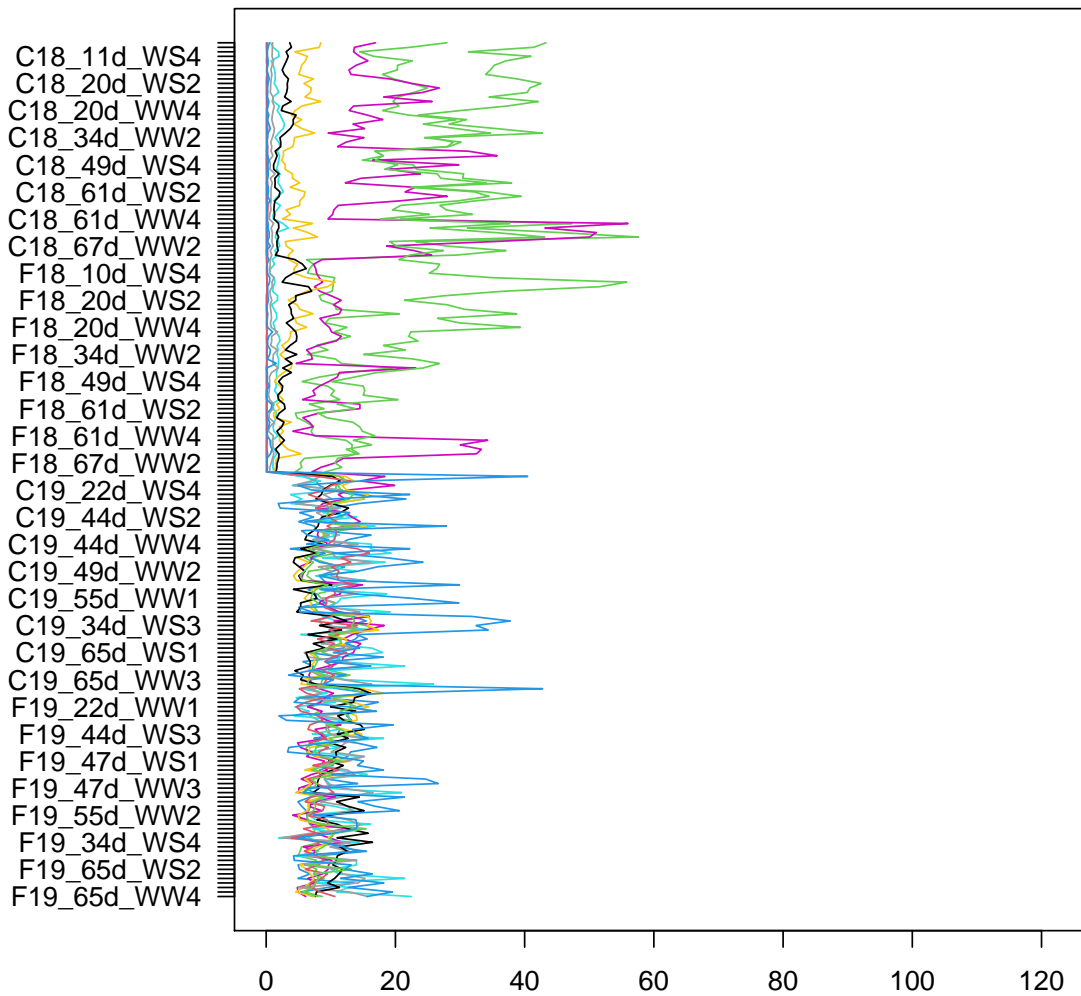
```

> x <- m1819[,m_num_id]
> for(k in seq(1,ncol(x),10)) {
+ if(interactive()) windows()
+   my.parallel( x, k:(k+9))
+ }

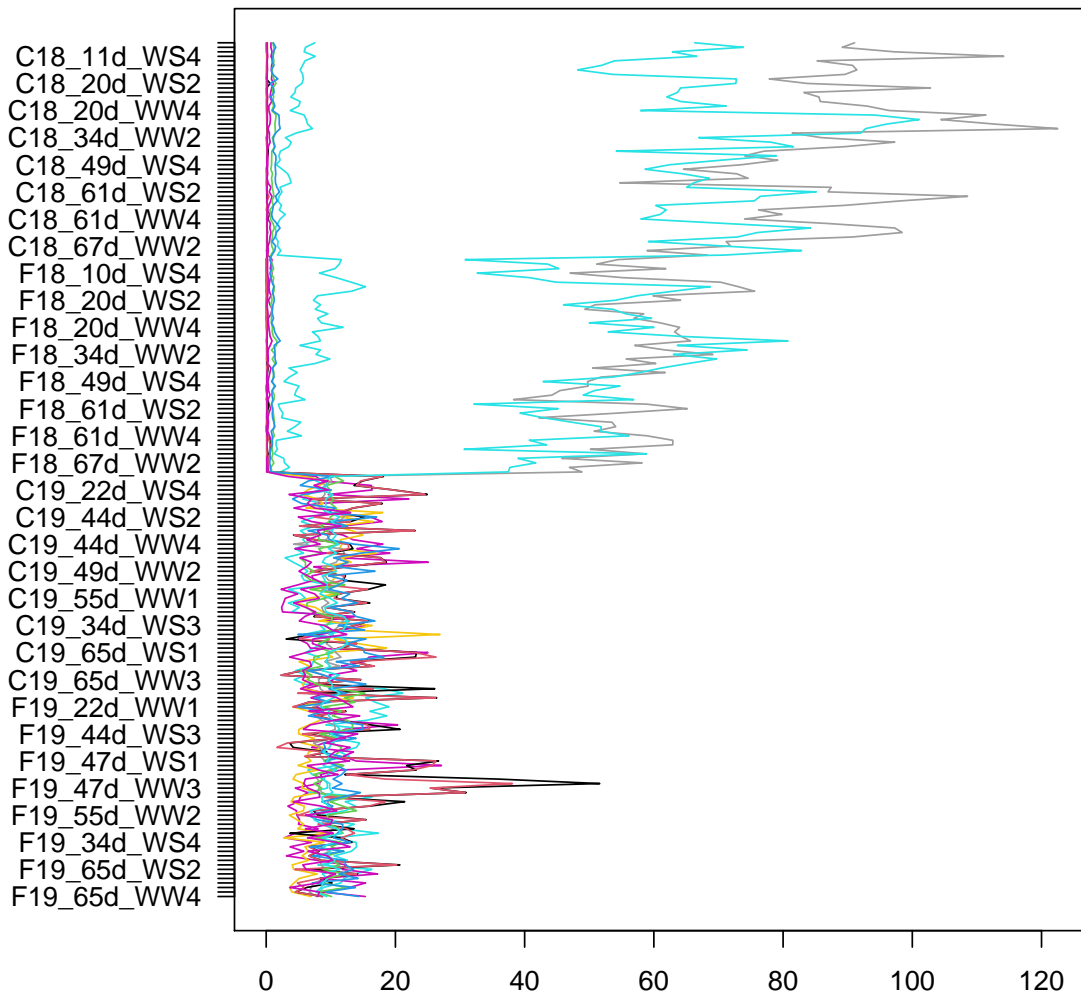
```

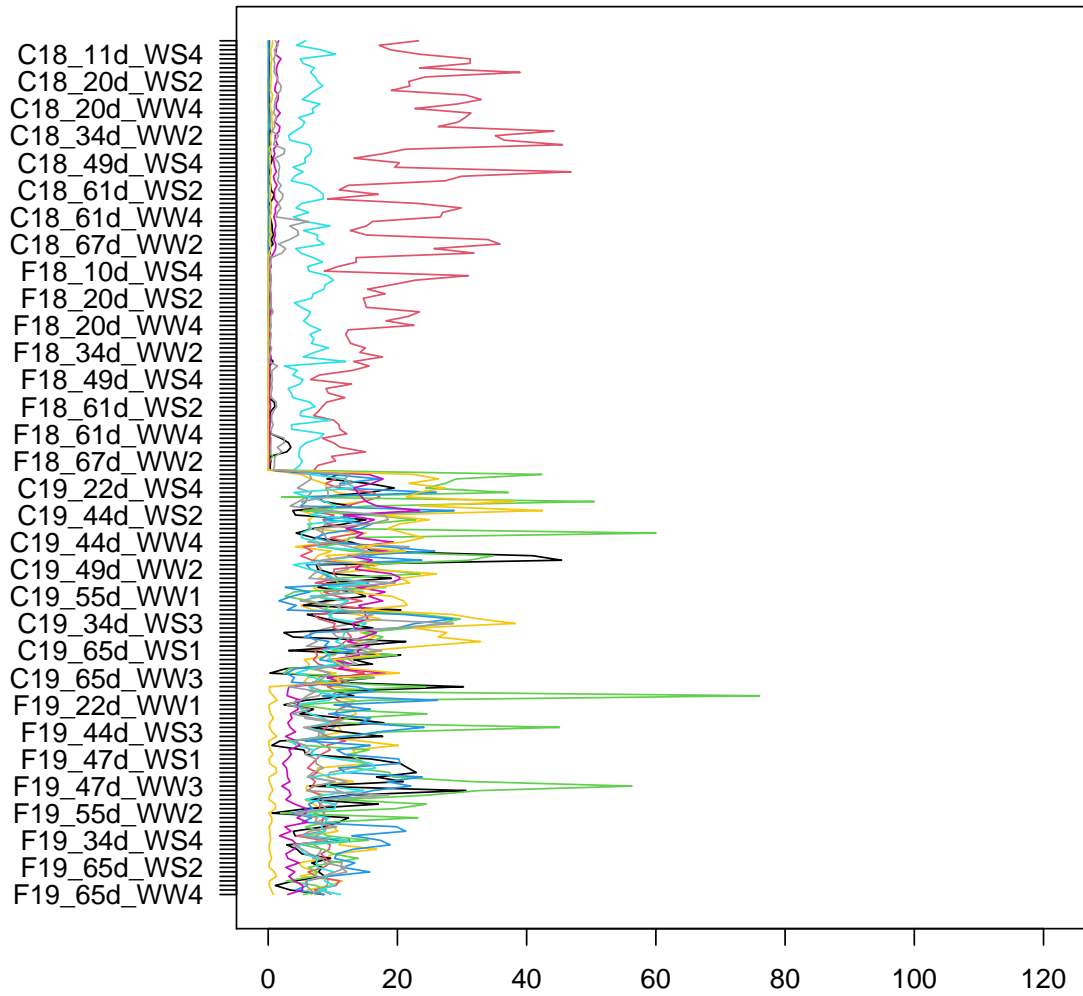
1:10

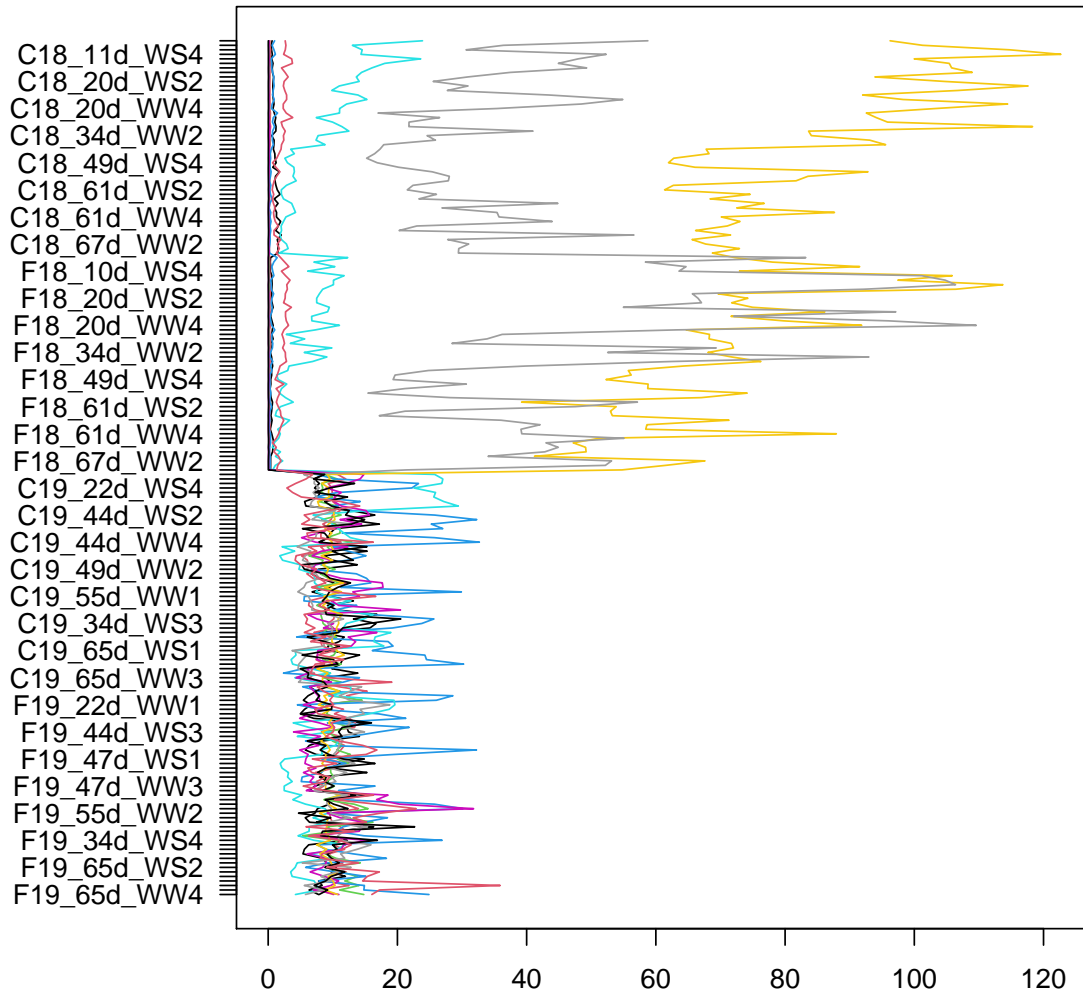


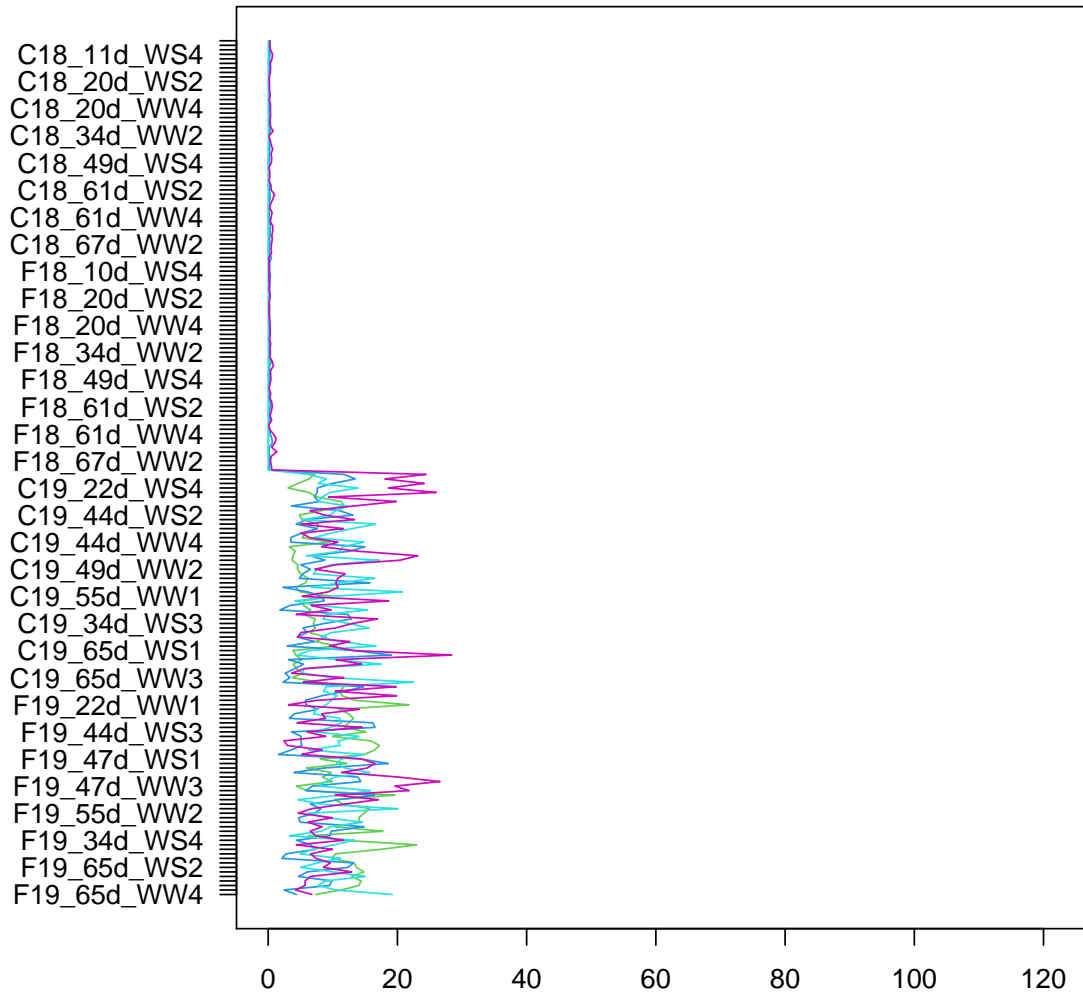


21:30





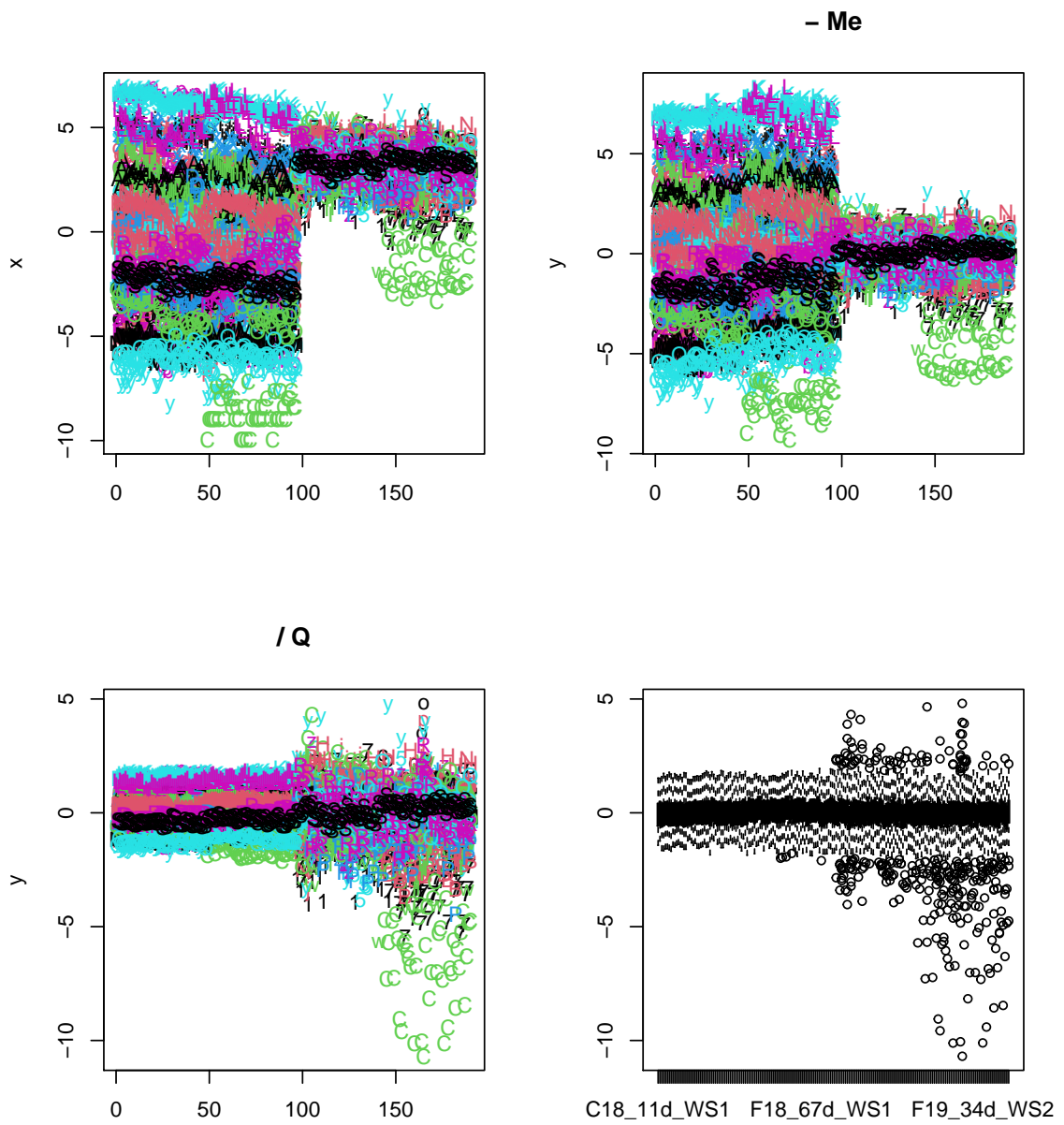




```

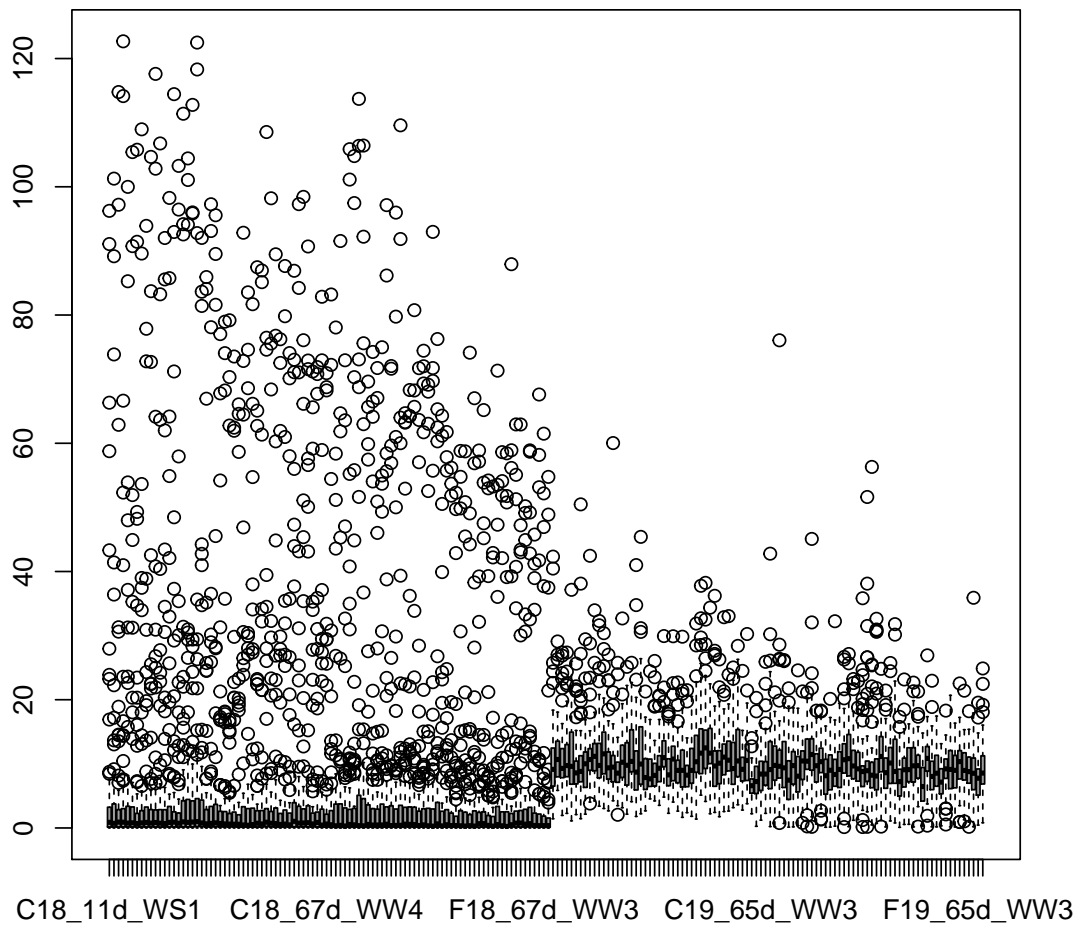
> par(mfrow=c(2,2))
> x <- log2(m1819[,m_num_id])
> matplot(x)
> dim(x)
[1] 190 55
> Me <- apply(x,1,median, na.rm=TRUE)
> Q <- apply(x,1,function(x) diff(quantile(x, probs=c(0.25,0.75), na.rm=TRUE)))
> y <- sweep(x,1,Me)
> matplot(y, main="- Me")
> y <- sweep(y,1,Q, "/")
> matplot(y, main="/ Q")
> boxplot(t(y))

```



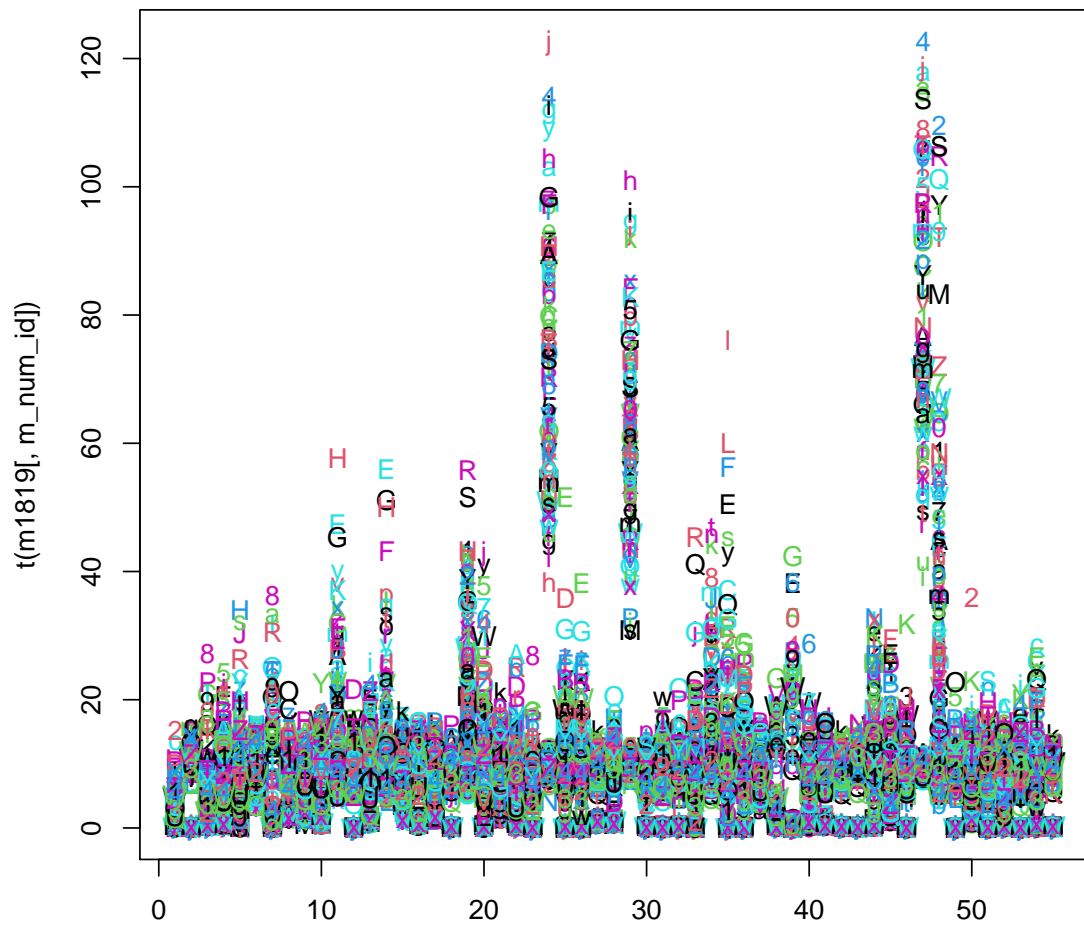
Data are highly skewed with marked differences between years

```
> boxplot(t(m1819[,m_num_id]), col=8)
```

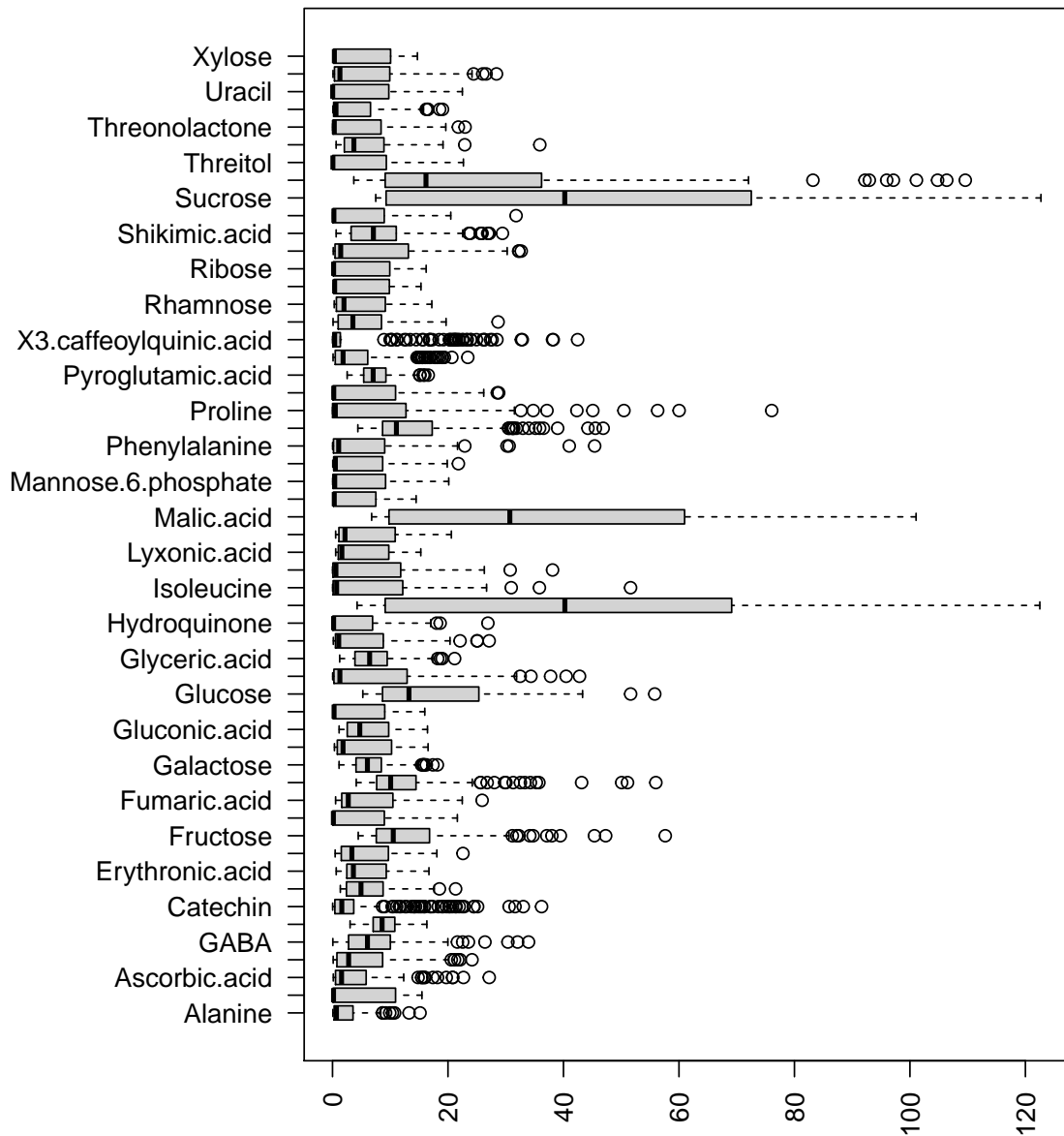



```
> matplot(t(m1819[,m_num_id]))
```

```
Warning in matplot(t(m1819[, m_num_id])): default 'pch' is smaller than number
```



```
> par(mar=c(4, 10, 3, 1))
> boxplot(m1819[,m_num_id], horizontal=TRUE, las=2)
```



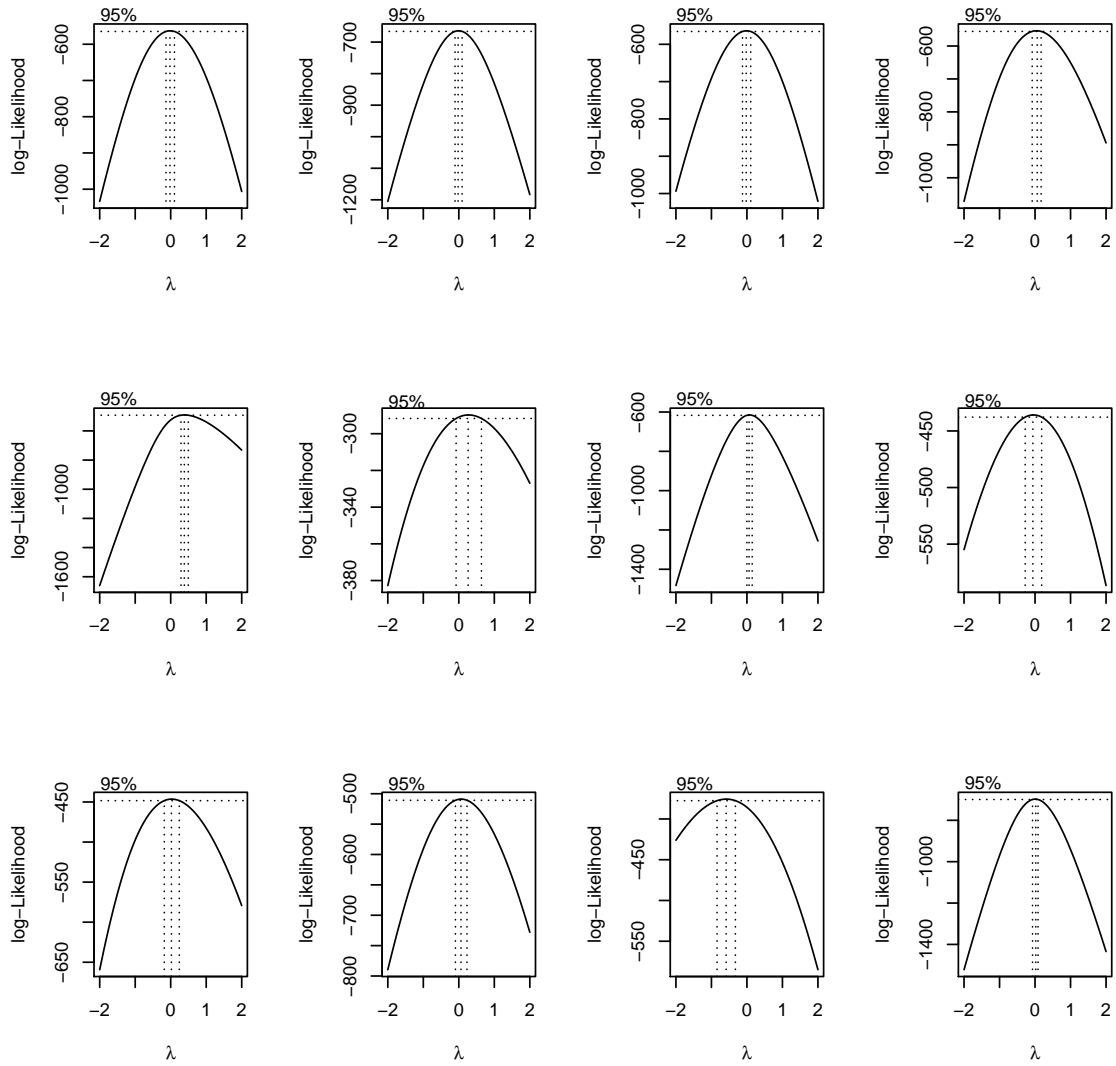
Box-Cox transformation

```

> library(MASS)
> par(mfrow=c(3,2))
> my.boxcox <- function(x, plotit=FALSE,...) {
+   require(MASS)
+   bc <- boxcox(x ~1, plotit=plotit, ...)
+   bc$x[which.max(bc$y)]
+ }

> par(mfrow=c(3,4))
> apply(m1819[,m_num_id[1:12] ], 2, my.boxcox , plotit=TRUE)

```

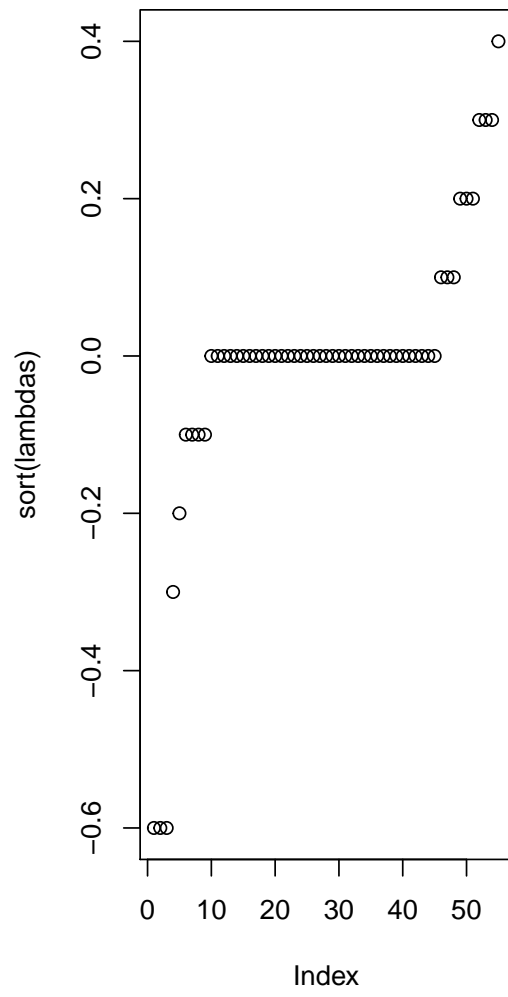
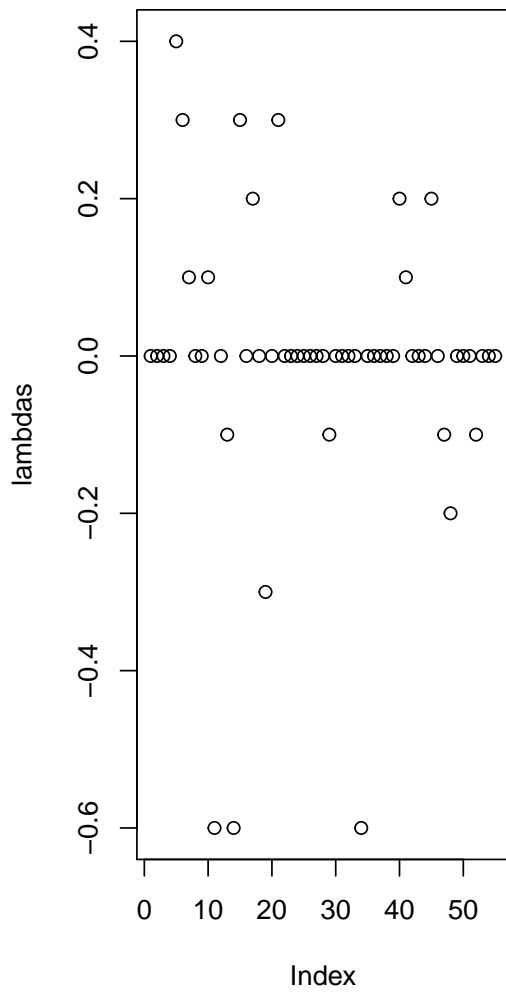


Alanine	Arabinose	Ascorbic.acid
-0.02020202	-0.02020202	-0.02020202
Aspartic.acid	GABA	Trans.Caffeic.acid
0.06060606	0.38383838	0.26262626
Catechin	Citric.acid	Erythronic.acid
0.06060606	-0.06060606	0.02020202
Ethanolamine	Fructose	Fructose.6.phosphate
0.06060606	-0.58585859	0.02020202

```

> lambdas <- apply(m1819[,m_num_id ], 2, my.boxcox )
> par(mfrow=c(1,2))
> plot(lambdas)
> plot(sort(lambdas))

```



It seems that logarithm is adequate.

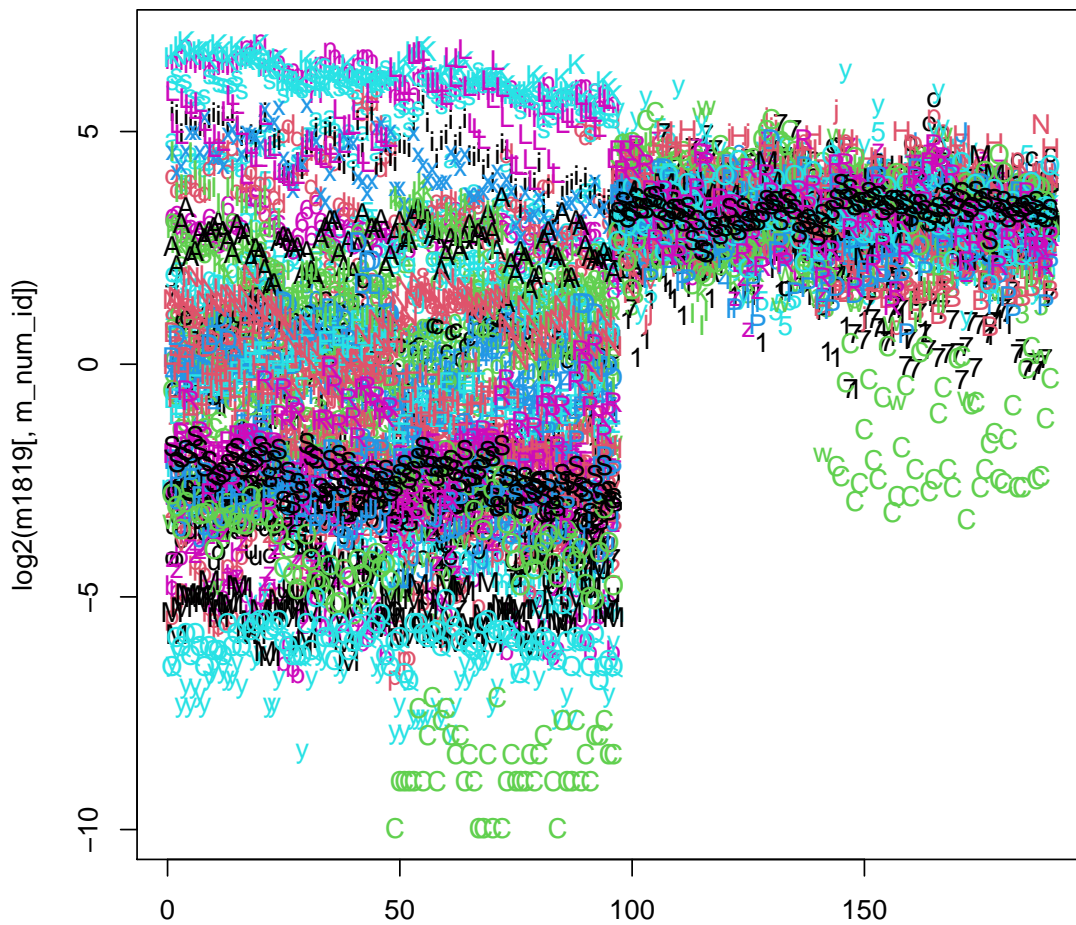
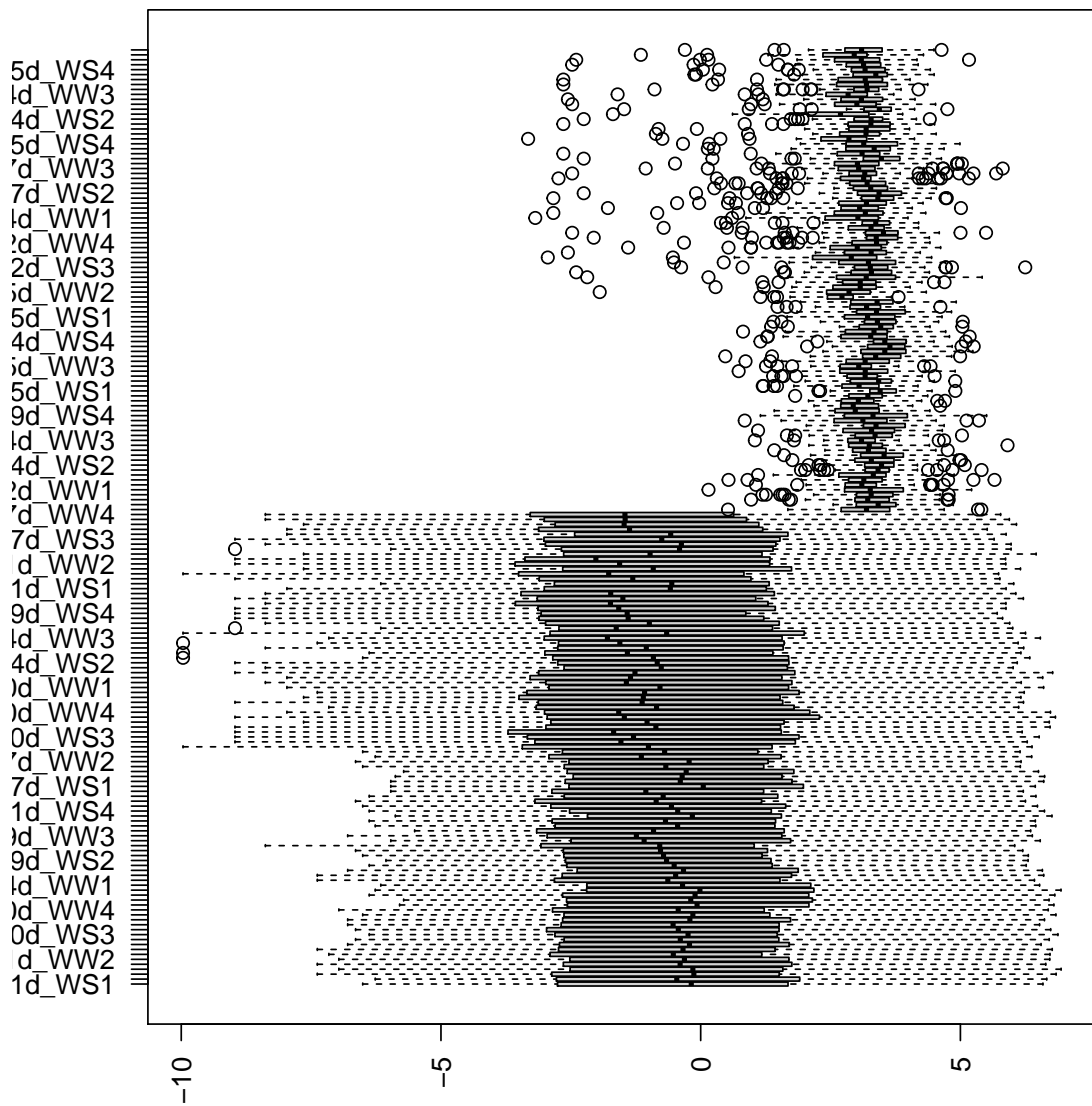


Figure 1: Comparison of variability between years

```
> matplot(log2(m1819[,m_num_id]))
```

Data are highly skewed with marked differences between years

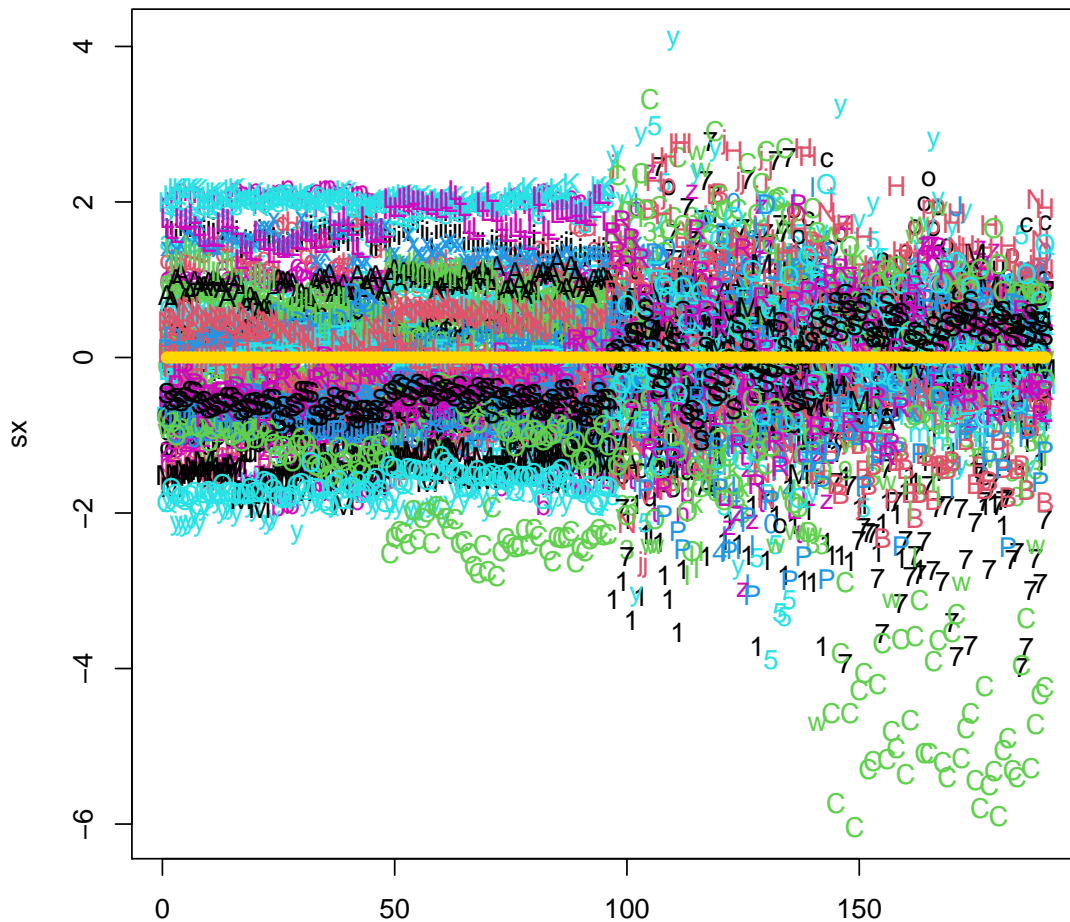
```
> boxplot(t(log2(m1819[,m_num_id]))
+ , horizontal=TRUE, las=2)
```



3.8 Standardization

For CCA standardized values are needed
Standardization across samples

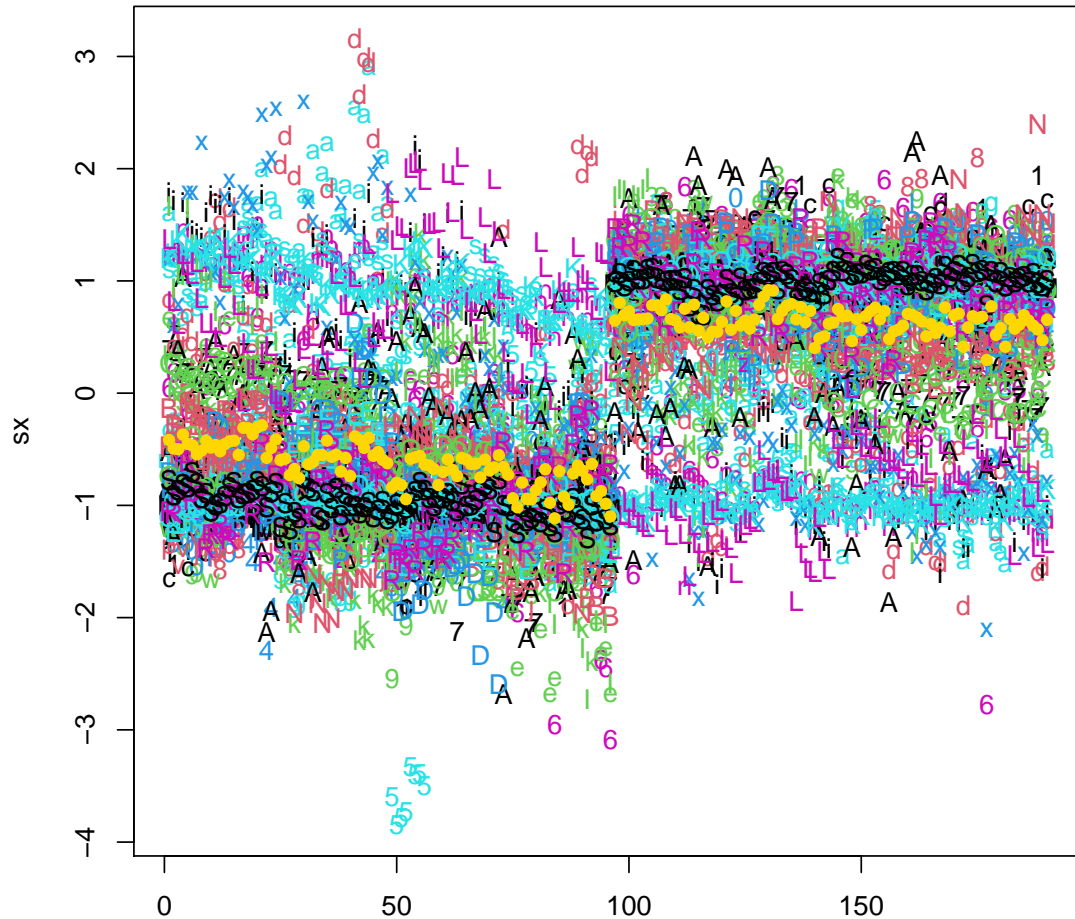
```
> x <- t(log2(m1819[,m_num_id]))
> dim(x)
[1] 55 190
> sx <- t(scale(x))
> matplot(sx)
> points(1:nrow(sx), apply(sx,1,mean, na.rm=TRUE), col="gold", pch=16)
```



```
> head(apply(sx,1,mean, na.rm=TRUE))
C18_11d_WS1 C18_11d_WS2 C18_11d_WS3 C18_11d_WS4 C18_11d_WW1
3.156507e-18 2.714645e-17 -5.391442e-19 2.231091e-17 9.791331e-18
C18_11d_WW2
-2.039877e-17
> head(apply(sx,1,sd, na.rm=TRUE))
C18_11d_WS1 C18_11d_WS2 C18_11d_WS3 C18_11d_WS4 C18_11d_WW1
1 1 1 1 1
C18_11d_WW2
1
```


Standardization across variables

```
> x <- log2(m1819[,m_num_id])
> sx <- scale(x)
> matplot(sx)
> points(1:nrow(sx), apply(sx,1,mean, na.rm=TRUE), col="gold", pch=16)
```



```
> head(apply(sx,2,mean, na.rm=TRUE))
      Alanine      Arabinose      Ascorbic.acid
-3.395835e-18 -1.762114e-17 -2.402592e-17
  Aspartic.acid      GABA  Trans.Caffeic.acid
 4.406198e-17 -1.204206e-17  1.468649e-16
```

```
> head(apply(sx,2,sd, na.rm=TRUE))
      Alanine      Arabinose      Ascorbic.acid
           1           1           1
  Aspartic.acid      GABA  Trans.Caffeic.acid
           1           1           1
```

```
> x <- log2(m18[,m_num_id])
> sx <- scale(x)
> sm18 <- sx
> matplot(t(sm18), ylim=c(-4,4))
```

2018

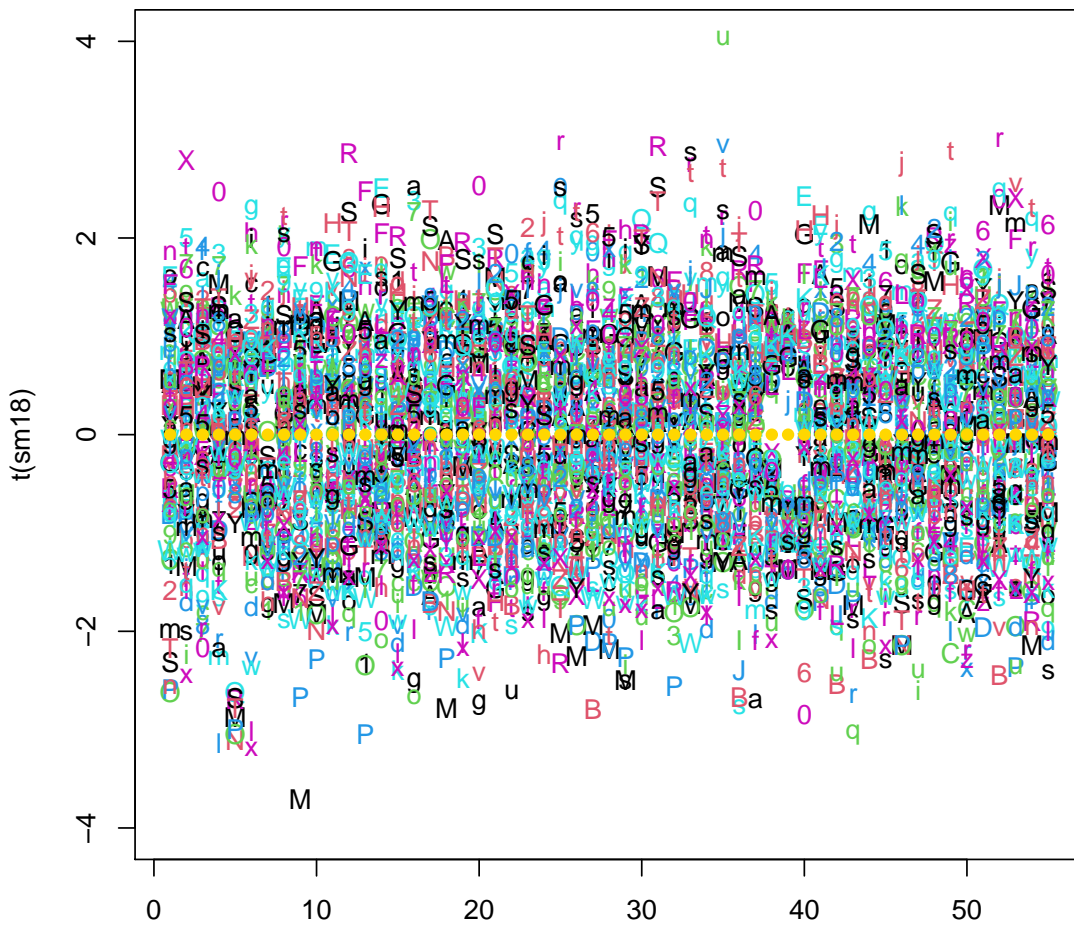


Figure 2: Plot of standardized data 2018 (log2).

Warning in matplot(t(sm18), ylim = c(-4, 4)): default 'pch' is smaller than nu

```
> title("2018")
```

```
> points(1:ncol(sx), apply(sx,2,mean, na.rm=TRUE), col="gold", pch=16)
```

```
> head(apply(sx,2,mean, na.rm=TRUE))
```

Alanine	Arabinose	Ascorbic.acid
1.220563e-16	9.353051e-17	2.322000e-17
Aspartic.acid	GABA	Trans.Caffeic.acid
3.373280e-17	-3.177277e-17	4.260915e-17

```
> head(apply(sx,2,sd, na.rm=TRUE))
```

Alanine	Arabinose	Ascorbic.acid
1	1	1
Aspartic.acid	GABA	Trans.Caffeic.acid
1	1	1

2019

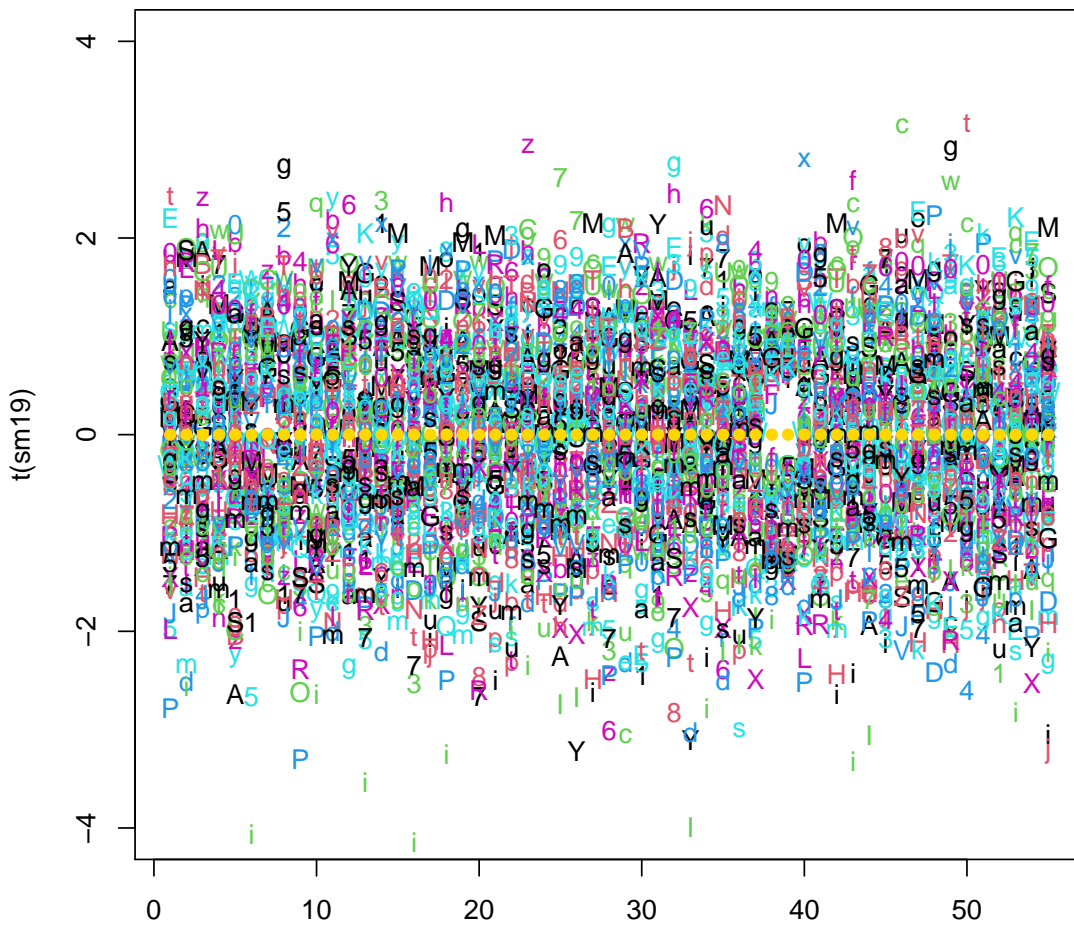


Figure 3: Plot of standardized data 2019 (log2).

```
> x <- log2(m19[,m_num_id])
> sx <- scale(x)
> sm19 <- sx
> matplot(t(sm19), ylim=c(-4,4))
Warning in matplot(t(sm19), ylim = c(-4, 4)): default 'pch' is smaller than nu
> title("2019")
> points(1:ncol(sx), apply(sx,2,mean, na.rm=TRUE), col="gold", pch=16)
> head(apply(sx,2,mean, na.rm=TRUE))
      Alanine      Arabinose      Ascorbic.acid
-1.607849e-17  3.325525e-16  -1.133143e-16
Aspartic.acid      GABA Trans.Caffeic.acid
 1.635755e-16  3.333345e-17  -2.079315e-16
> head(apply(sx,2,sd, na.rm=TRUE))
      Alanine      Arabinose      Ascorbic.acid
           1           1           1
Aspartic.acid      GABA Trans.Caffeic.acid
           1           1           1
```

```

> par(mar=c(4,10,3,1))
> sx <- sm18
> boxplot(sx, horizontal=TRUE, las=2, main="2018", ylim=range(sm1819))
Error in boxplot.default(groups, ...): object 'sm1819' not found
> points(apply(sx,2,mean, na.rm=TRUE),1:ncol(sx), col="gold", pch=16)
Error in plot.xy(xy.coords(x, y), type = type, ...): plot.new has not been cal

> par(mar=c(4,10,3,1))
> sx <- sm19
> boxplot(sx, horizontal=TRUE, las=2, main="2019", ylim=range(sm1819))
Error in boxplot.default(groups, ...): object 'sm1819' not found
> points(apply(sx,2,mean, na.rm=TRUE),1:ncol(sx), col="gold", pch=16)
Error in plot.xy(xy.coords(x, y), type = type, ...): plot.new has not been cal

> sm1819 <- rbind(sm18, sm19)
> dim(sm1819)
[1] 190 55
> matplot(t(sm1819))
Warning in matplot(t(sm1819)): default 'pch' is smaller than number of columns
> title("2018, 2019")
> sx <- sm1819
> points(1:ncol(sx), apply(sx,2,mean, na.rm=TRUE), col="gold", pch=16)
> head(apply(sx,2,mean, na.rm=TRUE))
      Alanine      Arabinose      Ascorbic.acid
5.373991e-17  2.118948e-16  -4.435749e-17
Aspartic.acid      GABA Trans.Caffeic.acid
9.802072e-17  4.516558e-19  -8.137280e-17

> head(apply(sx,2,sd, na.rm=TRUE))
      Alanine      Arabinose      Ascorbic.acid
0.997351      0.997351      0.997351
Aspartic.acid      GABA Trans.Caffeic.acid
0.997351      0.997351      0.997351

```

2018, 2019

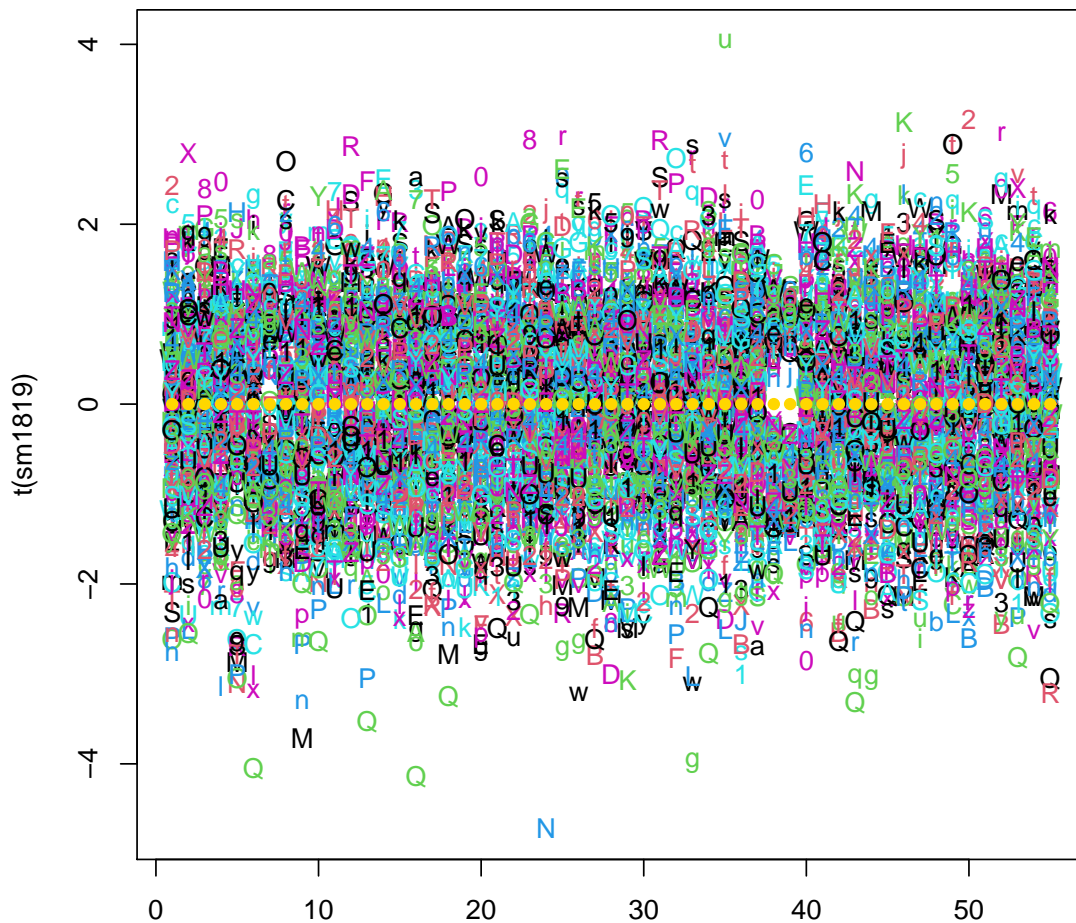


Figure 4: Plot of standardized data 2018, 2019 (log2).

3.9 Featuredata for metabolites

Rows in featuredata and columns of metabolite data ARE in the same order.

```
> (fmfn <- getMeta(.adesc, "Featuredata metabolites"))
[1] "Featuredata_metabolites_210127.txt"
> mfdata <- read.table(file.path(.iroot, fmfn), sep="\t", header=TRUE)
> cbind(colnames(sm1819), make.names(mfdata[,1]), mfdata[,1])

      [,1]                [,2]
[1,] "Alanine"           "Alanine"
[2,] "Arabinose"         "Arabinose"
[3,] "Ascorbic.acid"     "Ascorbic.acid"
[4,] "Aspartic.acid"     "Aspartic.acid"
[5,] "GABA"              "GABA"
[6,] "Trans.Caffeic.acid" "Trans.Caffeic.acid"
[7,] "Catechin"          "Catechin"
[8,] "Citric.acid"       "Citric.acid"
[9,] "Erythronic.acid"   "Erythronic.acid"
```

[10,]	"Ethanolamine"	"Ethanolamine"
[11,]	"Fructose"	"Fructose"
[12,]	"Fructose.6.phosphate"	"Fructose.6.phosphate"
[13,]	"Fumaric.acid"	"Fumaric.acid"
[14,]	"Galactinol"	"Galactinol"
[15,]	"Galactose"	"Galactose"
[16,]	"Gallic.acid"	"Gallic.acid"
[17,]	"Gluconic.acid"	"Gluconic.acid"
[18,]	"Glucopyranose..H2O."	"Glucopyranose..H2O."
[19,]	"Glucose"	"Glucose"
[20,]	"Glutamic.acid"	"Glutamic.acid"
[21,]	"Glyceric.acid"	"Glyceric.acid"
[22,]	"Glycine"	"Glycine"
[23,]	"Hydroquinone"	"Hydroquinone"
[24,]	"Myo.Inostol"	"Myo.Inostol"
[25,]	"Isoleucine"	"Isoleucine"
[26,]	"Leucine"	"Leucine"
[27,]	"Lyxonic.acid"	"Lyxonic.acid"
[28,]	"Maleic.acid"	"Maleic.acid"
[29,]	"Malic.acid"	"Malic.acid"
[30,]	"Malonic.acid"	"Malonic.acid"
[31,]	"Mannose.6.phosphate"	"Mannose.6.phosphate"
[32,]	"Melibiose"	"Melibiose"
[33,]	"Phenylalanine"	"Phenylalanine"
[34,]	"Phosphoric.acid"	"Phosphoric.acid"
[35,]	"Proline"	"Proline"
[36,]	"Putrescine"	"Putrescine"
[37,]	"Pyroglutamic.acid"	"Pyroglutamic.acid"
[38,]	"Quinic.acid"	"Quinic.acid"
[39,]	"X3.caffeoylquinic.acid"	"X3.caffeoylquinic.acid"
[40,]	"Raffinose"	"Raffinose"
[41,]	"Rhamnose"	"Rhamnose"
[42,]	"Ribonic.acid"	"Ribonic.acid"
[43,]	"Ribose"	"Ribose"
[44,]	"Serine"	"Serine"
[45,]	"Shikimic.acid"	"Shikimic.acid"
[46,]	"Succinic.acid"	"Succinic.acid"
[47,]	"Sucrose"	"Sucrose"
[48,]	"Tartaric.acid"	"Tartaric.acid"
[49,]	"Threitol"	"Threitol"
[50,]	"Threonic.acid"	"Threonic.acid"
[51,]	"Threonolactone"	"Threonolactone"
[52,]	"Threonine"	"Threonine"
[53,]	"Uracil"	"Uracil"
[54,]	"Valine"	"Valine"
[55,]	"Xylose"	"Xylose"
	[, 3]	
[1,]	"Alanine"	
[2,]	"Arabinose"	
[3,]	"Ascorbic acid"	
[4,]	"Aspartic acid"	
[5,]	"GABA"	
[6,]	"Trans-Caffeic acid"	

```

[7,] "Catechin"
[8,] "Citric acid"
[9,] "Erythronic acid"
[10,] "Ethanolamine"
[11,] "Fructose"
[12,] "Fructose-6-phosphate"
[13,] "Fumaric acid"
[14,] "Galactinol"
[15,] "Galactose"
[16,] "Gallic acid"
[17,] "Gluconic acid"
[18,] "Glucopyranose[-H2O]"
[19,] "Glucose"
[20,] "Glutamic acid"
[21,] "Glyceric acid"
[22,] "Glycine"
[23,] "Hydroquinone"
[24,] "Myo-Inositol"
[25,] "Isoleucine"
[26,] "Leucine"
[27,] "Lyxonic acid"
[28,] "Maleic acid"
[29,] "Malic acid"
[30,] "Malonic acid"
[31,] "Mannose-6-phosphate"
[32,] "Melibiose"
[33,] "Phenylalanine"
[34,] "Phosphoric acid"
[35,] "Proline"
[36,] "Putrescine"
[37,] "Pyroglutamic acid"
[38,] "Quinic acid"
[39,] "3-caffeoylquinic acid"
[40,] "Raffinose"
[41,] "Rhamnose"
[42,] "Ribonic acid"
[43,] "Ribose"
[44,] "Serine"
[45,] "Shikimic acid"
[46,] "Succinic acid"
[47,] "Sucrose"
[48,] "Tartaric acid"
[49,] "Threitol"
[50,] "Threonic acid"
[51,] "Threonolactone"
[52,] "Threonine"
[53,] "Uracil"
[54,] "Valine"
[55,] "Xylose"

> all(colnames(sm1819) == make.names(mfdata[,1]))
[1] TRUE

```

```

> rownames(mfdata) <- make.names(mfdata[,1])
> # to be sure, that the fdata rows are in the same order as data
> mfdata <- mfdata[colnames(sm1819),]
> head(mfdata)

```

	Metabolite	Bin
Alanine	Alanine	5.1001
Arabinose	Arabinose	10.6.1001
Ascorbic.acid	Ascorbic acid	17.5.1001
Aspartic.acid	Aspartic acid	13.1.1.2.1001
GABA	GABA	22.1002
Trans.Caffeic.acid	Trans-Caffeic acid	16.2.1.1006

Alanine		
Arabinose		cell wall
Ascorbic.acid		hormone metabol
Aspartic.acid	amino acid metabolism.synthesis.central	amino acid metaboli
GABA		pol
Trans.Caffeic.acid	secondary metabolism.phenylpropanoids.lignin	

3.10 Interesting bins

For Canonical Correlation Analysis (CCA) we will need to restrict the number of genes (and metabolites) to those that are of more interest. The Mapman bins of interest are provided in the file (at the investigation layer).

List of bins for metabolite and transcripts:

```
> (bfn <- getMeta(.adesc, "Interesting bins"))
[1] "Bin_selection_2.txt"
> bindata <- read.table(file.path(.iroot, bfn), sep="\t", header=TRUE, dec="$")
> head(bindata)
  bin      description
1  2      major CHO metabolism
2  3      minor CHO metabolism
3  8 TCA / organic transformation
4 10      cell wall
5 13      amino acid metabolism
6 16      secondary metabolism
> str(bindata)
'data.frame':      8 obs. of  2 variables:
 $ bin      : chr  "2" "3" "8" "10" ...
 $ description: chr  "major CHO metabolism" "minor CHO metabolism" "TCA / orga
```

At some point, the transcripts and metabolites that correspond to the bins of interest (including the sub-bins) will be selected. Let us try to find them for the first bin:

```
> bindata$bin
[1] "2" "3" "8" "10" "13" "16" "27.3" "35"
> paste0("^", bindata$bin[1], "\\.")
[1] "^2\\."
> ind <- grep(paste0("^", bindata$bin[1], "\\."), fdata$BINCODE)
> ind
 [1] 449 478 502 503 514 982 1128 1604 1654 1839 1904
[12] 2102 2191 2368 2495 2496 2497 3842 3973 4049 4398 4444
[23] 4544 5112 5424 5514 5714 5870 6003 6641 6913 7102 7367
[34] 7400 7507 7737 7987 8541 8807 8810 8952 9215 9405 9614
[45] 9783 10279 10410 10468 11168 11217 11324 11658 11967 12036 12225
[56] 12418 12547 12609 12663 13007 13181 13182 13252 13509 13868 14410
[67] 14612 14713 14740 14975 15363 15451 15526 15644 15665 15833 15871
[78] 16263 16349 16362 16622 16930
> fdata[ind, 2:3]
```

	geneID	BINCODE
449	Vitvi02g01845	2.2.1.3
478	Vitvi01g00025	2.2.1.1
502	Vitvi01g00052	2.2.2.6
503	Vitvi01g00053	2.2.2.6
514	Vitvi01g00064	2.2.2.2
982	Vitvi01g00681	2.2.2.3
1128	Vitvi01g00932	2.2.2.1.1
1604	Vitvi02g00184	2.2.2.1.2
1654	Vitvi02g00250	2.1.2.2
1839	Vitvi02g00512	2.2.1.3
1904	Vitvi02g00605	2.2.2.1.2
2102	Vitvi02g01232	2.2.2.1.2
2191	Vitvi03g00088	2.2.1.3.1
2368	Vitvi03g00304	2.1.2.1
2495	Vitvi03g00500	2.2.2.1.1
2496	Vitvi03g01571	2.2.2.1.1
2497	Vitvi03g01572	2.2.2.1.1
3842	Vitvi05g00164	2.2.1.3.1
3973	Vitvi05g00357	2.2.2.1.2
4049	Vitvi05g00442	2.1.2.1
4398	Vitvi05g01193	2.1.1.1
4444	Vitvi05g01324	2.2.2.3
4544	Vitvi05g01516	2.2.1.1
5112	Vitvi06g00583	2.2.2.2
5424	Vitvi06g01272	2.2.1.4
5514	Vitvi06g01427	2.2.1.3.1
5714	Vitvi07g00167	2.1.2.4
5870	Vitvi07g00353	2.2.1.5
6003	Vitvi07g00544	2.1.2.1
6641	Vitvi07g01830	2.2.2.4
6913	Vitvi08g00225	2.1.1.2
7102	Vitvi08g00930	2.2.2.4
7367	Vitvi08g01307	2.2.1.1
7400	Vitvi08g01353	2.2.2.4
7507	Vitvi08g01497	2.1.2.3
7737	Vitvi08g01815	2.1.2.60
7987	Vitvi09g00193	2.2.1.3
8541	Vitvi10g00094	2.1.2.2
8807	Vitvi10g00494	2.2.2.2
8810	Vitvi10g00499	2.2.2.6
8952	Vitvi10g00739	2.1.2.2
9215	Vitvi11g00030	2.2.1.5
9405	Vitvi11g00260	2.2.1.4
9614	Vitvi11g00542	2.1.1.1
9783	Vitvi11g00903	2.1.2.2
10279	Vitvi12g00558	2.2.2.1.2
10410	Vitvi12g00770	2.1.2.1
10468	Vitvi12g01618	2.1.1.2
11168	Vitvi13g00692	2.2.2.9
11217	Vitvi13g00792	2.2.1.3.1
11324	Vitvi13g01242	2.2.2.10
11658	Vitvi14g00070	2.2.1.3.1

```

11967 Vitvi14g00608      2.1.2
12036 Vitvi14g00950      2.2.1.1
12225 Vitvi14g01406      2.2.2.1.1
12418 Vitvi14g01692      2.2.1.99
12547 Vitvi14g01873      2.1.2.1
12609 Vitvi14g01968      2.1.2.2
12663 Vitvi14g02032      2.2.2.2
13007 Vitvi15g00885      2.1.2.2
13181 Vitvi15g01127      2.2.2.1.2
13182 Vitvi15g01128      2.2.2.1.2
13252 Vitvi16g00033      2.2.2.6
13509 Vitvi16g00895      2.2.1.1
13868 Vitvi16g01405      2.1.2.2
14410 Vitvi17g00738      2.1.2.1
14612 Vitvi17g01221      2.2.1.5
14713 Vitvi18g00111      2.1.2.3
14740 Vitvi18g00144      2.2.2.1.1
14975 Vitvi18g00454      2.1.2.4
15363 Vitvi18g02758      2.1.2.1
15451 Vitvi18g01114      2.2.1.4
15526 Vitvi18g01242      2.1.2
15644 Vitvi18g01657      2.1.1.3
15665 Vitvi18g01682      2.2.1.3.1
15833 Vitvi18g02365      2.1.1.1
15871 Vitvi18g02417      2.2.1.1
16263 Vitvi19g00538      2.1.2.3
16349 Vitvi19g00678      2.2.2.1.2
16362 Vitvi19g00704      2.2.1.99
16622 Vitvi10g02394      2.1.2.2
16930 Vitvi19g02064      2.2.2.1.2

```

```
> bindata$bin
```

```
[1] "2"      "3"      "8"      "10"     "13"     "16"     "27.3"  "35"
```

```
> paste0("^", "26.9", "$")
```

```
[1] "^26.9$"
```

```
> ind <- grep(paste0("^", "26.9", "$"), fdata$BINCODE)
```

```
> ind
```

```

[1]      1    590   1709   3285   3293   4238   4239   4240   4241   4311   4315
[12]  4940   5088   5806   5807   5809   6075   7120   7239   7242   7348   7349
[23]  9989   9990  10683  10689  10862  12705  12710  12903  13263  13264  13269
[34] 14299  14747  14797  16444  16446  16449  16457  16458  16459  16460  16461
[45] 16462  16463  16729  16933

```

```
> fdata[ind, 2:3]
```

```

           geneID BINCODE
1      Vitvi15g01736    26.9
590    Vitvi01g00149    26.9
1709   Vitvi02g00335    26.9
3285   Vitvi04g00880    26.9
3293   Vitvi04g00908    26.9
4238   Vitvi05g01959    26.9
4239   Vitvi05g01961    26.9

```

4240	Vitvi05g01962	26.9
4241	Vitvi05g01963	26.9
4311	Vitvi05g02001	26.9
4315	Vitvi05g00910	26.9
4940	Vitvi06g00372	26.9
5088	Vitvi06g01724	26.9
5806	Vitvi07g00283	26.9
5807	Vitvi07g00285	26.9
5809	Vitvi07g00286	26.9
6075	Vitvi07g00674	26.9
7120	Vitvi08g00957	26.9
7239	Vitvi08g01129	26.9
7242	Vitvi08g01137	26.9
7348	Vitvi08g02226	26.9
7349	Vitvi08g02228	26.9
9989	Vitvi12g00080	26.9
9990	Vitvi12g00081	26.9
10683	Vitvi12g02153	26.9
10689	Vitvi12g02159	26.9
10862	Vitvi13g00208	26.9
12705	Vitvi15g00172	26.9
12710	Vitvi15g00231	26.9
12903	Vitvi15g00746	26.9
13263	Vitvi16g01498	26.9
13264	Vitvi16g01499	26.9
13269	Vitvi16g01501	26.9
14299	Vitvi17g01467	26.9
14747	Vitvi18g00152	26.9
14797	Vitvi18g00219	26.9
16444	Vitvi19g02145	26.9
16446	Vitvi19g02150	26.9
16449	Vitvi19g01048	26.9
16457	Vitvi19g02189	26.9
16458	Vitvi19g02192	26.9
16459	Vitvi19g01305	26.9
16460	Vitvi19g01311	26.9
16461	Vitvi19g01326	26.9
16462	Vitvi19g02197	26.9
16463	Vitvi19g01338	26.9
16729	Vitvi07g02188	26.9
16933	Vitvi19g01328	26.9

```

> xx <- "27.3"
> bindata$bin
[1] "2"      "3"      "8"      "10"     "13"     "16"     "27.3"  "35"
> paste0("^",xx,"$")
[1] "^27.3$"
> ind1 <- grep(paste0("^",xx,"$"),fdata$BINCODE)
> ind1
[1] 1421 3631 4719 8064 11915 13247 13534 14400 14799 14801
> fdata[ind1,2:3]

```

```

      geneID BINCODE
1421 Vitvi01g01749 27.3
3631 Vitvi04g01606 27.3
4719 Vitvi06g00072 27.3
8064 Vitvi09g00288 27.3
11915 Vitvi14g00482 27.3
13247 Vitvi16g00026 27.3
13534 Vitvi16g00956 27.3
14400 Vitvi17g00725 27.3
14799 Vitvi18g00222 27.3
14801 Vitvi18g00224 27.3

```

```
> bindata$bin
```

```
[1] "2" "3" "8" "10" "13" "16" "27.3" "35"
```

```
> paste0("^",xx,"\\.")
```

```
[1] "^27.3\\."
```

```
> ind2 <- grep(paste0("^",xx,"\\."),fdata$BINCODE)
```

```
> ind2
```

```

 [1] 20 23 73 83 86 96 107 111 195 223
[11] 226 265 293 309 316 337 343 345 377 415
[21] 512 513 519 545 577 610 633 651 655 660
[31] 662 677 688 690 737 738 754 765 779 814
[41] 816 838 839 841 862 893 894 909 958 979
[51] 1029 1066 1080 1086 1090 1098 1117 1119 1129 1130
[61] 1131 1132 1139 1141 1144 1151 1176 1177 1186 1198
[71] 1199 1203 1215 1226 1231 1243 1268 1269 1270 1284
[81] 1286 1287 1296 1308 1364 1365 1374 1382 1391 1408
[91] 1411 1413 1425 1427 1437 1454 1464 1482 1485 1516
[101] 1534 1552 1557 1583 1586 1601 1614 1617 1618 1642
[111] 1647 1650 1667 1669 1680 1699 1704 1706 1718 1727
[121] 1736 1745 1751 1765 1775 1778 1780 1803 1805 1809
[131] 1838 1923 1929 1937 1942 1948 1961 1966 1972 1977
[141] 1992 2002 2003 2004 2013 2053 2088 2125 2135 2137
[151] 2144 2152 2155 2156 2161 2170 2172 2173 2178 2198
[161] 2214 2226 2232 2249 2250 2293 2308 2316 2338 2343
[171] 2346 2359 2360 2380 2383 2387 2390 2400 2403 2428
[181] 2444 2461 2473 2484 2508 2509 2541 2555 2578 2579
[191] 2592 2594 2644 2646 2662 2681 2706 2727 2752 2765
[201] 2770 2781 2802 2803 2804 2807 2828 2829 2841 2843
[211] 2854 2876 2888 2896 2928 2931 2940 2941 2942 2956
[221] 2966 2994 3023 3024 3025 3046 3054 3061 3064 3081
[231] 3082 3095 3105 3127 3139 3140 3142 3152 3166 3191
[241] 3217 3221 3222 3223 3224 3232 3252 3258 3259 3274
[251] 3283 3295 3304 3353 3357 3358 3362 3374 3387 3388
[261] 3389 3390 3391 3411 3416 3449 3451 3453 3504 3506
[271] 3510 3515 3528 3540 3546 3564 3571 3572 3613 3617
[281] 3621 3622 3626 3630 3650 3659 3667 3668 3669 3687
[291] 3696 3704 3745 3772 3774 3785 3791 3799 3824 3882
[301] 3907 3911 3913 3915 3950 3951 3953 3954 3983 4039
[311] 4056 4084 4115 4158 4162 4163 4176 4195 4210 4235
[321] 4263 4279 4286 4293 4320 4352 4353 4358 4359 4370
[331] 4478 4482 4484 4547 4555 4573 4585 4607 4631 4654
[341] 4678 4686 4712 4735 4762 4789 4790 4795 4799 4801

```

[351]	4806	4813	4826	4830	4846	4847	4848	4849	4850	4863
[361]	4864	4867	4879	4886	4889	4893	4896	4897	4898	4913
[371]	4921	4932	4938	4942	4945	4952	4967	5006	5021	5027
[381]	5032	5040	5042	5053	5060	5104	5119	5130	5142	5150
[391]	5163	5200	5246	5247	5287	5288	5293	5301	5312	5316
[401]	5323	5336	5353	5370	5373	5399	5403	5419	5431	5454
[411]	5464	5469	5470	5481	5490	5491	5495	5513	5527	5537
[421]	5547	5563	5564	5578	5586	5593	5594	5605	5617	5618
[431]	5621	5623	5624	5640	5641	5677	5687	5695	5703	5712
[441]	5724	5741	5744	5780	5789	5794	5797	5827	5872	5879
[451]	5899	5922	5923	5924	5927	5936	5938	5947	5973	5983
[461]	5984	5985	5988	5994	6007	6014	6045	6049	6061	6062
[471]	6064	6073	6087	6120	6121	6131	6138	6151	6161	6162
[481]	6178	6180	6315	6325	6326	6385	6394	6408	6421	6467
[491]	6474	6501	6539	6541	6553	6561	6564	6568	6580	6586
[501]	6593	6612	6622	6633	6634	6652	6663	6671	6673	6675
[511]	6680	6711	6712	6718	6754	6762	6775	6776	6780	6788
[521]	6789	6794	6795	6799	6845	6849	6862	6880	6893	6900
[531]	6909	6923	6928	6933	6939	6949	6951	6969	6978	6980
[541]	7010	7046	7053	7066	7067	7075	7082	7099	7116	7124
[551]	7125	7131	7148	7151	7157	7167	7180	7187	7201	7241
[561]	7249	7253	7260	7284	7295	7297	7306	7310	7313	7327
[571]	7341	7358	7364	7387	7407	7416	7431	7501	7518	7524
[581]	7532	7553	7571	7589	7592	7595	7598	7609	7637	7665
[591]	7666	7668	7671	7674	7716	7722	7743	7748	7751	7752
[601]	7758	7759	7760	7764	7768	7774	7778	7779	7786	7790
[611]	7793	7803	7818	7821	7828	7875	7876	7880	7887	7925
[621]	7927	7932	7939	7940	7962	7985	8001	8007	8010	8017
[631]	8020	8059	8072	8075	8079	8098	8101	8117	8138	8139
[641]	8146	8157	8158	8220	8231	8333	8345	8405	8431	8438
[651]	8461	8476	8478	8482	8504	8519	8525	8563	8574	8599
[661]	8603	8622	8625	8628	8656	8657	8666	8672	8673	8676
[671]	8677	8691	8696	8699	8702	8726	8729	8733	8745	8760
[681]	8787	8788	8794	8797	8809	8816	8828	8858	8872	8883
[691]	8893	8914	8923	8948	8960	8998	9009	9013	9014	9039
[701]	9040	9047	9072	9073	9084	9092	9094	9120	9121	9124
[711]	9125	9126	9148	9149	9158	9166	9174	9202	9209	9210
[721]	9211	9226	9265	9271	9272	9282	9285	9287	9294	9309
[731]	9318	9321	9325	9331	9337	9339	9354	9358	9373	9381
[741]	9382	9383	9391	9446	9448	9455	9460	9472	9489	9493
[751]	9508	9513	9517	9539	9543	9544	9587	9588	9593	9607
[761]	9633	9661	9683	9703	9712	9775	9788	9807	9813	9829
[771]	9840	9842	9844	9846	9851	9872	9895	9898	9902	9933
[781]	9968	9981	9988	10000	10006	10023	10062	10073	10074	10076
[791]	10097	10108	10123	10145	10154	10164	10183	10199	10200	10216
[801]	10217	10218	10233	10262	10267	10268	10269	10278	10283	10286
[811]	10313	10318	10319	10348	10349	10371	10391	10401	10411	10471
[821]	10485	10573	10596	10598	10600	10617	10650	10651	10663	10671
[831]	10679	10681	10696	10726	10739	10745	10747	10758	10762	10764
[841]	10789	10790	10824	10829	10843	10848	10857	10867	10914	10918
[851]	10945	10951	10954	10955	10996	11035	11042	11048	11057	11058
[861]	11060	11062	11063	11071	11080	11085	11108	11113	11114	11126
[871]	11127	11128	11141	11157	11171	11172	11227	11265	11266	11282

```

[881] 11293 11298 11316 11319 11326 11337 11360 11366 11378 11395
[891] 11397 11403 11406 11407 11408 11412 11459 11460 11491 11514
[901] 11528 11534 11544 11551 11558 11579 11581 11584 11619 11626
[911] 11631 11638 11639 11681 11707 11759 11761 11763 11770 11805
[921] 11807 11830 11870 11891 11896 11910 11912 11916 11918 11928
[931] 11944 11947 11954 11968 11971 11975 11996 11998 12012 12015
[941] 12032 12033 12045 12052 12072 12108 12132 12162 12177 12185
[951] 12218 12240 12267 12278 12288 12293 12294 12297 12312 12317
[961] 12349 12355 12360 12361 12367 12377 12378 12383 12385 12388
[971] 12437 12461 12486 12487 12507 12518 12540 12544 12565 12566
[981] 12587 12591 12602 12603 12612 12622 12623 12665 12676 12680
[991] 12707 12733 12759 12810 12820 12842 12843 12878 12889 12898
[1001] 12914 12918 12919 12939 12941 12960 12982 12983 12984 12985
[1011] 12986 12990 13001 13018 13020 13025 13034 13039 13041 13048
[1021] 13049 13052 13053 13065 13066 13082 13083 13102 13125 13152
[1031] 13155 13159 13160 13163 13173 13180 13184 13201 13206 13236
[1041] 13245 13250 13255 13278 13282 13299 13303 13305 13336 13348
[1051] 13354 13376 13421 13432 13457 13489 13511 13521 13544 13561
[1061] 13572 13592 13593 13603 13628 13674 13696 13697 13698 13761
[1071] 13787 13798 13799 13810 13813 13815 13821 13845 13872 13880
[1081] 13896 13905 13906 13912 13928 13940 13944 13958 13983 13991
[1091] 13995 13997 14024 14051 14083 14103 14111 14113 14125 14137
[1101] 14154 14155 14172 14199 14203 14225 14229 14232 14243 14251
[1111] 14253 14267 14291 14293 14302 14313 14317 14323 14330 14349
[1121] 14350 14357 14358 14364 14376 14401 14406 14423 14425 14428
[1131] 14442 14451 14459 14489 14497 14503 14515 14556 14557 14566
[1141] 14568 14570 14609 14611 14616 14641 14650 14653 14660 14662
[1151] 14677 14691 14694 14696 14701 14745 14750 14770 14783 14789
[1161] 14798 14812 14815 14822 14824 14848 14873 14885 14889 14896
[1171] 14909 14926 14930 14950 14961 14972 15000 15021 15025 15033
[1181] 15043 15044 15046 15057 15059 15067 15068 15103 15106 15113
[1191] 15132 15164 15171 15176 15180 15188 15197 15202 15203 15246
[1201] 15248 15253 15265 15268 15291 15292 15304 15312 15319 15348
[1211] 15373 15386 15404 15406 15407 15409 15418 15419 15426 15431
[1221] 15448 15449 15476 15483 15490 15507 15511 15516 15525 15558
[1231] 15588 15618 15619 15620 15621 15625 15633 15635 15640 15641
[1241] 15650 15734 15775 15853 15854 15862 15881 15889 15904 15926
[1251] 15967 15977 15979 15988 15990 15993 16009 16073 16074 16087
[1261] 16107 16153 16166 16174 16180 16228 16231 16250 16257 16264
[1271] 16275 16285 16289 16294 16296 16312 16313 16317 16329 16344
[1281] 16352 16403 16423 16428 16496 16524 16525 16526 16556 16560
[1291] 16564 16593 16594 16600 16604 16608 16641 16651 16667 16678
[1301] 16682 16686 16746 16753 16755 16786 16793 16838 16862 16871
[1311] 16882 16909 16919 16935

```

```
> head(sort(fdata[ind2,3]))
```

```
[1] "27.3.1" "27.3.1" "27.3.1" "27.3.10" "27.3.10" "27.3.10"
```

```
> ind <- c(ind1,ind2)
```

```
> table(fdata[ind,3])
```

```

 27.3  27.3.1 27.3.10 27.3.11 27.3.12 27.3.13 27.3.14 27.3.15
   10     3     5     73     20     2     7     5
27.3.16 27.3.17 27.3.18 27.3.19 27.3.2 27.3.20 27.3.21 27.3.22

```

```

      5      4      3      2      6      7      14      41
27.3.23 27.3.24 27.3.25 27.3.26 27.3.27 27.3.29 27.3.3 27.3.30
      15      12      90      15      44      11      34      21
27.3.32 27.3.34 27.3.35 27.3.36 27.3.37 27.3.38 27.3.39 27.3.4
      46      10      55      10      8      2      3      18
27.3.40 27.3.41 27.3.42 27.3.44 27.3.46 27.3.47 27.3.48 27.3.49
      18      2      4      27      5      2      7      2
  27.3.5 27.3.50 27.3.51 27.3.52 27.3.53 27.3.54 27.3.55 27.3.57
      18      27      3      11      1      12      12      21
27.3.58 27.3.59 27.3.6 27.3.60 27.3.61 27.3.62 27.3.63 27.3.64
      4      10      78      5      2      10      13      3
27.3.65 27.3.66 27.3.67 27.3.68 27.3.69 27.3.7 27.3.70 27.3.71
      3      2      117      10      28      21      3      11
27.3.72 27.3.73 27.3.75 27.3.8 27.3.80 27.3.81 27.3.82 27.3.84
      1      7      2      20      7      2      2      4
27.3.85 27.3.86 27.3.9 27.3.99
      6      3      12      165

```

```
> length(ind)
```

```
[1] 1324
```

```
> #@ function to find a bin and sub-bins
```

```
> in_bins <- function(x, bincodes) {
```

```
+   in1 <- grep(paste0("^", x, "\\."), bincodes)
```

```
+   in2 <- grep(paste0("^", x, "$"), bincodes)
```

```
+   in3 <- c(in2, in1)
```

```
+   return(in3)
```

```
+ }
```

```
> in_bins("2", c("2", "2.1", "2.1.1", "22", "22.2.1"))
```

```
[1] 1 2 3
```

3.10.1 Genes in bins of interest

Find genes in sub-bins

```
> intgenes1 <- sapply(bindata$bin, function(x) fdata$geneID[grep(paste0("^", x,
> str(intgenes1)
```

```
List of 8
```

```
$ 2 : chr [1:82] "Vitvi02g01845" "Vitvi01g00025" "Vitvi01g00052" "Vitvi01g0
```

```
$ 3 : chr [1:84] "Vitvi02g01701" "Vitvi10g02295" "Vitvi00g01259" "Vitvi01g0
```

```
$ 8 : chr [1:56] "Vitvi07g03070" "Vitvi01g00274" "Vitvi01g00698" "Vitvi01g0
```

```
$ 10 : chr [1:201] "Vitvi02g01716" "Vitvi07g02990" "Vitvi10g02284" "Vitvi10g
```

```
$ 13 : chr [1:187] "Vitvi07g03049" "Vitvi00g00869" "Vitvi02g01749" "Vitvi01g
```

```
$ 16 : chr [1:237] "Vitvi02g01683" "Vitvi02g01722" "Vitvi10g02185" "Vitvi07g
```

```
$ 27.3: chr [1:1314] "Vitvi02g01753" "Vitvi02g01762" "Vitvi02g01795" "Vitvi02
```

```
$ 35 : chr [1:6244] "Vitvi07g02832" "Vitvi07g02830" "Vitvi07g02812" "Vitvi07
```

Find genes in bins (no dot at the end)

```
> intgenes2 <- sapply(bindata$bin, function(x) fdata$geneID[grep(paste0("^",
> str(intgenes2)
```



```

List of 8
 $ 2   : chr(0)
 $ 3   : chr(0)
 $ 8   : chr(0)
 $ 10  : chr(0)
 $ 13  : chr [1:8] "Vitvi01g01704" "Vitvi02g00467" "Vitvi04g00506" "Vitvi07g01
 $ 16  : chr "Vitvi08g01337"
 $ 27.3: chr [1:10] "Vitvi01g01749" "Vitvi04g01606" "Vitvi06g00072" "Vitvi09g0
 $ 35  : chr(0)

```

Combine two sets

```

> intgenes <- Map(c, intgenes1, intgenes2)
> str(intgenes)
List of 8
 $ 2   : chr [1:82] "Vitvi02g01845" "Vitvi01g00025" "Vitvi01g00052" "Vitvi01g0
 $ 3   : chr [1:84] "Vitvi02g01701" "Vitvi10g02295" "Vitvi00g01259" "Vitvi01g0
 $ 8   : chr [1:56] "Vitvi07g03070" "Vitvi01g00274" "Vitvi01g00698" "Vitvi01g0
 $ 10  : chr [1:201] "Vitvi02g01716" "Vitvi07g02990" "Vitvi10g02284" "Vitvi10g
 $ 13  : chr [1:195] "Vitvi07g03049" "Vitvi00g00869" "Vitvi02g01749" "Vitvi01g
 $ 16  : chr [1:238] "Vitvi02g01683" "Vitvi02g01722" "Vitvi10g02185" "Vitvi07g
 $ 27.3: chr [1:1324] "Vitvi02g01753" "Vitvi02g01762" "Vitvi02g01795" "Vitvi02
 $ 35  : chr [1:6244] "Vitvi07g02832" "Vitvi07g02830" "Vitvi07g02812" "Vitvi07

```

```

> # unlist(intgenes)table(fdata$BINCODE)
> head(fdata[fdata[, "geneID"]%in% unlist(intgenes),2:3], 10)

```

	geneID	BINCODE
2	Vitvi07g02832	35.2
3	Vitvi07g02830	35.2
4	Vitvi07g02812	35.2
5	Vitvi07g02811	35.2
6	Vitvi09g02033	35.2
7	Vitvi09g02034	35.2
8	Vitvi08g02428	35.2
10	Vitvi07g02753	35.2
16	Vitvi07g03006	35.2
20	Vitvi02g01753	27.3.30

```

> tail(fdata[fdata[, "geneID"]%in% unlist(intgenes),2:3], 10)

```

	geneID	BINCODE
16919	Vitvi18g01322	27.3.21
16921	Vitvi18g01950	35.2
16922	Vitvi18g03096	35.2
16927	Vitvi19g01926	35.2
16928	Vitvi19g01994	35.2
16929	Vitvi19g02044	35.2
16930	Vitvi19g02064	2.2.2.1.2
16932	Vitvi19g01155	35.2
16934	Vitvi19g01515	35.2
16935	Vitvi19g01589	27.3.37

```

> length(unique(unlist(intgenes)))

```

```

[1] 8338

```

```

> nn <- t(t(sapply(intgenes, length)))
> nn
      [,1]
2      82
3      84
8      56
10     201
13     195
16     238
27.3 1324
35     6244

```

Number of interesting genes: 8338.

3.10.2 Metabolites in bins of interest

```

> intmtbs <- sapply(bindata$bin, function(x) rownames(mfdata)[in_bins(x, mfdata$Bin)])
> str(intmtbs)

```

```

List of 8
 $ 2   : chr [1:3] "Fructose" "Glucose" "Sucrose"
 $ 3   : chr [1:7] "Erythronic.acid" "Galactinol" "Galactose" "Myo.Inostol" ..
 $ 8   : chr [1:2] "Fumaric.acid" "Succinic.acid"
 $ 10  : chr [1:5] "Arabinose" "Melibiose" "Rhamnose" "Ribonic.acid" ...
 $ 13  : chr [1:8] "Aspartic.acid" "Isoleucine" "Leucine" "Phenylalanine" ...
 $ 16  : chr [1:6] "Trans.Caffeic.acid" "Catechin" "Gallic.acid" "Hydroquinone"
 $ 27.3: chr(0)
 $ 35  : chr [1:8] "Gluconic.acid" "Glucopyranose..H2O." "Lyxonic.acid" "Malei

```

```

> x <- "2"
> mfdata$Bin[grep(paste0("^", x), mfdata$Bin)]
[1] "22.1002"      "2.2.1.1001" "2.2.1.1002" "21.1009"      "22.1003"
[6] "21.1010"      "2.1.1.1006" "21.1011"      "23.2.1002"
> mfdata$Bin[in_bins(x, mfdata$Bin)]
[1] "2.2.1.1001" "2.2.1.1002" "2.1.1.1006"

```

```

> nn <- t(t(sapply(intmtbs, length)))
> nn
      [,1]
2         3
3         7
8         2
10        5
13        8
16        6
27.3      0
35        8
> # unlist(intmtbs)
> head(mfdata[unlist(intmtbs), 2:3], 10)

```

```

                Bin
Fructose      2.2.1.1001
Glucose       2.2.1.1002
Sucrose       2.1.1.1006
Erythronic.acid 3.99.1007
Galactinol    3.1.1001
Galactose     3.99.1010
Myo.Inostol   3.4.1001
Raffinose     3.1.1002
Threitol      3.3.1011
Xylose        3.99.1027

```

```

                Description
Fructose      major CHO metabolism.degradation.sucrose.fructose
Glucose       major CHO metabolism.degradation.sucrose.glucose
Sucrose       major CHO metabolism.synthesis.sucrose.sucrose
Erythronic.acid  minor CHO metabolism.misc.erythronic acid
Galactinol    minor CHO metabolism.raffinose family.galactinol
Galactose     minor CHO metabolism.misc.galactose
Myo.Inostol   minor CHO metabolism.myo-inositol.inositol
Raffinose     minor CHO metabolism.raffinose family.raffinose
Threitol      minor CHO metabolism.sugar alcohols.threitol
Xylose        minor CHO metabolism.misc.xylose

```

```
> tail(mfdata[unlist(intmtbs),2:3], 10)
```

```

                Bin
Quinic.acid      16.2.99.1044
X3.caffeoylquinic.acid 16.2.99.1008
Gluconic.acid    35.1.1012
Glucopyranose..H2O. 35.1.1038
Lyxonic.acid     35.1.1023
Maleic.acid      35.1.1024
Malonic.acid     35.1.1036
Phosphoric.acid 35.1.1035
Tartaric.acid   35.1.1037
Threonolactone   35.1.1034

```

```

Quinic.acid      secondary metabolism.phenylpropanoids.unspecified
X3.caffeoylquinic.acid secondary metabolism.phenylpropanoids.unspecified.3-caf
Gluconic.acid    not assigned.no ontology.g
Glucopyranose..H2O. not assigned.no ontology.g
Lyxonic.acid     not assigned.no ontology.g
Maleic.acid      not assigned.no onto
Malonic.acid     not assigned.no ontology.g
Phosphoric.acid not assigned.no ontology.pho
Tartaric.acid   not assigned.no ontology.t
Threonolactone   not assigned.no ontology.th

```

```
> length(unique(unlist(intmtbs)))
```

```
[1] 39
```

3.11 Created objects overview

```
> #if(!exists("addObject")) {
> addObject <- function(x=NULL, desc="", x0=my.objects) {
+ if(is.null(x)) x0 <- data.frame(name="", description="", class="", nrow=NA,
+ nr <- length(x)
+ nc <- ncol(x)
+ if( is.data.frame(x) | is.matrix(x) | is.array(x) ) {
+   nr <- nrow(x)
+   nc <- ncol(x)
+ }
+ if(is.null(nc)) nc <- NA
+ x0 <- rbind(x0, c(deparse(substitute(x)), desc, class(x)[1], nr, nc))
+ x0 <- x0[x0$name!="", ]
+ }
+ rownames(x0) <- 1:nrow(x0)
+ return(x0)
+ }
> #}

> (my.objects <- addObject())
  name description class nrow ncol
1                NA    NA

> (my.objects <- addObject(t18, "Transcripts for 2018"))
  name      description      class  nrow ncol
1  t18 Transcripts for 2018 data.frame 15242  48

> (my.objects <- addObject(t19, "Transcripts for 2019"))
  name      description      class  nrow ncol
1  t18 Transcripts for 2018 data.frame 15242  48
2  t19 Transcripts for 2019 data.frame 15242  32

> (my.objects <- addObject(t1819, "Transcripts for 18/19"))
  name      description      class  nrow ncol
1  t18 Transcripts for 2018 data.frame 15242  48
2  t19 Transcripts for 2019 data.frame 15242  32
3 t1819 Transcripts for 18/19 data.frame 15242  80

> (my.objects <- addObject(pdata18, "Phenodata for 2018"))
  name      description      class  nrow ncol
1  t18 Transcripts for 2018 data.frame 15242  48
2  t19 Transcripts for 2019 data.frame 15242  32
3  t1819 Transcripts for 18/19 data.frame 15242  80
4 pdata18 Phenodata for 2018 data.frame  48  22

> (my.objects <- addObject(pdata19, "Phenodata for 2019"))
  name      description      class  nrow ncol
1  t18 Transcripts for 2018 data.frame 15242  48
2  t19 Transcripts for 2019 data.frame 15242  32
3  t1819 Transcripts for 18/19 data.frame 15242  80
4 pdata18 Phenodata for 2018 data.frame  48  22
5 pdata19 Phenodata for 2019 data.frame  32  22

> (my.objects <- addObject(pdata1819, "Phenodata for 2018/19"))
```

```

      name      description      class  nrow ncol
1      t18  Transcripts for 2018 data.frame 15242  48
2      t19  Transcripts for 2019 data.frame 15242  32
3     t1819 Transcripts for 18/19 data.frame 15242  80
4    pdata18  Phenodata for 2018 data.frame   48  22
5    pdata19  Phenodata for 2019 data.frame   32  22
6  pdata1819 Phenodata for 2018/19 data.frame   80  22
> (my.objects <- addObject(fdata,"Trans. featuredata 18/19"))
      name      description      class  nrow ncol
1      t18  Transcripts for 2018 data.frame 15242  48
2      t19  Transcripts for 2019 data.frame 15242  32
3     t1819 Transcripts for 18/19 data.frame 15242  80
4    pdata18  Phenodata for 2018 data.frame   48  22
5    pdata19  Phenodata for 2019 data.frame   32  22
6  pdata1819 Phenodata for 2018/19 data.frame   80  22
7      fdata Trans. featuredata 18/19 data.frame 16936  5
> (my.objects <- addObject(m18,"Metabolites for 2018"))
      name      description      class  nrow ncol
1      t18  Transcripts for 2018 data.frame 15242  48
2      t19  Transcripts for 2019 data.frame 15242  32
3     t1819 Transcripts for 18/19 data.frame 15242  80
4    pdata18  Phenodata for 2018 data.frame   48  22
5    pdata19  Phenodata for 2019 data.frame   32  22
6  pdata1819 Phenodata for 2018/19 data.frame   80  22
7      fdata Trans. featuredata 18/19 data.frame 16936  5
8      m18  Metabolites for 2018 data.frame   96  60
> (my.objects <- addObject(m19,"Metabolites for 2019"))
      name      description      class  nrow ncol
1      t18  Transcripts for 2018 data.frame 15242  48
2      t19  Transcripts for 2019 data.frame 15242  32
3     t1819 Transcripts for 18/19 data.frame 15242  80
4    pdata18  Phenodata for 2018 data.frame   48  22
5    pdata19  Phenodata for 2019 data.frame   32  22
6  pdata1819 Phenodata for 2018/19 data.frame   80  22
7      fdata Trans. featuredata 18/19 data.frame 16936  5
8      m18  Metabolites for 2018 data.frame   96  60
9      m19  Metabolites for 2019 data.frame   94  60
> (my.objects <- addObject(m1819,"Metabolites for 18/19"))
      name      description      class  nrow ncol
1      t18  Transcripts for 2018 data.frame 15242  48
2      t19  Transcripts for 2019 data.frame 15242  32
3     t1819 Transcripts for 18/19 data.frame 15242  80
4    pdata18  Phenodata for 2018 data.frame   48  22
5    pdata19  Phenodata for 2019 data.frame   32  22
6  pdata1819 Phenodata for 2018/19 data.frame   80  22
7      fdata Trans. featuredata 18/19 data.frame 16936  5
8      m18  Metabolites for 2018 data.frame   96  60
9      m19  Metabolites for 2019 data.frame   94  60
10     m1819 Metabolites for 18/19 data.frame  190  60
> (my.objects <- addObject(sm18,"Standardized metabolites 18"))

```

	name	description	class	nrow	ncol
1	t18	Transcripts for 2018	data.frame	15242	48
2	t19	Transcripts for 2019	data.frame	15242	32
3	t1819	Transcripts for 18/19	data.frame	15242	80
4	pdata18	Phenodata for 2018	data.frame	48	22
5	pdata19	Phenodata for 2019	data.frame	32	22
6	pdata1819	Phenodata for 2018/19	data.frame	80	22
7	fdata	Trans. featuredata 18/19	data.frame	16936	5
8	m18	Metabolites for 2018	data.frame	96	60
9	m19	Metabolites for 2019	data.frame	94	60
10	m1819	Metabolites for 18/19	data.frame	190	60
11	sm18	Standardized metabolites 18	matrix	96	55

```
> (my.objects <- addObject(sm19, "Standardized metabolites 19"))
```

	name	description	class	nrow	ncol
1	t18	Transcripts for 2018	data.frame	15242	48
2	t19	Transcripts for 2019	data.frame	15242	32
3	t1819	Transcripts for 18/19	data.frame	15242	80
4	pdata18	Phenodata for 2018	data.frame	48	22
5	pdata19	Phenodata for 2019	data.frame	32	22
6	pdata1819	Phenodata for 2018/19	data.frame	80	22
7	fdata	Trans. featuredata 18/19	data.frame	16936	5
8	m18	Metabolites for 2018	data.frame	96	60
9	m19	Metabolites for 2019	data.frame	94	60
10	m1819	Metabolites for 18/19	data.frame	190	60
11	sm18	Standardized metabolites 18	matrix	96	55
12	sm19	Standardized metabolites 19	matrix	94	55

```
> (my.objects <- addObject(sm1819, "Standardized metabolites 18/19"))
```

	name	description	class	nrow	ncol
1	t18	Transcripts for 2018	data.frame	15242	48
2	t19	Transcripts for 2019	data.frame	15242	32
3	t1819	Transcripts for 18/19	data.frame	15242	80
4	pdata18	Phenodata for 2018	data.frame	48	22
5	pdata19	Phenodata for 2019	data.frame	32	22
6	pdata1819	Phenodata for 2018/19	data.frame	80	22
7	fdata	Trans. featuredata 18/19	data.frame	16936	5
8	m18	Metabolites for 2018	data.frame	96	60
9	m19	Metabolites for 2019	data.frame	94	60
10	m1819	Metabolites for 18/19	data.frame	190	60
11	sm18	Standardized metabolites 18	matrix	96	55
12	sm19	Standardized metabolites 19	matrix	94	55
13	sm1819	Standardized metabolites 18/19	matrix	190	55

```
> (my.objects <- addObject(mfdata, "Metabolites featuredata 18/19"))
```

	name	description	class	nrow	ncol
1	t18	Transcripts for 2018	data.frame	15242	48
2	t19	Transcripts for 2019	data.frame	15242	32
3	t1819	Transcripts for 18/19	data.frame	15242	80
4	pdata18	Phenodata for 2018	data.frame	48	22
5	pdata19	Phenodata for 2019	data.frame	32	22
6	pdata1819	Phenodata for 2018/19	data.frame	80	22
7	fdata	Trans. featuredata 18/19	data.frame	16936	5
8	m18	Metabolites for 2018	data.frame	96	60
9	m19	Metabolites for 2019	data.frame	94	60

```

10     m1819      Metabolites for 18/19 data.frame   190    60
11     sm18       Standardized metabolites 18      matrix    96    55
12     sm19       Standardized metabolites 19      matrix    94    55
13     sm1819    Standardized metabolites 18/19    matrix   190    55
14     mfddata   Metabolites featuredata 18/19    data.frame  55     3

```

```
> (my.objects <- addObject(intgenes, "Names of genes in bins"))
```

```

      name          description      class  nrow ncol
1      t18      Transcripts for 2018 data.frame 15242  48
2      t19      Transcripts for 2019 data.frame 15242  32
3     t1819    Transcripts for 18/19 data.frame 15242  80
4    pdata18      Phenodata for 2018 data.frame   48   22
5    pdata19      Phenodata for 2019 data.frame   32   22
6  pdata1819    Phenodata for 2018/19 data.frame   80   22
7      fdata    Trans. featuredata 18/19 data.frame 16936   5
8      m18      Metabolites for 2018 data.frame   96   60
9      m19      Metabolites for 2019 data.frame   94   60
10     m1819    Metabolites for 18/19 data.frame  190   60
11     sm18     Standardized metabolites 18      matrix   96   55
12     sm19     Standardized metabolites 19      matrix   94   55
13    sm1819   Standardized metabolites 18/19    matrix  190   55
14    mfddata  Metabolites featuredata 18/19    data.frame  55    3
15  intgenes   Names of genes in bins          list     8 <NA>

```

```
> (my.objects <- addObject(intmtbs, "Names of metabolites in bins"))
```

```

      name          description      class  nrow ncol
1      t18      Transcripts for 2018 data.frame 15242  48
2      t19      Transcripts for 2019 data.frame 15242  32
3     t1819    Transcripts for 18/19 data.frame 15242  80
4    pdata18      Phenodata for 2018 data.frame   48   22
5    pdata19      Phenodata for 2019 data.frame   32   22
6  pdata1819    Phenodata for 2018/19 data.frame   80   22
7      fdata    Trans. featuredata 18/19 data.frame 16936   5
8      m18      Metabolites for 2018 data.frame   96   60
9      m19      Metabolites for 2019 data.frame   94   60
10     m1819    Metabolites for 18/19 data.frame  190   60
11     sm18     Standardized metabolites 18      matrix   96   55
12     sm19     Standardized metabolites 19      matrix   94   55
13    sm1819   Standardized metabolites 18/19    matrix  190   55
14    mfddata  Metabolites featuredata 18/19    data.frame  55    3
15  intgenes   Names of genes in bins          list     8 <NA>
16  intmtbs   Names of metabolites in bins     list     8 <NA>

```

```
> my.objects
```

```

      name          description      class  nrow ncol
1      t18      Transcripts for 2018 data.frame 15242  48
2      t19      Transcripts for 2019 data.frame 15242  32
3     t1819    Transcripts for 18/19 data.frame 15242  80
4    pdata18      Phenodata for 2018 data.frame   48   22
5    pdata19      Phenodata for 2019 data.frame   32   22
6  pdata1819    Phenodata for 2018/19 data.frame   80   22
7      fdata    Trans. featuredata 18/19 data.frame 16936   5
8      m18      Metabolites for 2018 data.frame   96   60

```

9	m19	Metabolites for 2019	data.frame	94	60
10	m1819	Metabolites for 18/19	data.frame	190	60
11	sm18	Standardized metabolites 18	matrix	96	55
12	sm19	Standardized metabolites 19	matrix	94	55
13	sm1819	Standardized metabolites 18/19	matrix	190	55
14	mfddata	Metabolites featuredata 18/19	data.frame	55	3
15	intgenes	Names of genes in bins	list	8	<NA>
16	intmtbs	Names of metabolites in bins	list	8	<NA>

List of data objects:

```
> my.objects
```

	name	description	class	nrow	ncol
1	t18	Transcripts for 2018	data.frame	15242	48
2	t19	Transcripts for 2019	data.frame	15242	32
3	t1819	Transcripts for 18/19	data.frame	15242	80
4	pdata18	Phenodata for 2018	data.frame	48	22
5	pdata19	Phenodata for 2019	data.frame	32	22
6	pdata1819	Phenodata for 2018/19	data.frame	80	22
7	fdata	Trans. featuredata 18/19	data.frame	16936	5
8	m18	Metabolites for 2018	data.frame	96	60
9	m19	Metabolites for 2019	data.frame	94	60
10	m1819	Metabolites for 18/19	data.frame	190	60
11	sm18	Standardized metabolites 18	matrix	96	55
12	sm19	Standardized metabolites 19	matrix	94	55
13	sm1819	Standardized metabolites 18/19	matrix	190	55
14	mfddata	Metabolites featuredata 18/19	data.frame	55	3
15	intgenes	Names of genes in bins	list	8	<NA>
16	intmtbs	Names of metabolites in bins	list	8	<NA>

List of interesting genes, based on typification on 50* reports (key: Transcript stats. Typification is extracted from the file in the relevant output directory. To allow manual curation, the info is extracted from the Type column.

```
> tsfn <- getMeta(.adesc, "Transcript stats")
> stat4 <- read.table(file.path(.aroot, tsfn), sep="\t", header=TRUE)
> colnames(stat4)

[1] "X" "A" "Coef.mc"
[4] "Coef.c" "Coef.mf.mc" "Coef.f.c"
[7] "Kcoef.f" "t.mc" "t.c"
[10] "t.mf.mc" "t.f.c" "P.value.mc"
[13] "P.value.c" "P.value.mf.mc" "P.value.f.c"
[16] "P.value.adj.mc" "P.value.adj.c" "P.value.adj.mf.mc"
[19] "P.value.adj.f.c" "F" "F.p.value"
[22] "Type" "interestingC" "interestingFC"
[25] "geneID" "BINCODE" "NAME"
[28] "DESCRIPTION"

> rownames(stat4) <- stat4[,1]
> head(stat4)
```


	X	A	Coef.mc	Coef.c	
Vitvi03g00325	Vitvi03g00325	2.29529737	3.1923214	5.400429	
Vitvi17g00601	Vitvi17g00601	-0.09054575	-1.1866150	4.512791	
Vitvi05g00011	Vitvi05g00011	3.00782492	2.7136299	4.101024	
Vitvi13g02005	Vitvi13g02005	2.33061623	2.9484215	4.078080	
Vitvi02g00695	Vitvi02g00695	-0.45420732	-0.8399601	4.047520	
Vitvi14g01381	Vitvi14g01381	-0.72321946	-1.1530667	3.935431	
	Coef.mf.mc	Coef.f.c	Koef.f	t.mc	t.c
Vitvi03g00325	-1.7940481	-0.29162541	5.108803	4.801209	2.919406
Vitvi17g00601	2.1921385	-0.33971975	4.173071	-2.341540	3.200805
Vitvi05g00011	0.5883901	0.03610049	4.137125	8.336348	4.528348
Vitvi13g02005	-1.2356105	1.22197743	5.300058	3.618330	1.798857
Vitvi02g00695	0.7715056	-1.08372917	2.963791	-1.857328	3.216927
Vitvi14g01381	0.8596946	0.58827898	4.523710	-1.546481	1.897163
	t.mf.mc	t.f.c	P.value.mc	P.value.c	
Vitvi03g00325	-1.9079328	-0.10592036	9.103982e-05	0.0080809276	
Vitvi17g00601	3.0587557	-0.16189100	2.892715e-02	0.0042173372	
Vitvi05g00011	1.2781316	0.02678234	3.604890e-08	0.0001755321	
Vitvi13g02005	-1.0722232	0.36215284	1.571115e-03	0.0861338096	
Vitvi02g00695	1.2062964	-0.57870975	7.705364e-02	0.0040613495	
Vitvi14g01381	0.8153037	0.19053890	1.366212e-01	0.0713562714	
	P.value.mf.mc	P.value.f.c	P.value.adj.mc	P.value.adj.c	
Vitvi03g00325	0.069880852	0.9166302	1.006038e-04	0.28511458	
Vitvi17g00601	0.005867102	0.8729075	3.029669e-02	0.23762225	
Vitvi05g00011	0.214866433	0.9788810	4.329163e-08	0.07644173	
Vitvi13g02005	0.295539873	0.7207826	1.694639e-03	0.58897413	
Vitvi02g00695	0.240844889	0.5688147	7.988693e-02	0.23288283	
Vitvi14g01381	0.423862612	0.8506793	1.407775e-01	0.56306208	
	P.value.adj.mf.mc	P.value.adj.f.c	F		
Vitvi03g00325	0.29606841	0.9999488	9.550446		
Vitvi17g00601	0.05151288	0.9999488	37.060841		
Vitvi05g00011	0.56223076	0.9999488	84.152716		
Vitvi13g02005	0.66176271	0.9999488	6.968467		
Vitvi02g00695	0.59836313	0.9999488	35.455608		
Vitvi14g01381	0.76054331	0.9999488	19.816526		
	F.p.value	Type	interestingC	interestingFC	
Vitvi03g00325	1.356555e-04	Type5	*		
Vitvi17g00601	2.421190e-09	Type5	*		
Vitvi05g00011	8.630059e-13	Type1	*		
Vitvi13g02005	9.348733e-04	Type1.4			
Vitvi02g00695	3.642503e-09		*		
Vitvi14g01381	5.922355e-07	Type1			
	geneID	BINCODE			
Vitvi03g00325	Vitvi03g00325	27.3.25			
Vitvi17g00601	Vitvi17g00601	17.6.3			
Vitvi05g00011	Vitvi05g00011	35.2			
Vitvi13g02005	Vitvi13g02005	29.5.1			
Vitvi02g00695	Vitvi02g00695	20.1			
Vitvi14g01381	Vitvi14g01381	35.2			
Vitvi03g00325	RNA.regulation of transcription.MYB domain transcription factor				
Vitvi17g00601	hormone metabolism.gibberelin.induced-regulated-responsive-act				
Vitvi05g00011	not assigned.u				

```

Vitvi13g02005 protein.degradation.subt
Vitvi02g00695 stress.
Vitvi14g01381 not assigned.u

Vitvi03g00325 RAD-like
Vitvi17g00601 Gibberellin-regulated family prote
Vitvi05g00011 Pollen Ole e 1 allergen and extensin family pro
Vitvi13g02005 Subtilisin-like serine endopeptidase family prote
Vitvi02g00695 Polyketide cyclase/dehydrase and lipid transport superfamily pro
Vitvi14g01381

```

Select interesting genes

```

> filter <- unlist(intgenes) #stat4$Type!="
> length(filter)
[1] 8424
> statig <- stat4[filter,]
> dim(statig)
[1] 8424 28

> (my.objects <- addObject(stat4, "Statistics from transcript analysis (50*.Rnw)
name description class
1 t18 Transcripts for 2018 data.frame
2 t19 Transcripts for 2019 data.frame
3 t1819 Transcripts for 18/19 data.frame
4 pdata18 Phenodata for 2018 data.frame
5 pdata19 Phenodata for 2019 data.frame
6 pdata1819 Phenodata for 2018/19 data.frame
7 fdata Trans. featuredata 18/19 data.frame
8 m18 Metabolites for 2018 data.frame
9 m19 Metabolites for 2019 data.frame
10 m1819 Metabolites for 18/19 data.frame
11 sm18 Standardized metabolites 18 matrix
12 sm19 Standardized metabolites 19 matrix
13 sm1819 Standardized metabolites 18/19 matrix
14 mfdata Metabolites featuredata 18/19 data.frame
15 intgenes Names of genes in bins list
16 intmtbs Names of metabolites in bins list
17 stat4 Statistics from transcript analysis (50*.Rnw) data.frame

nrow ncol
1 15242 48
2 15242 32
3 15242 80
4 48 22
5 32 22
6 80 22
7 16936 5
8 96 60
9 94 60
10 190 60
11 96 55
12 94 55
13 190 55
14 55 3
15 8 <NA>
16 8 <NA>
17 15242 28

```

```
> (my.objects <- addObject(statig, "Statistics for interesting genes"))
```

	name	description	class
1	t18	Transcripts for 2018	data.frame
2	t19	Transcripts for 2019	data.frame
3	t1819	Transcripts for 18/19	data.frame
4	pdata18	Phenodata for 2018	data.frame
5	pdata19	Phenodata for 2019	data.frame
6	pdata1819	Phenodata for 2018/19	data.frame
7	fdata	Trans. featuredata 18/19	data.frame
8	m18	Metabolites for 2018	data.frame
9	m19	Metabolites for 2019	data.frame
10	m1819	Metabolites for 18/19	data.frame
11	sm18	Standardized metabolites 18	matrix
12	sm19	Standardized metabolites 19	matrix
13	sm1819	Standardized metabolites 18/19	matrix
14	mfddata	Metabolites featuredata 18/19	data.frame
15	intgenes	Names of genes in bins	list
16	intmtbs	Names of metabolites in bins	list
17	stat4	Statistics from transcript analysis (50*.Rnw)	data.frame
18	statig	Statistics for interesting genes	data.frame

	nrow	ncol
1	15242	48
2	15242	32
3	15242	80
4	48	22
5	32	22
6	80	22
7	16936	5
8	96	60
9	94	60
10	190	60
11	96	55
12	94	55
13	190	55
14	55	3
15	8	<NA>
16	8	<NA>
17	15242	28
18	8424	28

How many interesting genes are in the interesting bins?

```
> filter <- rownames(statig) %in% unlist(intgenes)
```

```
> sum(filter)
```

```
[1] 7617
```

```
> rownames(statig)[filter]
```

```
[1] "Vitvi01g00025" "Vitvi01g00052" "Vitvi01g00053" "Vitvi01g00064"
[5] "Vitvi01g00681" "Vitvi01g00932" "Vitvi02g00184" "Vitvi02g00250"
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[17] "Vitvi05g00357" "Vitvi05g00442" "Vitvi05g01193" "Vitvi05g01324"
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```

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Vitvi01g00053  2.2.2.6
Vitvi01g00064 Type3.F  2.2.2.2
Vitvi01g00681      35.2

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Vitvi01g00932

2.2.2.1.1

Vitvi01g00025

fructokinase-like protein | Chr1:2

Vitvi01g00052

Glucose-6-phosphate/phosphate translocator-like protein | Chr5:1

Vitvi01g00053

Glucose-6-phosphate/phosphate translocator-like protein | Chr5:1

Vitvi01g00064

Glycosyl transferase%2C family 35 | Chr3:1

Vitvi01g00681

phosphoglucan%2C water dikinase | Chr4:12

Vitvi01g00932

alpha-amylase-like 3 | Chr1:2

3.12 Data overview

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> x <- my.objects
> x$name[1]
[1] "t18"
> for(i in 1:nrow(x)){
+ cat("\n\n----- ", x[i,1], ": ", x[i,2], " -----")
+ print(x[i,])
+ cat("\n")
+ print(head(eval(as.name(x[i,1]))))
+ }

```

----- t18: Transcripts for 2018 -----

name	description	class	nrow	ncol
1 t18	Transcripts for 2018	data.frame	15242	48
	C18_11d_WS1 C18_11d_WS2 C18_11d_WS3 C18_11d_WS4			
Vitvi15g01736	4.6646023 4.109897 4.4482203 4.013955			
Vitvi07g02832	-2.4786650 -1.601359 -1.4274193 -2.066419			
Vitvi07g02830	-1.8506337 -2.218030 -1.0648492 -1.580992			
Vitvi07g02812	-0.5805446 -1.266940 -0.3278837 -1.019113			
Vitvi07g02811	2.2762225 1.823447 1.9550503 1.648742			
Vitvi09g02033	-1.6901691 -2.218030 -2.6498117 -1.449747			
	C18_11d_WW1 C18_11d_WW2 C18_11d_WW3 C18_11d_WW4			
Vitvi15g01736	4.717536 4.154237 4.475261 4.466465			
Vitvi07g02832	-2.608485 -1.801183 -1.886474 -3.583384			
Vitvi07g02830	-1.544354 -1.405255 -1.755229 -1.381750			
Vitvi07g02812	-0.858463 -0.434401 -1.234397 -1.047331			
Vitvi07g02811	2.086902 2.105707 2.134837 1.908469			
Vitvi09g02033	-2.608485 -1.961648 -3.108866 -4.320350			
	C18_34d_WS1 C18_34d_WS2 C18_34d_WS3 C18_34d_WS4			
Vitvi15g01736	4.400333 4.641164 4.4191231 4.534438			
Vitvi07g02832	-5.683146 -2.452000 -4.4274120 -1.876949			
Vitvi07g02830	-2.513221 -2.452000 -4.4274120 -2.263972			
Vitvi07g02812	-2.875791 -2.693008 -2.5529429 -3.157057			
Vitvi07g02811	1.414886 1.409803 0.3970164 1.273993			
Vitvi09g02033	-2.513221 -3.345084 -2.1054839 -1.716484			
	C18_34d_WW1 C18_34d_WW2 C18_34d_WW3 C18_34d_WW4			
Vitvi15g01736	4.924801 5.085690 5.3397350 5.263629			
Vitvi07g02832	-2.827859 -3.130016 -4.6322874 -3.196388			
Vitvi07g02830	-3.190429 -1.545054 -2.1297871 -3.196388			
Vitvi07g02812	-2.827859 -1.182484 -3.4098950 -2.303303			
Vitvi07g02811	1.517916 1.357250 0.9626591 1.344985			
Vitvi09g02033	-4.412821 -3.130016 -2.3103594 -4.418780			
	C18_67d_WS1 C18_67d_WS2 C18_67d_WS3 C18_67d_WS4			
Vitvi15g01736	2.05871733 2.2895026 2.9234830 2.575657			
Vitvi07g02832	-5.85417200 -4.4786818 -4.4659691 -2.468738			
Vitvi07g02830	-2.15373228 -3.2562893 -3.7290035 -2.709746			
Vitvi07g02812	-1.21031581 -2.3632045 -2.3504919 -2.999252			
Vitvi07g02811	0.02847105 0.5362686 0.9824914 1.231702			
Vitvi09g02033	-4.26920950 -3.7417162 -2.5915000 -3.847249			

	C18_67d_WW1	C18_67d_WW2	C18_67d_WW3	C18_67d_WW4
Vitvi15g01736	3.891121	3.040087	4.216897	3.763086
Vitvi07g02832	-2.726982	-4.610965	-2.436913	-3.355855
Vitvi07g02830	-2.726982	-3.873999	-1.493496	-2.075747
Vitvi07g02812	-1.672534	-3.388572	-2.230462	-2.462770
Vitvi07g02811	1.436764	1.552266	1.811015	1.352490
Vitvi09g02033	-2.520531	-2.736495	-3.815424	-2.993285
F18_10d_WS1	F18_10d_WS2	F18_10d_WS3	F18_10d_WS4	
Vitvi15g01736	4.8591409	4.8669435	4.9805904	4.9453953
Vitvi07g02832	2.8881443	2.7032394	2.2757651	2.0553154
Vitvi07g02830	2.8426117	2.4130023	2.2135368	2.0169691
Vitvi07g02812	0.8222302	0.9363895	0.7209384	0.3104447
Vitvi07g02811	3.5853480	3.5007015	3.2965509	3.2141610
Vitvi09g02033	1.0793880	1.0850336	0.6285743	0.8579324
F18_10d_WW1	F18_10d_WW2	F18_10d_WW3	F18_10d_WW4	
Vitvi15g01736	5.705006	4.619238	4.808548	5.276124
Vitvi07g02832	2.747538	3.349854	2.537311	2.621336
Vitvi07g02830	2.287722	3.464855	1.989824	2.321031
Vitvi07g02812	1.073767	1.384521	1.472076	1.016877
Vitvi07g02811	3.818074	4.000053	3.198197	3.501954
Vitvi09g02033	1.073767	1.566041	1.237985	1.268416
F18_34d_WS1	F18_34d_WS2	F18_34d_WS3	F18_34d_WS4	
Vitvi15g01736	4.0647184	3.8504580	5.1418322	4.6269091
Vitvi07g02832	2.2592790	2.1761360	2.3686660	1.2127920
Vitvi07g02830	2.1186455	1.9359657	2.6356368	1.5129655
Vitvi07g02812	0.4316833	-0.6220298	-0.5985972	-0.2055206
Vitvi07g02811	2.9490282	2.7595613	3.1300210	2.4602029
Vitvi09g02033	1.5459405	1.3699329	1.4288835	1.4477479
F18_34d_WW1	F18_34d_WW2	F18_34d_WW3	F18_34d_WW4	
Vitvi15g01736	4.0961725	4.80222138	4.3291524	4.4419027
Vitvi07g02832	1.8816574	1.82887526	1.7895023	1.6490200
Vitvi07g02830	2.1414534	1.62149541	2.1153419	2.1541121
Vitvi07g02812	0.1031253	-0.09471163	0.3053805	0.2855497
Vitvi07g02811	3.0025984	2.73213947	2.8750439	3.0458304
Vitvi09g02033	1.7460934	1.45938991	2.0072243	1.2444445
F18_67d_WS1	F18_67d_WS2	F18_67d_WS3	F18_67d_WS4	
Vitvi15g01736	1.5831276	1.7408584	2.97840555	4.1664059
Vitvi07g02832	1.3093668	1.9241330	2.01456521	2.9813855
Vitvi07g02830	0.8382242	1.4203666	2.18501915	2.8731326
Vitvi07g02812	0.1974739	0.4558154	-0.03878421	-0.0111843
Vitvi07g02811	1.8823168	2.5341865	2.65708760	2.9461961
Vitvi09g02033	1.6066824	1.5040245	1.25520700	0.9986635
F18_67d_WW1	F18_67d_WW2	F18_67d_WW3	F18_67d_WW4	
Vitvi15g01736	3.5580430	3.569471	4.4112089	4.3597440
Vitvi07g02832	2.5563386	2.180583	2.6637430	2.7538173
Vitvi07g02830	2.0265816	2.154972	2.7552617	2.3663511
Vitvi07g02812	0.6773148	0.106805	-0.2413317	-0.1063251
Vitvi07g02811	3.0678390	2.949349	2.9761917	2.9625700
Vitvi09g02033	1.7205680	1.541742	1.6527997	1.3320450

----- t19: Transcripts for 2019 -----

name	description	class	nrow	ncol
2	t19 Transcripts for 2019	data.frame	15242	32
	C19_22d_WS1	C19_22d_WS2	C19_22d_WS3	C19_22d_WS4
Vitvi15g01736	3.18	3.50	3.19	3.61
Vitvi07g02832	-5.05	-3.02	-0.80	-0.02
Vitvi07g02830	-5.05	-5.35	-2.39	-3.99
Vitvi07g02812	-1.35	-0.95	-0.15	0.26
Vitvi07g02811	2.09	2.29	2.34	2.63
Vitvi09g02033	-5.05	-3.76	-2.39	-2.76
	C19_22d_WW1	C19_22d_WW2	C19_22d_WW3	C19_22d_WW4
Vitvi15g01736	5.42	5.35	4.68	4.92
Vitvi07g02832	-1.36	-2.10	-1.15	-3.81
Vitvi07g02830	-2.28	-1.81	-2.07	-2.59
Vitvi07g02812	-1.36	0.16	-0.29	-0.75
Vitvi07g02811	2.85	3.16	3.09	2.89
Vitvi09g02033	-5.45	-3.68	-1.78	-3.81
	C19_44d_WS1	C19_44d_WS2	C19_44d_WS3	C19_44d_WS4
Vitvi15g01736	5.13	5.19	5.71	5.29
Vitvi07g02832	-3.13	-1.28	-3.18	-2.29
Vitvi07g02830	-3.87	-5.53	-5.50	-2.29
Vitvi07g02812	-0.10	-1.01	-0.29	-0.04
Vitvi07g02811	2.11	1.92	2.40	2.29
Vitvi09g02033	-3.87	-3.95	-2.69	-2.00
	C19_44d_WW1	C19_44d_WW2	C19_44d_WW3	C19_44d_WW4
Vitvi15g01736	6.19	5.62	5.62	6.19
Vitvi07g02832	-5.54	-5.31	-3.73	-1.81
Vitvi07g02830	-3.22	-5.31	-3.73	-2.05
Vitvi07g02812	-0.59	-0.45	-1.07	-0.55
Vitvi07g02811	2.44	2.13	2.08	2.26
Vitvi09g02033	-3.22	-3.72	-3.73	-3.18
	F19_22d_WS1	F19_22d_WS2	F19_22d_WS3	F19_22d_WS4
Vitvi15g01736	3.03	3.93	4.24	2.85
Vitvi07g02832	3.35	3.23	3.02	2.98
Vitvi07g02830	2.28	2.39	1.75	1.35
Vitvi07g02812	2.19	1.04	0.89	1.56
Vitvi07g02811	3.84	3.72	3.01	3.43
Vitvi09g02033	2.70	1.61	1.94	2.47
	F19_22d_WW1	F19_22d_WW2	F19_22d_WW3	F19_22d_WW4
Vitvi15g01736	5.00	4.97	4.47	5.51
Vitvi07g02832	3.18	3.04	2.81	3.05
Vitvi07g02830	2.59	2.07	1.81	2.20
Vitvi07g02812	1.72	1.08	1.42	1.35
Vitvi07g02811	3.79	3.43	3.24	3.33
Vitvi09g02033	2.24	1.90	2.08	1.25
	F19_44d_WS1	F19_44d_WS2	F19_44d_WS3	F19_44d_WS4
Vitvi15g01736	5.84	5.83	4.56	5.23
Vitvi07g02832	2.54	3.14	3.01	2.16
Vitvi07g02830	1.13	2.25	2.18	1.90
Vitvi07g02812	0.99	0.84	0.85	0.22
Vitvi07g02811	2.91	3.10	3.18	2.61
Vitvi09g02033	1.21	1.67	2.52	2.21
	F19_44d_WW1	F19_44d_WW2	F19_44d_WW3	F19_44d_WW4

Vitvi15g01736	5.44	6.13	5.96	6.16
Vitvi07g02832	3.84	3.05	3.89	4.02
Vitvi07g02830	2.74	2.79	2.90	2.88
Vitvi07g02812	1.03	0.93	1.49	1.29
Vitvi07g02811	3.32	2.89	3.35	3.26
Vitvi09g02033	1.99	1.14	1.65	1.29

----- t1819: Transcripts for 18/19 -----

name	description	class	nrow	ncol
3 t1819	Transcripts for 18/19	data.frame	15242	80
	C18_11d_WS1	C18_11d_WS2	C18_11d_WS3	C18_11d_WS4
Vitvi15g01736	4.6646023	4.109897	4.4482203	4.013955
Vitvi07g02832	-2.4786650	-1.601359	-1.4274193	-2.066419
Vitvi07g02830	-1.8506337	-2.218030	-1.0648492	-1.580992
Vitvi07g02812	-0.5805446	-1.266940	-0.3278837	-1.019113
Vitvi07g02811	2.2762225	1.823447	1.9550503	1.648742
Vitvi09g02033	-1.6901691	-2.218030	-2.6498117	-1.449747
	C18_11d_WW1	C18_11d_WW2	C18_11d_WW3	C18_11d_WW4
Vitvi15g01736	4.717536	4.154237	4.475261	4.466465
Vitvi07g02832	-2.608485	-1.801183	-1.886474	-3.583384
Vitvi07g02830	-1.544354	-1.405255	-1.755229	-1.381750
Vitvi07g02812	-0.858463	-0.434401	-1.234397	-1.047331
Vitvi07g02811	2.086902	2.105707	2.134837	1.908469
Vitvi09g02033	-2.608485	-1.961648	-3.108866	-4.320350
	C18_34d_WS1	C18_34d_WS2	C18_34d_WS3	C18_34d_WS4
Vitvi15g01736	4.400333	4.641164	4.4191231	4.534438
Vitvi07g02832	-5.683146	-2.452000	-4.4274120	-1.876949
Vitvi07g02830	-2.513221	-2.452000	-4.4274120	-2.263972
Vitvi07g02812	-2.875791	-2.693008	-2.5529429	-3.157057
Vitvi07g02811	1.414886	1.409803	0.3970164	1.273993
Vitvi09g02033	-2.513221	-3.345084	-2.1054839	-1.716484
	C18_34d_WW1	C18_34d_WW2	C18_34d_WW3	C18_34d_WW4
Vitvi15g01736	4.924801	5.085690	5.3397350	5.263629
Vitvi07g02832	-2.827859	-3.130016	-4.6322874	-3.196388
Vitvi07g02830	-3.190429	-1.545054	-2.1297871	-3.196388
Vitvi07g02812	-2.827859	-1.182484	-3.4098950	-2.303303
Vitvi07g02811	1.517916	1.357250	0.9626591	1.344985
Vitvi09g02033	-4.412821	-3.130016	-2.3103594	-4.418780
	C18_67d_WS1	C18_67d_WS2	C18_67d_WS3	C18_67d_WS4
Vitvi15g01736	2.05871733	2.2895026	2.9234830	2.575657
Vitvi07g02832	-5.85417200	-4.4786818	-4.4659691	-2.468738
Vitvi07g02830	-2.15373228	-3.2562893	-3.7290035	-2.709746
Vitvi07g02812	-1.21031581	-2.3632045	-2.3504919	-2.999252
Vitvi07g02811	0.02847105	0.5362686	0.9824914	1.231702
Vitvi09g02033	-4.26920950	-3.7417162	-2.5915000	-3.847249
	C18_67d_WW1	C18_67d_WW2	C18_67d_WW3	C18_67d_WW4
Vitvi15g01736	3.891121	3.040087	4.216897	3.763086
Vitvi07g02832	-2.726982	-4.610965	-2.436913	-3.355855
Vitvi07g02830	-2.726982	-3.873999	-1.493496	-2.075747
Vitvi07g02812	-1.672534	-3.388572	-2.230462	-2.462770

Vitvi07g02811	1.436764	1.552266	1.811015	1.352490
Vitvi09g02033	-2.520531	-2.736495	-3.815424	-2.993285
	F18_10d_WS1	F18_10d_WS2	F18_10d_WS3	F18_10d_WS4
Vitvi15g01736	4.8591409	4.8669435	4.9805904	4.9453953
Vitvi07g02832	2.8881443	2.7032394	2.2757651	2.0553154
Vitvi07g02830	2.8426117	2.4130023	2.2135368	2.0169691
Vitvi07g02812	0.8222302	0.9363895	0.7209384	0.3104447
Vitvi07g02811	3.5853480	3.5007015	3.2965509	3.2141610
Vitvi09g02033	1.0793880	1.0850336	0.6285743	0.8579324
	F18_10d_WW1	F18_10d_WW2	F18_10d_WW3	F18_10d_WW4
Vitvi15g01736	5.705006	4.619238	4.808548	5.276124
Vitvi07g02832	2.747538	3.349854	2.537311	2.621336
Vitvi07g02830	2.287722	3.464855	1.989824	2.321031
Vitvi07g02812	1.073767	1.384521	1.472076	1.016877
Vitvi07g02811	3.818074	4.000053	3.198197	3.501954
Vitvi09g02033	1.073767	1.566041	1.237985	1.268416
	F18_34d_WS1	F18_34d_WS2	F18_34d_WS3	F18_34d_WS4
Vitvi15g01736	4.0647184	3.8504580	5.1418322	4.6269091
Vitvi07g02832	2.2592790	2.1761360	2.3686660	1.2127920
Vitvi07g02830	2.1186455	1.9359657	2.6356368	1.5129655
Vitvi07g02812	0.4316833	-0.6220298	-0.5985972	-0.2055206
Vitvi07g02811	2.9490282	2.7595613	3.1300210	2.4602029
Vitvi09g02033	1.5459405	1.3699329	1.4288835	1.4477479
	F18_34d_WW1	F18_34d_WW2	F18_34d_WW3	F18_34d_WW4
Vitvi15g01736	4.0961725	4.80222138	4.3291524	4.4419027
Vitvi07g02832	1.8816574	1.82887526	1.7895023	1.6490200
Vitvi07g02830	2.1414534	1.62149541	2.1153419	2.1541121
Vitvi07g02812	0.1031253	-0.09471163	0.3053805	0.2855497
Vitvi07g02811	3.0025984	2.73213947	2.8750439	3.0458304
Vitvi09g02033	1.7460934	1.45938991	2.0072243	1.2444445
	F18_67d_WS1	F18_67d_WS2	F18_67d_WS3	F18_67d_WS4
Vitvi15g01736	1.5831276	1.7408584	2.97840555	4.1664059
Vitvi07g02832	1.3093668	1.9241330	2.01456521	2.9813855
Vitvi07g02830	0.8382242	1.4203666	2.18501915	2.8731326
Vitvi07g02812	0.1974739	0.4558154	-0.03878421	-0.0111843
Vitvi07g02811	1.8823168	2.5341865	2.65708760	2.9461961
Vitvi09g02033	1.6066824	1.5040245	1.25520700	0.9986635
	F18_67d_WW1	F18_67d_WW2	F18_67d_WW3	F18_67d_WW4
Vitvi15g01736	3.5580430	3.569471	4.4112089	4.3597440
Vitvi07g02832	2.5563386	2.180583	2.6637430	2.7538173
Vitvi07g02830	2.0265816	2.154972	2.7552617	2.3663511
Vitvi07g02812	0.6773148	0.106805	-0.2413317	-0.1063251
Vitvi07g02811	3.0678390	2.949349	2.9761917	2.9625700
Vitvi09g02033	1.7205680	1.541742	1.6527997	1.3320450
	C19_22d_WS1	C19_22d_WS2	C19_22d_WS3	C19_22d_WS4
Vitvi15g01736	3.18	3.50	3.19	3.61
Vitvi07g02832	-5.05	-3.02	-0.80	-0.02
Vitvi07g02830	-5.05	-5.35	-2.39	-3.99
Vitvi07g02812	-1.35	-0.95	-0.15	0.26
Vitvi07g02811	2.09	2.29	2.34	2.63
Vitvi09g02033	-5.05	-3.76	-2.39	-2.76
	C19_22d_WW1	C19_22d_WW2	C19_22d_WW3	C19_22d_WW4
Vitvi15g01736	5.42	5.35	4.68	4.92

Vitvi07g02832	-1.36	-2.10	-1.15	-3.81
Vitvi07g02830	-2.28	-1.81	-2.07	-2.59
Vitvi07g02812	-1.36	0.16	-0.29	-0.75
Vitvi07g02811	2.85	3.16	3.09	2.89
Vitvi09g02033	-5.45	-3.68	-1.78	-3.81
C19_44d_WS1	C19_44d_WS2	C19_44d_WS3	C19_44d_WS4	
Vitvi15g01736	5.13	5.19	5.71	5.29
Vitvi07g02832	-3.13	-1.28	-3.18	-2.29
Vitvi07g02830	-3.87	-5.53	-5.50	-2.29
Vitvi07g02812	-0.10	-1.01	-0.29	-0.04
Vitvi07g02811	2.11	1.92	2.40	2.29
Vitvi09g02033	-3.87	-3.95	-2.69	-2.00
C19_44d_WW1	C19_44d_WW2	C19_44d_WW3	C19_44d_WW4	
Vitvi15g01736	6.19	5.62	5.62	6.19
Vitvi07g02832	-5.54	-5.31	-3.73	-1.81
Vitvi07g02830	-3.22	-5.31	-3.73	-2.05
Vitvi07g02812	-0.59	-0.45	-1.07	-0.55
Vitvi07g02811	2.44	2.13	2.08	2.26
Vitvi09g02033	-3.22	-3.72	-3.73	-3.18
F19_22d_WS1	F19_22d_WS2	F19_22d_WS3	F19_22d_WS4	
Vitvi15g01736	3.03	3.93	4.24	2.85
Vitvi07g02832	3.35	3.23	3.02	2.98
Vitvi07g02830	2.28	2.39	1.75	1.35
Vitvi07g02812	2.19	1.04	0.89	1.56
Vitvi07g02811	3.84	3.72	3.01	3.43
Vitvi09g02033	2.70	1.61	1.94	2.47
F19_22d_WW1	F19_22d_WW2	F19_22d_WW3	F19_22d_WW4	
Vitvi15g01736	5.00	4.97	4.47	5.51
Vitvi07g02832	3.18	3.04	2.81	3.05
Vitvi07g02830	2.59	2.07	1.81	2.20
Vitvi07g02812	1.72	1.08	1.42	1.35
Vitvi07g02811	3.79	3.43	3.24	3.33
Vitvi09g02033	2.24	1.90	2.08	1.25
F19_44d_WS1	F19_44d_WS2	F19_44d_WS3	F19_44d_WS4	
Vitvi15g01736	5.84	5.83	4.56	5.23
Vitvi07g02832	2.54	3.14	3.01	2.16
Vitvi07g02830	1.13	2.25	2.18	1.90
Vitvi07g02812	0.99	0.84	0.85	0.22
Vitvi07g02811	2.91	3.10	3.18	2.61
Vitvi09g02033	1.21	1.67	2.52	2.21
F19_44d_WW1	F19_44d_WW2	F19_44d_WW3	F19_44d_WW4	
Vitvi15g01736	5.44	6.13	5.96	6.16
Vitvi07g02832	3.84	3.05	3.89	4.02
Vitvi07g02830	2.74	2.79	2.90	2.88
Vitvi07g02812	1.03	0.93	1.49	1.29
Vitvi07g02811	3.32	2.89	3.35	3.26
Vitvi09g02033	1.99	1.14	1.65	1.29

----- pdata18: Phenodata for 2018 -----

name	description	class	nrow	ncol
4	pdata18 Phenodata for 2018	data.frame	48	22

	ID	Variety	Date	variety	year	day
C18_11d_WS1	C18_11d_WS1	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WS2	C18_11d_WS2	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WS3	C18_11d_WS3	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WS4	C18_11d_WS4	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WW1	C18_11d_WW1	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WW2	C18_11d_WW2	Cabernet Volos	12.06.2018	C	18	11
	treat	rep	project.name	species	plant.name	tissue
C18_11d_WS1	WS	1	EnViros Vitis	vinifera	grapevine	leaf
C18_11d_WS2	WS	2	EnViros Vitis	vinifera	grapevine	leaf
C18_11d_WS3	WS	3	EnViros Vitis	vinifera	grapevine	leaf
C18_11d_WS4	WS	4	EnViros Vitis	vinifera	grapevine	leaf
C18_11d_WW1	WW	1	EnViros Vitis	vinifera	grapevine	leaf
C18_11d_WW2	WW	2	EnViros Vitis	vinifera	grapevine	leaf
	health.status	plant.number	growth.location			
C18_11d_WS1	water stress	R1	Udine			
C18_11d_WS2	water stress	R2	Udine			
C18_11d_WS3	water stress	R3	Udine			
C18_11d_WS4	water stress	R4	Udine			
C18_11d_WW1	well watered	R1	Udine			
C18_11d_WW2	well watered	R2	Udine			
	growth.conditions					
C18_11d_WS1	outside					
C18_11d_WS2	outside					
C18_11d_WS3	outside					
C18_11d_WS4	outside					
C18_11d_WW1	outside					
C18_11d_WW2	outside					
C18_11d_WS1	several leaves ground together and stored at -80oC;	an aliquot was				
C18_11d_WS2	several leaves ground together and stored at -80oC;	an aliquot was				
C18_11d_WS3	several leaves ground together and stored at -80oC;	an aliquot was				
C18_11d_WS4	several leaves ground together and stored at -80oC;	an aliquot was				
C18_11d_WW1	several leaves ground together and stored at -80oC;	an aliquot was				
C18_11d_WW2	several leaves ground together and stored at -80oC;	an aliquot was				
	Metabolites.Order	Metabolites.File.Name				
C18_11d_WS1	2	1019Cabernet Volos_6_12_WS1.D				
C18_11d_WS2	4	1019Cabernet Volos_6_12_WS2.D				
C18_11d_WS3	102	1219Cabernet Volos_6_12_WS3.D				
C18_11d_WS4	100	1219Cabernet Volos_6_12_WS4.D				
C18_11d_WW1	6	1019Cabernet Volos_6_12_WW1.D				
C18_11d_WW2	8	1019Cabernet Volos_6_12_WW2.D				
	Metabolites.File	Transcripts.ID	date			
C18_11d_WS1	Cabernet Volos6_12WS	C1_S1	2018-06-12			
C18_11d_WS2	Cabernet Volos6_12WS	C1_S2	2018-06-12			
C18_11d_WS3	Cabernet Volos6_12WS	C1_S3	2018-06-12			
C18_11d_WS4	Cabernet Volos6_12WS	C1_S4	2018-06-12			
C18_11d_WW1	Cabernet Volos6_12WW	C1_W1	2018-06-12			
C18_11d_WW2	Cabernet Volos6_12WW	C1_W2	2018-06-12			

----- pdata19: Phenodata for 2019 -----

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name          description          class nrow ncol
5 pdata19 Phenodata for 2019 data.frame 32 22

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          ID          Variety          Date variety year day
C19_22d_WS1 C19_22d_WS1 Cabernet Volos 26.06.2019      C   19  22
C19_22d_WS2 C19_22d_WS2 Cabernet Volos 26.06.2019      C   19  22
C19_22d_WS3 C19_22d_WS3 Cabernet Volos 26.06.2019      C   19  22
C19_22d_WS4 C19_22d_WS4 Cabernet Volos 26.06.2019      C   19  22
C19_22d_WW1 C19_22d_WW1 Cabernet Volos 26.06.2019      C   19  22
C19_22d_WW2 C19_22d_WW2 Cabernet Volos 26.06.2019      C   19  22

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          treat rep project.name          species plant.name tissue
C19_22d_WS1      WS    1      EnViros Vitis vinifera grapevine leaf
C19_22d_WS2      WS    2      EnViros Vitis vinifera grapevine leaf
C19_22d_WS3      WS    3      EnViros Vitis vinifera grapevine leaf
C19_22d_WS4      WS    4      EnViros Vitis vinifera grapevine leaf
C19_22d_WW1      WW    1      EnViros Vitis vinifera grapevine leaf
C19_22d_WW2      WW    2      EnViros Vitis vinifera grapevine leaf

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          health.status plant.number growth.location
C19_22d_WS1 water stress           R1           Udine
C19_22d_WS2 water stress           R2           Udine
C19_22d_WS3 water stress           R3           Udine
C19_22d_WS4 water stress           R4           Udine
C19_22d_WW1 well watered          R1           Udine
C19_22d_WW2 well watered          R2           Udine

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          growth.conditions
C19_22d_WS1           outside
C19_22d_WS2           outside
C19_22d_WS3           outside
C19_22d_WS4           outside
C19_22d_WW1           outside
C19_22d_WW2           outside

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C19_22d_WS1 several leaves ground together and stored at -80oC; an aliquot was
C19_22d_WS2 several leaves ground together and stored at -80oC; an aliquot was
C19_22d_WS3 several leaves ground together and stored at -80oC; an aliquot was
C19_22d_WS4 several leaves ground together and stored at -80oC; an aliquot was
C19_22d_WW1 several leaves ground together and stored at -80oC; an aliquot was
C19_22d_WW2 several leaves ground together and stored at -80oC; an aliquot was

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          Metabolites.Order          Metabolites.File Name
C19_22d_WS1           2 11120Cabernet_Volos_26_06_2019WS_1.D
C19_22d_WS2           27 11120Cabernet_Volos_26_06_2019WS_2.D
C19_22d_WS3           53 11420Cabernet_Volos_26_06_2019WS_3.D
C19_22d_WS4           79 11420Cabernet_Volos_26_06_2019WS_4.D
C19_22d_WW1           3 11120Cabernet_Volos_26_06_2019WW_1.D
C19_22d_WW2           28 11120Cabernet_Volos_26_06_2019WW_2.D

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          Metabolites.File Transcripts.ID          date
C19_22d_WS1 Cabernet Volos26_06WS           C1S1 2019-06-26
C19_22d_WS2 Cabernet Volos26_06WS           C1S2 2019-06-26
C19_22d_WS3 Cabernet Volos26_06WS           C1S3 2019-06-26
C19_22d_WS4 Cabernet Volos26_06WS           C1S4 2019-06-26
C19_22d_WW1 Cabernet Volos26_06WW           C1W1 2019-06-26
C19_22d_WW2 Cabernet Volos26_06WW           C1W2 2019-06-26

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----- pdata1819: Phenodata for 2018/19 -----

name	description	class	nrow	ncol
6 pdata1819	Phenodata for 2018/19	data.frame	80	22

	ID	Variety	Date	variety	year	day
C18_11d_WS1	C18_11d_WS1	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WS2	C18_11d_WS2	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WS3	C18_11d_WS3	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WS4	C18_11d_WS4	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WW1	C18_11d_WW1	Cabernet Volos	12.06.2018	C	18	11
C18_11d_WW2	C18_11d_WW2	Cabernet Volos	12.06.2018	C	18	11

	treat	rep	project.name	species	plant.name	tissue
C18_11d_WS1	WS	1	EnViros Vitis	vinifera	grapevine	leaf
C18_11d_WS2	WS	2	EnViros Vitis	vinifera	grapevine	leaf
C18_11d_WS3	WS	3	EnViros Vitis	vinifera	grapevine	leaf
C18_11d_WS4	WS	4	EnViros Vitis	vinifera	grapevine	leaf
C18_11d_WW1	WW	1	EnViros Vitis	vinifera	grapevine	leaf
C18_11d_WW2	WW	2	EnViros Vitis	vinifera	grapevine	leaf

	health.status	plant.number	growth.location
C18_11d_WS1	water stress	R1	Udine
C18_11d_WS2	water stress	R2	Udine
C18_11d_WS3	water stress	R3	Udine
C18_11d_WS4	water stress	R4	Udine
C18_11d_WW1	well watered	R1	Udine
C18_11d_WW2	well watered	R2	Udine

	growth.conditions
C18_11d_WS1	outside
C18_11d_WS2	outside
C18_11d_WS3	outside
C18_11d_WS4	outside
C18_11d_WW1	outside
C18_11d_WW2	outside

C18_11d_WS1	several leaves ground together and stored at -80oC; an aliquot was
C18_11d_WS2	several leaves ground together and stored at -80oC; an aliquot was
C18_11d_WS3	several leaves ground together and stored at -80oC; an aliquot was
C18_11d_WS4	several leaves ground together and stored at -80oC; an aliquot was
C18_11d_WW1	several leaves ground together and stored at -80oC; an aliquot was
C18_11d_WW2	several leaves ground together and stored at -80oC; an aliquot was

	Metabolites.Order	Metabolites.File.Name
C18_11d_WS1	2	1019Cabernet Volos_6_12_WS1.D
C18_11d_WS2	4	1019Cabernet Volos_6_12_WS2.D
C18_11d_WS3	102	1219Cabernet Volos_6_12_WS3.D
C18_11d_WS4	100	1219Cabernet Volos_6_12_WS4.D
C18_11d_WW1	6	1019Cabernet Volos_6_12_WW1.D
C18_11d_WW2	8	1019Cabernet Volos_6_12_WW2.D

	Metabolites.File	Transcripts.ID	date
C18_11d_WS1	Cabernet Volos6_12WS	C1_S1	2018-06-12
C18_11d_WS2	Cabernet Volos6_12WS	C1_S2	2018-06-12
C18_11d_WS3	Cabernet Volos6_12WS	C1_S3	2018-06-12

C18_11d_WS4	Cabernet Volos6_12WS	C1_S4	2018-06-12
C18_11d_WW1	Cabernet Volos6_12WW	C1_W1	2018-06-12
C18_11d_WW2	Cabernet Volos6_12WW	C1_W2	2018-06-12

----- fdata: Trans. featuredata 18/19 -----

name	description	class	nrow	ncol
7 fdata	Trans. featuredata 18/19	data.frame	16936	5

YEAR	geneID	BINCODE	NAME
1 2018	Vitvi15g01736	26.9	misc.glutathione S transferases
2 2018	Vitvi07g02832	35.2	not assigned.unknown
3 2018	Vitvi07g02830	35.2	not assigned.unknown
4 2018	Vitvi07g02812	35.2	not assigned.unknown
5 2018	Vitvi07g02811	35.2	not assigned.unknown
6 2018	Vitvi09g02033	35.2	not assigned.unknown

1	glutathione S-transferase tau 7 Chr2:12618111-1261887
2	Disease resistance protein (TIR-NBS-LRR class) family Chr4:7197325-7201393
3	
4	
5	
6	Disease resistance protein (CC-NBS-LRR class) family Chr1:4145374-414768

----- m18: Metabolites for 2018 -----

name	description	class	nrow	ncol
8 m18	Metabolites for 2018	data.frame	96	60

	ID	Variety	Date	Treat	Rep	Alanine
C18_11d_WS1	C18_11d_WS1	Cabernet Volos	12.06.2018	WS	1	0.133
C18_11d_WS2	C18_11d_WS2	Cabernet Volos	12.06.2018	WS	2	0.115
C18_11d_WS3	C18_11d_WS3	Cabernet Volos	12.06.2018	WS	3	0.624
C18_11d_WS4	C18_11d_WS4	Cabernet Volos	12.06.2018	WS	4	0.649
C18_11d_WW1	C18_11d_WW1	Cabernet Volos	12.06.2018	WW	1	0.136
C18_11d_WW2	C18_11d_WW2	Cabernet Volos	12.06.2018	WW	2	0.230
	Arabinose	Ascorbic.acid	Aspartic.acid	GABA		
C18_11d_WS1	0.116	0.885	0.882	1.950		
C18_11d_WS2	0.120	1.147	0.613	1.054		
C18_11d_WS3	0.094	1.080	0.715	1.401		
C18_11d_WS4	0.112	1.587	0.827	1.705		
C18_11d_WW1	0.160	0.769	0.549	1.929		
C18_11d_WW2	0.147	0.903	0.625	1.513		
	Trans.Caffeic.acid	Catechin	Citric.acid	Erythronic.acid		
C18_11d_WS1	8.751	2.828	2.236	1.696		
C18_11d_WS2	9.164	3.595	2.152	2.039		
C18_11d_WS3	7.039	2.059	1.996	1.791		
C18_11d_WS4	7.379	1.880	2.619	1.587		
C18_11d_WW1	8.197	1.930	2.290	1.999		
C18_11d_WW2	7.093	1.419	2.056	1.301		
	Ethanolamine	Fructose	Fructose.6.phosphate	Fumaric.acid		

C18_11d_WS1	1.052	27.962		0.037	0.683
C18_11d_WS2	0.865	22.260		0.035	1.069
C18_11d_WS3	1.335	14.477		0.058	1.907
C18_11d_WS4	0.901	16.651		0.079	2.009
C18_11d_WW1	0.861	22.621		0.033	0.760
C18_11d_WW2	0.805	20.318		0.089	1.094
Galactinol Galactose Gallic.acid Gluconic.acid					
C18_11d_WS1	16.908	8.455	1.002	3.640	
C18_11d_WS2	13.660	8.218	0.958	3.911	
C18_11d_WS3	13.524	4.445	0.936	3.079	
C18_11d_WS4	14.463	6.358	0.905	3.421	
C18_11d_WW1	15.754	6.071	1.143	3.133	
C18_11d_WW2	13.891	5.467	0.917	2.805	
Glucopyranose..H2O. Glucose Glutamic.acid Glyceric.acid					
C18_11d_WS1		0.143	43.309	0.491	7.540
C18_11d_WS2		0.155	41.412	0.298	6.082
C18_11d_WS3		0.178	31.305	0.140	5.876
C18_11d_WS4		0.153	40.981	0.202	7.596
C18_11d_WW1		0.169	37.135	0.303	6.008
C18_11d_WW2		0.156	35.269	0.263	5.638
Glycine Hydroquinone Myo.Inostol Isoleucine Leucine					
C18_11d_WS1	0.742	0.161	91.071	0.055	0.022
C18_11d_WS2	0.727	0.208	89.166	0.054	0.027
C18_11d_WS3	0.913	0.099	97.202	0.072	0.033
C18_11d_WS4	0.742	0.176	114.143	0.150	0.079
C18_11d_WW1	1.031	0.101	85.266	0.146	0.077
C18_11d_WW2	0.847	0.161	90.745	0.096	0.050
Lyxonic.acid Maleic.acid Malic.acid Malonic.acid					
C18_11d_WS1	1.028	1.093	66.326	0.166	
C18_11d_WS2	1.187	1.486	73.855	0.140	
C18_11d_WS3	1.237	1.108	62.877	0.183	
C18_11d_WS4	1.020	1.084	66.638	0.233	
C18_11d_WW1	1.205	0.879	53.894	0.156	
C18_11d_WW2	1.096	1.202	51.917	0.170	
Mannose.6.phosphate Melibiose Phenylalanine					
C18_11d_WS1		0.088	0.227	0.095	
C18_11d_WS2		0.102	0.204	0.116	
C18_11d_WS3		0.134	0.069	0.103	
C18_11d_WS4		0.174	0.092	0.203	
C18_11d_WW1		0.082	0.178	0.104	
C18_11d_WW2		0.198	0.164	0.179	
Phosphoric.acid Proline Putrescine Pyroglutamic.acid					
C18_11d_WS1	23.244	0.011	0.054	5.790	
C18_11d_WS2	17.190	0.013	0.046	4.354	
C18_11d_WS3	18.909	0.006	0.104	8.169	
C18_11d_WS4	23.579	0.008	0.111	10.445	
C18_11d_WW1	31.264	0.006	0.066	4.966	
C18_11d_WW2	31.233	0.008	0.058	5.408	
Quinic.acid X3.caffeoylquinic.acid Raffinose Rhamnose					
C18_11d_WS1	1.615		0.677	1.117	0.603
C18_11d_WS2	1.304		0.635	1.544	0.472
C18_11d_WS3	1.254		0.378	0.961	0.516
C18_11d_WS4	1.106		0.423	1.164	0.634

C18_11d_WW1	1.985		0.576	0.870	0.534
C18_11d_WW2	1.513		0.462	1.001	0.747
	Ribonic.acid	Ribose	Serine	Shikimic.acid	Succinic.acid
C18_11d_WS1	0.270	0.151	0.997	23.883	0.139
C18_11d_WS2	0.237	0.152	0.609	13.037	0.133
C18_11d_WS3	0.361	0.162	0.895	14.551	0.089
C18_11d_WS4	0.293	0.154	1.116	14.364	0.106
C18_11d_WW1	0.326	0.164	0.465	23.607	0.140
C18_11d_WW2	0.326	0.164	0.551	18.038	0.119
	Sucrose	Tartaric.acid	Threitol	Threonic.acid	
C18_11d_WS1	96.247	58.764	0.025	2.632	
C18_11d_WS2	101.273	36.403	0.018	2.741	
C18_11d_WS3	114.777	30.620	0.027	2.645	
C18_11d_WS4	122.686	52.311	0.033	2.387	
C18_11d_WW1	99.984	47.986	0.032	3.639	
C18_11d_WW2	105.411	44.912	0.031	3.759	
	Threonolactone	Threonine	Uracil	Valine	Xylose
C18_11d_WS1	0.145	0.244	0.011	0.267	0.297
C18_11d_WS2	0.121	0.187	0.017	0.259	0.231
C18_11d_WS3	0.086	0.216	0.015	0.360	0.251
C18_11d_WS4	0.129	0.274	0.018	0.684	0.278
C18_11d_WW1	0.160	0.184	0.013	0.582	0.266
C18_11d_WW2	0.334	0.212	0.018	0.319	0.342

----- m19: Metabolites for 2019 -----

name	description	class	nrow	ncol
9	m19 Metabolites for 2019	data.frame	94	60

	ID	Variety	Date	Treat	Rep	Alanine
C19_22d_WS1	C19_22d_WS1	Cabernet Volos	26.06.2019	WS	1	1.44
C19_22d_WS2	C19_22d_WS2	Cabernet Volos	26.06.2019	WS	2	5.97
C19_22d_WS3	C19_22d_WS3	Cabernet Volos	26.06.2019	WS	3	1.96
C19_22d_WS4	C19_22d_WS4	Cabernet Volos	26.06.2019	WS	4	2.98
C19_22d_WW1	C19_22d_WW1	Cabernet Volos	26.06.2019	WW	1	1.11
C19_22d_WW2	C19_22d_WW2	Cabernet Volos	26.06.2019	WW	2	3.63
	Arabinose	Ascorbic.acid	Aspartic.acid	GABA		
C19_22d_WS1	13.26	3.60	14.69	3.20		
C19_22d_WS2	15.15	11.03	10.00	8.23		
C19_22d_WS3	10.67	3.34	12.62	9.39		
C19_22d_WS4	12.01	2.39	16.12	6.30		
C19_22d_WW1	15.16	2.86	15.53	10.23		
C19_22d_WW2	13.30	4.67	8.74	7.91		
	Trans.Caffeic.acid	Catechin	Citric.acid	Erythronic.acid		
C19_22d_WS1	6.18	13.63	5.35	5.97		
C19_22d_WS2	12.17	17.04	6.65	6.24		
C19_22d_WS3	9.24	3.24	8.05	5.03		
C19_22d_WS4	14.66	2.28	8.26	4.81		
C19_22d_WW1	5.00	15.94	8.76	5.13		
C19_22d_WW2	8.99	9.05	7.03	4.67		
	Ethanolamine	Fructose	Fructose.6.phosphate	Fumaric.acid		
C19_22d_WS1	6.68	11.71	5.83	5.54		

C19_22d_WS2	10.69	12.24		7.71	8.39
C19_22d_WS3	8.34	10.82		8.21	4.05
C19_22d_WS4	5.94	10.75		9.73	11.80
C19_22d_WW1	8.68	14.73		13.54	3.79
C19_22d_WW2	6.65	15.20		21.62	5.53
Galactinol Galactose Gallic.acid Gluconic.acid					
C19_22d_WS1	18.37	12.24	8.35	10.27	
C19_22d_WS2	14.63	13.59	6.97	11.45	
C19_22d_WS3	19.87	11.75	4.26	9.88	
C19_22d_WS4	12.76	12.42	11.59	8.80	
C19_22d_WW1	11.17	15.85	8.83	8.55	
C19_22d_WW2	12.18	14.85	10.23	7.69	
Glucopyranose..H2O. Glucose Glutamic.acid Glyceric.acid					
C19_22d_WS1		7.92	11.29	40.46	6.60
C19_22d_WS2		13.10	12.31	14.61	7.73
C19_22d_WS3		10.09	11.09	4.98	9.26
C19_22d_WS4		9.05	11.56	7.01	8.10
C19_22d_WW1		6.55	13.44	22.22	6.49
C19_22d_WW2		8.59	12.87	10.24	5.66
Glycine Hydroquinone Myo.Inostol Isoleucine Leucine					
C19_22d_WS1	7.71	5.76	9.85	18.14	17.99
C19_22d_WS2	8.66	9.66	10.15	14.90	14.61
C19_22d_WS3	16.27	8.18	9.06	13.60	13.96
C19_22d_WS4	16.39	6.41	9.00	19.76	19.64
C19_22d_WW1	9.61	5.49	9.09	24.93	24.59
C19_22d_WW2	22.09	7.93	8.50	6.72	6.62
Lyxonic.acid Maleic.acid Malic.acid Malonic.acid					
C19_22d_WS1	10.61	10.45	9.97	3.38	
C19_22d_WS2	11.95	6.52	9.34	9.58	
C19_22d_WS3	9.29	5.28	9.45	7.40	
C19_22d_WS4	9.88	10.07	8.48	9.60	
C19_22d_WW1	9.79	5.66	10.31	3.53	
C19_22d_WW2	10.41	4.09	9.20	7.04	
Mannose.6.phosphate Melibiose Phenylalanine					
C19_22d_WS1		5.06	6.44	10.84	
C19_22d_WS2		6.60	6.69	9.08	
C19_22d_WS3		8.72	5.63	15.50	
C19_22d_WS4		9.13	4.52	19.50	
C19_22d_WW1		10.64	6.69	15.80	
C19_22d_WW2		14.61	6.01	7.75	
Phosphoric.acid Proline Putrescine Pyroglutamic.acid					
C19_22d_WS1	8.37	42.32	9.77	6.61	
C19_22d_WS2	10.42	29.09	17.21	11.75	
C19_22d_WS3	8.93	27.38	11.09	12.15	
C19_22d_WS4	10.84	24.39	11.07	13.65	
C19_22d_WW1	16.18	37.14	26.02	4.01	
C19_22d_WW2	17.27	2.09	6.03	6.17	
Quinic.acid X3.caffeoylquinic.acid Raffinose Rhamnose					
C19_22d_WS1	15.57		22.62	12.69	8.65
C19_22d_WS2	17.79		26.34	11.81	7.16
C19_22d_WS3	16.01		21.82	12.97	7.92
C19_22d_WS4	13.27		27.36	8.93	7.18
C19_22d_WW1	13.87		23.22	9.27	8.09

C19_22d_WW2	14.48			21.36	9.21	7.63
	Ribonic.acid	Ribose	Serine	Shikimic.acid	Succinic.acid	
C19_22d_WS1	10.72	8.67	7.52	25.77		14.82
C19_22d_WS2	12.01	10.68	10.27	27.03		14.23
C19_22d_WS3	9.29	8.24	23.26	26.88		12.09
C19_22d_WS4	9.88	8.45	22.24	22.56		8.96
C19_22d_WW1	9.90	8.22	10.73	25.90		11.32
C19_22d_WW2	10.44	9.96	7.29	25.63		10.23
	Sucrose	Tartaric.acid	Threitol	Threonic.acid		
C19_22d_WS1	9.14		5.49	8.74		6.89
C19_22d_WS2	9.58		7.20	7.03		5.86
C19_22d_WS3	8.30		7.11	13.37		4.25
C19_22d_WS4	9.41		7.89	9.49		2.89
C19_22d_WW1	9.33		6.96	7.26		3.78
C19_22d_WW2	9.51		7.55	12.28		5.13
	Threonolactone	Threonine	Uracil	Valine	Xylose	
C19_22d_WS1	7.24		11.78	6.22	24.43	9.58
C19_22d_WS2	6.31		13.48	9.03	18.07	11.19
C19_22d_WS3	4.64		9.94	7.75	24.16	8.97
C19_22d_WS4	3.07		7.60	13.93	18.60	9.80
C19_22d_WW1	6.32		7.58	9.59	26.02	11.86
C19_22d_WW2	7.49		7.14	8.35	9.35	10.21

----- m1819: Metabolites for 18/19 -----

name description class nrow ncol
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	ID	Variety	Date	Treat	Rep	Alanine
C18_11d_WS1	C18_11d_WS1	Cabernet Volos	12.06.2018	WS	1	0.133
C18_11d_WS2	C18_11d_WS2	Cabernet Volos	12.06.2018	WS	2	0.115
C18_11d_WS3	C18_11d_WS3	Cabernet Volos	12.06.2018	WS	3	0.624
C18_11d_WS4	C18_11d_WS4	Cabernet Volos	12.06.2018	WS	4	0.649
C18_11d_WW1	C18_11d_WW1	Cabernet Volos	12.06.2018	WW	1	0.136
C18_11d_WW2	C18_11d_WW2	Cabernet Volos	12.06.2018	WW	2	0.230
	Arabinose	Ascorbic.acid	Aspartic.acid	GABA		
C18_11d_WS1	0.116	0.885	0.882	1.950		
C18_11d_WS2	0.120	1.147	0.613	1.054		
C18_11d_WS3	0.094	1.080	0.715	1.401		
C18_11d_WS4	0.112	1.587	0.827	1.705		
C18_11d_WW1	0.160	0.769	0.549	1.929		
C18_11d_WW2	0.147	0.903	0.625	1.513		
	Trans.Caffeic.acid	Catechin	Citric.acid	Erythronic.acid		
C18_11d_WS1	8.751	2.828	2.236			1.696
C18_11d_WS2	9.164	3.595	2.152			2.039
C18_11d_WS3	7.039	2.059	1.996			1.791
C18_11d_WS4	7.379	1.880	2.619			1.587
C18_11d_WW1	8.197	1.930	2.290			1.999
C18_11d_WW2	7.093	1.419	2.056			1.301
	Ethanolamine	Fructose	Fructose.6.phosphate	Fumaric.acid		
C18_11d_WS1	1.052	27.962		0.037		0.683
C18_11d_WS2	0.865	22.260		0.035		1.069

C18_11d_WS3	1.335	14.477		0.058	1.907
C18_11d_WS4	0.901	16.651		0.079	2.009
C18_11d_WW1	0.861	22.621		0.033	0.760
C18_11d_WW2	0.805	20.318		0.089	1.094
Galactinol Galactose Gallic.acid Gluconic.acid					
C18_11d_WS1	16.908	8.455	1.002		3.640
C18_11d_WS2	13.660	8.218	0.958		3.911
C18_11d_WS3	13.524	4.445	0.936		3.079
C18_11d_WS4	14.463	6.358	0.905		3.421
C18_11d_WW1	15.754	6.071	1.143		3.133
C18_11d_WW2	13.891	5.467	0.917		2.805
Glucopyranose..H2O. Glucose Glutamic.acid Glyceric.acid					
C18_11d_WS1		0.143	43.309	0.491	7.540
C18_11d_WS2		0.155	41.412	0.298	6.082
C18_11d_WS3		0.178	31.305	0.140	5.876
C18_11d_WS4		0.153	40.981	0.202	7.596
C18_11d_WW1		0.169	37.135	0.303	6.008
C18_11d_WW2		0.156	35.269	0.263	5.638
Glycine Hydroquinone Myo.Inostol Isoleucine Leucine					
C18_11d_WS1	0.742	0.161	91.071	0.055	0.022
C18_11d_WS2	0.727	0.208	89.166	0.054	0.027
C18_11d_WS3	0.913	0.099	97.202	0.072	0.033
C18_11d_WS4	0.742	0.176	114.143	0.150	0.079
C18_11d_WW1	1.031	0.101	85.266	0.146	0.077
C18_11d_WW2	0.847	0.161	90.745	0.096	0.050
Lyxonic.acid Maleic.acid Malic.acid Malonic.acid					
C18_11d_WS1	1.028	1.093	66.326		0.166
C18_11d_WS2	1.187	1.486	73.855		0.140
C18_11d_WS3	1.237	1.108	62.877		0.183
C18_11d_WS4	1.020	1.084	66.638		0.233
C18_11d_WW1	1.205	0.879	53.894		0.156
C18_11d_WW2	1.096	1.202	51.917		0.170
Mannose.6.phosphate Melibiose Phenylalanine					
C18_11d_WS1		0.088	0.227		0.095
C18_11d_WS2		0.102	0.204		0.116
C18_11d_WS3		0.134	0.069		0.103
C18_11d_WS4		0.174	0.092		0.203
C18_11d_WW1		0.082	0.178		0.104
C18_11d_WW2		0.198	0.164		0.179
Phosphoric.acid Proline Putrescine Pyroglutamic.acid					
C18_11d_WS1	23.244	0.011	0.054		5.790
C18_11d_WS2	17.190	0.013	0.046		4.354
C18_11d_WS3	18.909	0.006	0.104		8.169
C18_11d_WS4	23.579	0.008	0.111		10.445
C18_11d_WW1	31.264	0.006	0.066		4.966
C18_11d_WW2	31.233	0.008	0.058		5.408
Quinic.acid X3.caffeoylquinic.acid Raffinose Rhamnose					
C18_11d_WS1	1.615		0.677	1.117	0.603
C18_11d_WS2	1.304		0.635	1.544	0.472
C18_11d_WS3	1.254		0.378	0.961	0.516
C18_11d_WS4	1.106		0.423	1.164	0.634
C18_11d_WW1	1.985		0.576	0.870	0.534
C18_11d_WW2	1.513		0.462	1.001	0.747

	Ribonic.acid	Ribose	Serine	Shikimic.acid	Succinic.acid
C18_11d_WS1	0.270	0.151	0.997	23.883	0.139
C18_11d_WS2	0.237	0.152	0.609	13.037	0.133
C18_11d_WS3	0.361	0.162	0.895	14.551	0.089
C18_11d_WS4	0.293	0.154	1.116	14.364	0.106
C18_11d_WW1	0.326	0.164	0.465	23.607	0.140
C18_11d_WW2	0.326	0.164	0.551	18.038	0.119
	Sucrose	Tartaric.acid	Threitol	Threonic.acid	
C18_11d_WS1	96.247	58.764	0.025	2.632	
C18_11d_WS2	101.273	36.403	0.018	2.741	
C18_11d_WS3	114.777	30.620	0.027	2.645	
C18_11d_WS4	122.686	52.311	0.033	2.387	
C18_11d_WW1	99.984	47.986	0.032	3.639	
C18_11d_WW2	105.411	44.912	0.031	3.759	
	Threonolactone	Threonine	Uracil	Valine	Xylose
C18_11d_WS1	0.145	0.244	0.011	0.267	0.297
C18_11d_WS2	0.121	0.187	0.017	0.259	0.231
C18_11d_WS3	0.086	0.216	0.015	0.360	0.251
C18_11d_WS4	0.129	0.274	0.018	0.684	0.278
C18_11d_WW1	0.160	0.184	0.013	0.582	0.266
C18_11d_WW2	0.334	0.212	0.018	0.319	0.342

----- sm18: Standardized metabolites 18 -----

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name                description  class nrow ncol
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	Alanine	Arabinose	Ascorbic.acid	Aspartic.acid
C18_11d_WS1	-1.3216743	0.81081282	0.9519808	0.2446827
C18_11d_WS2	-1.5850312	0.93631495	1.3802355	-0.3672711
C18_11d_WS3	1.4778432	0.03230802	1.2808356	-0.1083831
C18_11d_WS4	1.5489853	0.68090622	1.9164602	0.1363838
C18_11d_WW1	-1.2812775	2.00130194	0.7199549	-0.5527383
C18_11d_WW2	-0.3297096	1.68759346	0.9852330	-0.3346628
	GABA	Trans.Caffeic.acid	Catechin	Citric.acid
C18_11d_WS1	-0.2519906	0.80770699	1.3115059	-0.3119820
C18_11d_WS2	-0.7511261	0.98244100	1.4698917	-0.4326158
C18_11d_WS3	-0.5202380	-0.01718941	1.1020518	-0.6696953
C18_11d_WS4	-0.3609178	0.16155056	1.0420246	0.1861175
C18_11d_WW1	-0.2607749	0.55990013	1.0593488	-0.2368019
C18_11d_WW2	-0.4578432	0.01176799	0.8563507	-0.5763877
	Erythronic.acid	Ethanolamine	Fructose	
C18_11d_WS1	-0.8746026	-0.53998192	0.99215264	
C18_11d_WS2	-0.3362622	-0.95018249	0.62627555	
C18_11d_WS3	-0.7153057	-0.04066661	-0.06395835	
C18_11d_WS4	-1.0687552	-0.86472198	0.16050289	
C18_11d_WW1	-0.3941699	-0.95989684	0.65208504	
C18_11d_WW2	-1.6495500	-1.10084850	0.47982506	
	Fructose.6.phosphate	Fumaric.acid	Galactinol	Galactose
C18_11d_WS1	0.202208256	-2.3255100	0.393038165	1.5647978
C18_11d_WS2	0.107588328	-0.9958129	0.016625527	1.5087783
C18_11d_WS3	0.967623882	0.7221923	-0.001030863	0.2979007

C18_11d_WS4	1.493772779	0.8768515	0.117421853	1.0031588
C18_11d_WW1	0.007399398	-2.0084384	0.268295050	0.9121470
C18_11d_WW2	1.696717518	-0.9271973	0.046216383	0.7056666
	Gallic.acid	Gluconic.acid	Glucopyranose..H2O.	Glucose
C18_11d_WS1	0.6552556	0.7966034	-0.26091191	1.3799156
C18_11d_WS2	0.5208686	0.9581338	0.01215372	1.2776290
C18_11d_WS3	0.4513421	0.4200960	0.48101331	0.6386647
C18_11d_WS4	0.3505476	0.6570243	-0.03185639	1.2537365
C18_11d_WW1	1.0492647	0.4592050	0.30518953	1.0286806
C18_11d_WW2	0.3899686	0.2104459	0.03394629	0.9109430
	Glutamic.acid	Glyceric.acid	Glycine	Hydroquinone
C18_11d_WS1	0.9684981	0.9242754	0.9424986	1.6367642
C18_11d_WS2	0.2974240	0.5873407	0.8985881	2.1123854
C18_11d_WS3	-0.7178217	0.5333133	1.3883966	0.7337700
C18_11d_WS4	-0.2251164	0.9358776	0.9424986	1.8021784
C18_11d_WW1	0.3197855	0.5681464	1.6497358	0.7709097
C18_11d_WW2	0.1295187	0.4684835	1.2270649	1.6367642
	Myo.Inostol	Isoleucine	Leucine	Lyxonic.acid
C18_11d_WS1	1.0265053	-0.9370969	-1.28454542	0.05849233
C18_11d_WS2	0.9470041	-0.9635587	-1.06278206	0.71877226
C18_11d_WS3	1.2715262	-0.5486835	-0.84548411	0.90820545
C18_11d_WS4	1.8757337	0.5097963	0.09978663	0.02262333
C18_11d_WW1	0.7788075	0.4708175	0.07201951	0.78787203
C18_11d_WW2	1.0130190	-0.1338082	-0.39553971	0.35256857
	Maleic.acid	Malic.acid	Malonic.acid	Mannose.6.phosphate
C18_11d_WS1	-0.07418252	0.4657714	0.1067572	0.18623208
C18_11d_WS2	0.86317474	0.8806769	-0.4627324	0.44188513
C18_11d_WS3	-0.03258706	0.2597052	0.4327085	0.91439372
C18_11d_WS4	-0.09941469	0.4838808	1.2402534	1.36672608
C18_11d_WW1	-0.73913172	-0.3351734	-0.1009585	0.06394756
C18_11d_WW2	0.21591145	-0.4793880	0.1863597	1.59047486
	Melibiose	Phenylalanine	Phosphoric.acid	Proline
C18_11d_WS1	0.10278882	-0.68751145	0.605441967	-0.26080192
C18_11d_WS2	-0.08996645	-0.49153302	-0.009926619	-0.08957458
C18_11d_WS3	-2.04587081	-0.60817138	0.184462362	-0.88207988
C18_11d_WS4	-1.52680105	0.05761733	0.634626673	-0.58721109
C18_11d_WW1	-0.33596066	-0.59869016	1.210003081	-0.88207988
C18_11d_WW2	-0.48376515	-0.06584986	1.207979760	-0.58721109
	Putrescine	Pyroglutamic.acid	Quinic.acid	
C18_11d_WS1	-0.7704912	-0.06071954	1.2968180	
C18_11d_WS2	-1.0928669	-0.97572564	1.0387640	
C18_11d_WS3	0.5472319	1.04425188	0.9915949	
C18_11d_WS4	0.6781973	1.83322698	0.8400808	
C18_11d_WW1	-0.3670343	-0.55353059	1.5456855	
C18_11d_WW2	-0.6268200	-0.27982021	1.2181097	
	X3.caffeoylquinic.acid	Raffinose	Rhamnose	
C18_11d_WS1		1.195696	0.24980597	-0.3189756
C18_11d_WS2		1.170472	0.66225216	-0.8333650
C18_11d_WS3		0.966176	0.05815485	-0.6461897
C18_11d_WS4		1.010474	0.30231675	-0.2136950
C18_11d_WW1		1.132066	-0.06858858	-0.5741800
C18_11d_WW2		1.045208	0.11011083	0.1307519
	Ribonic.acid	Ribose	Serine	Shikimic.acid

C18_11d_WS1	0.4479829	0.5847252	1.5289606	1.883613
C18_11d_WS2	-0.2222956	0.6177648	0.6076439	1.186979
C18_11d_WS3	1.9414142	0.9366934	1.3272393	1.313410
C18_11d_WS4	0.8683185	0.6831970	1.7397064	1.298525
C18_11d_WW1	1.4170629	0.9981112	0.1034093	1.870237
C18_11d_WW2	1.4170629	0.9981112	0.4205826	1.560614
	Succinic.acid	Sucrose	Tartaric.acid	Threitol
C18_11d_WS1	1.12378351	0.9727693	0.88130394	-0.1755182
C18_11d_WS2	1.00592868	1.1753149	-0.05506813	-1.0908375
C18_11d_WS3	-0.06702313	1.6733871	-0.39333848	0.0389203
C18_11d_WS4	0.39986495	1.9385446	0.65385286	0.5980542
C18_11d_WW1	1.14293013	1.1243437	0.48511174	0.5123143
C18_11d_WW2	0.70885137	1.3346676	0.35565954	0.4238521
	Threonic.acid	Threonolactone	Threonine	Uracil
C18_11d_WS1	0.7865698	0.79978675	0.4644569	-1.5690944
C18_11d_WS2	0.8761986	0.52235743	-0.2077896	0.1557937
C18_11d_WS3	0.7974524	-0.00115696	0.1564808	-0.3401481
C18_11d_WS4	0.5707593	0.62051830	0.7574499	0.3822761
C18_11d_WW1	1.5021302	0.95071900	-0.2486532	-0.9071655
C18_11d_WW2	1.5737913	2.07913320	0.1092518	0.3822761
	Valine	Xylose		
C18_11d_WS1	-0.33710795	1.5782722		
C18_11d_WS2	-0.38696882	0.5869536		
C18_11d_WS3	0.15273094	0.9144890		
C18_11d_WS4	1.20476153	1.3174949		
C18_11d_WW1	0.94007550	1.1434431		
C18_11d_WW2	-0.04545139	2.1347616		

----- sm19: Standardized metabolites 19 -----

name	description	class	nrow	ncol
12 sm19	Standardized metabolites 19 matrix		94	55

	Alanine	Arabinose	Ascorbic.acid	Aspartic.acid
C19_22d_WS1	-1.53885853	1.0634664	-0.9440966	1.04172567
C19_22d_WS2	0.85684023	1.7818921	0.9077321	0.25315899
C19_22d_WS3	-1.01948914	-0.1082187	-1.0680767	0.73029456
C19_22d_WS4	-0.31367073	0.5296293	-1.6215943	1.23219983
C19_22d_WW1	-1.97733559	1.7854497	-1.3246754	1.15574442
C19_22d_WW2	0.01871974	1.0797062	-0.5137143	-0.02298542
	GABA	Trans.Caffeic.acid	Catechin	Citric.acid
C19_22d_WS1	-1.67750979	-1.9076588	0.8462298	-1.62534104
C19_22d_WS2	-0.15362857	0.5198699	1.0319866	-0.94395431
C19_22d_WS3	0.05908622	-0.4667952	-0.3489711	-0.34546992
C19_22d_WS4	-0.58473308	1.1867005	-0.6413015	-0.26479970
C19_22d_WW1	0.19730366	-2.6666662	0.9764719	-0.08069736
C19_22d_WW2	-0.21760505	-0.5650527	0.5055568	-0.76988063
	Erythronic.acid	Ethanolamine	Fructose	
C19_22d_WS1	-1.0760658	-0.9069005	1.0159463	
C19_22d_WS2	-0.9541824	0.3338993	1.1734210	
C19_22d_WS3	-1.5481519	-0.3212029	0.7347403	
C19_22d_WS4	-1.6713845	-1.2167341	0.7116506	

C19_22d_WW1	-1.4939087	-0.2157557	1.8321799	
C19_22d_WW2	-1.7527756	-0.9187787	1.9439168	
	Fructose.6.phosphate	Fumaric.acid	Galactinol	Galactose
C19_22d_WS1	-1.1899680	-1.2704112	2.1517037	0.9754190
C19_22d_WS2	-0.4381251	-0.3767642	1.4888870	1.2595376
C19_22d_WS3	-0.2691031	-1.9449380	2.3802436	0.8644708
C19_22d_WS4	0.1878136	0.3575794	1.0906958	1.0150634
C19_22d_WW1	1.0766640	-2.0877998	0.7032057	1.6772912
C19_22d_WW2	2.3354791	-1.2743013	0.9552469	1.5003171
	Gallic.acid	Gluconic.acid	Glucopyranose..H2O.	Glucose
C19_22d_WS1	-0.56205667	0.37818134	-0.41928333	0.8610600
C19_22d_WS2	-1.09115900	0.69707303	1.57621483	1.1899955
C19_22d_WS3	-2.53321141	0.26467059	0.54096494	0.7930875
C19_22d_WS4	0.39828768	-0.07473945	0.10960291	0.9509372
C19_22d_WW1	-0.39834799	-0.15924083	-1.17242614	1.5239844
C19_22d_WW2	0.03270293	-0.47006307	-0.09725887	1.3591781
	Glutamic.acid	Glyceric.acid	Glycine	Hydroquinone
C19_22d_WS1	1.8529397	-0.6396577	-0.209104316	-0.6134590
C19_22d_WS2	0.3557271	-0.2781580	-0.002480725	0.5769813
C19_22d_WS3	-1.2262505	0.1349363	1.118882580	0.1940983
C19_22d_WS4	-0.7236932	-0.1712099	1.131949816	-0.3672885
C19_22d_WW1	0.9720205	-0.6781024	0.182613833	-0.7239927
C19_22d_WW2	-0.1666686	-0.9911078	1.662667532	0.1226357
	Myo.Inostol	Isoleucine	Leucine	Lyxonic.acid
C19_22d_WS1	0.3929796	0.8083210	0.8838270	0.54364849
C19_22d_WS2	0.5757377	0.4685375	0.5263301	1.06677525
C19_22d_WS3	-0.1162823	0.3108856	0.4481515	-0.04072261
C19_22d_WS4	-0.1567574	0.9560411	1.0345703	0.23010740
C19_22d_WW1	-0.0961452	1.3573957	1.4206893	0.18985694
C19_22d_WW2	-0.5049368	-0.9065531	-0.8335214	0.45994567
	Maleic.acid	Malic.acid	Malonic.acid	Mannose.6.phosphate
C19_22d_WS1	-0.04124987	0.1562938	-2.45252896	-1.63479120
C19_22d_WS2	-1.53286791	-0.3889474	0.69455598	-0.90732420
C19_22d_WS3	-2.19989323	-0.2911455	-0.08540974	-0.14468505
C19_22d_WS4	-0.15837558	-1.1958191	0.70085590	-0.01888854
C19_22d_WW1	-1.98013894	0.4364033	-2.32135889	0.40015919
C19_22d_WW2	-3.00741667	-0.5151019	-0.23606303	1.26830623
	Melibiose	Phenylalanine	Phosphoric.acid	Proline
C19_22d_WS1	-0.9152522	0.29408257	-0.3869678	1.5641890
C19_22d_WS2	-0.8013869	0.09057998	0.4244065	1.0713501
C19_22d_WS3	-1.3171307	0.70483165	-0.1471089	0.9917029
C19_22d_WS4	-1.9736705	0.96852929	0.5707605	0.8396704
C19_22d_WW1	-0.8013869	0.72685092	2.0542015	1.3925337
C19_22d_WW2	-1.1218548	-0.09134266	2.2956621	-2.3905820
	Putrescine	Pyroglutamic.acid	Quinic.acid	
C19_22d_WS1	-0.09701713	-0.6880501	1.0134041	
C19_22d_WS2	0.83102826	0.9161958	1.1903568	
C19_22d_WS3	0.11070822	1.0095496	1.0504003	
C19_22d_WS4	0.10774946	1.3341814	0.8012049	
C19_22d_WW1	1.50861243	-2.0818139	0.8599133	
C19_22d_WW2	-0.88802174	-0.8801482	0.9170522	
	X3.caffeoylquinic.acid	Raffinose	Rhamnose	Ribonic.acid
C19_22d_WS1		1.0129338	0.90171619	-0.1916521
				0.57322107

C19_22d_WS2	1.0843258	0.73515622	-0.7853709	1.07373454
C19_22d_WS3	0.9960499	0.95229701	-0.4685484	-0.05742907
C19_22d_WS4	1.1021410	0.08731885	-0.7766106	0.21379339
C19_22d_WW1	1.0252094	0.17392032	-0.4018509	0.22270103
C19_22d_WW2	0.9860591	0.15887096	-0.5857015	0.45664047
	Ribose	Serine	Shikimic.acid	Succinic.acid
C19_22d_WS1	-0.725378295	-0.9819851	1.714582	1.19133927
C19_22d_WS2	0.377731531	-0.4214763	1.783446	1.08770110
C19_22d_WS3	-0.994502494	1.0487770	1.775418	0.67194197
C19_22d_WS4	-0.861358940	0.9681294	1.522672	-0.09238325
C19_22d_WW1	-1.007359346	-0.3426742	1.721841	0.50406151
C19_22d_WW2	0.008470629	-1.0378498	1.706724	0.24577356
	Sucrose	Tartaric.acid	Threitol	Threonic.acid
C19_22d_WS1	-0.21803504	-1.2815236	-0.269263695	-0.6126150
C19_22d_WS2	0.24534671	-0.5084729	-0.990507684	-0.9834298
C19_22d_WS3	-1.16816077	-0.5443348	1.138958898	-1.7190770
C19_22d_WS4	0.06888615	-0.2475653	0.003462422	-2.6022790
C19_22d_WW1	-0.01526017	-0.6051255	-0.883862796	-1.9874641
C19_22d_WW2	0.17306871	-0.3731469	0.857245383	-1.2881125
	Threonolactone	Threonine	Uracil	Valine
C19_22d_WS1	-0.2910448	0.9810916	-1.14948726	1.69636873
C19_22d_WS2	-0.5700936	1.2110984	-0.17321780	1.14513724
C19_22d_WS3	-1.1940542	0.6913117	-0.57353729	1.67605390
C19_22d_WS4	-2.0323783	0.2333270	0.96204325	1.19798021
C19_22d_WW1	-0.5668796	0.2288310	-0.01564416	1.81162694
C19_22d_WW2	-0.2221427	0.1267976	-0.37825144	-0.05925025
	Xylose			
C19_22d_WS1	-0.14129898			
C19_22d_WS2	0.67576048			
C19_22d_WS3	-0.48734567			
C19_22d_WS4	-0.02187825			
C19_22d_WW1	0.98161745			
C19_22d_WW2	0.19369245			

----- sm1819: Standardized metabolites 18/19 -----

name	description	class	nrow	ncol
13 sm1819	Standardized metabolites 18/19 matrix		190	55
	Alanine	Arabinose	Ascorbic.acid	Aspartic.acid
C18_11d_WS1	-1.3216743	0.81081282	0.9519808	0.2446827
C18_11d_WS2	-1.5850312	0.93631495	1.3802355	-0.3672711
C18_11d_WS3	1.4778432	0.03230802	1.2808356	-0.1083831
C18_11d_WS4	1.5489853	0.68090622	1.9164602	0.1363838
C18_11d_WW1	-1.2812775	2.00130194	0.7199549	-0.5527383
C18_11d_WW2	-0.3297096	1.68759346	0.9852330	-0.3346628
	GABA	Trans.Caffeic.acid	Catechin	Citric.acid
C18_11d_WS1	-0.2519906	0.80770699	1.3115059	-0.3119820
C18_11d_WS2	-0.7511261	0.98244100	1.4698917	-0.4326158
C18_11d_WS3	-0.5202380	-0.01718941	1.1020518	-0.6696953
C18_11d_WS4	-0.3609178	0.16155056	1.0420246	0.1861175
C18_11d_WW1	-0.2607749	0.55990013	1.0593488	-0.2368019

C18_11d_WW2	-0.4578432	0.01176799	0.8563507	-0.5763877
	Erythronic.acid	Ethanolamine	Fructose	
C18_11d_WS1	-0.8746026	-0.53998192	0.99215264	
C18_11d_WS2	-0.3362622	-0.95018249	0.62627555	
C18_11d_WS3	-0.7153057	-0.04066661	-0.06395835	
C18_11d_WS4	-1.0687552	-0.86472198	0.16050289	
C18_11d_WW1	-0.3941699	-0.95989684	0.65208504	
C18_11d_WW2	-1.6495500	-1.10084850	0.47982506	
	Fructose.6.phosphate	Fumaric.acid	Galactinol	Galactose
C18_11d_WS1	0.202208256	-2.3255100	0.393038165	1.5647978
C18_11d_WS2	0.107588328	-0.9958129	0.016625527	1.5087783
C18_11d_WS3	0.967623882	0.7221923	-0.001030863	0.2979007
C18_11d_WS4	1.493772779	0.8768515	0.117421853	1.0031588
C18_11d_WW1	0.007399398	-2.0084384	0.268295050	0.9121470
C18_11d_WW2	1.696717518	-0.9271973	0.046216383	0.7056666
	Gallic.acid	Gluconic.acid	Glucopyranose..H2O.	Glucose
C18_11d_WS1	0.6552556	0.7966034	-0.26091191	1.3799156
C18_11d_WS2	0.5208686	0.9581338	0.01215372	1.2776290
C18_11d_WS3	0.4513421	0.4200960	0.48101331	0.6386647
C18_11d_WS4	0.3505476	0.6570243	-0.03185639	1.2537365
C18_11d_WW1	1.0492647	0.4592050	0.30518953	1.0286806
C18_11d_WW2	0.3899686	0.2104459	0.03394629	0.9109430
	Glutamic.acid	Glyceric.acid	Glycine	Hydroquinone
C18_11d_WS1	0.9684981	0.9242754	0.9424986	1.6367642
C18_11d_WS2	0.2974240	0.5873407	0.8985881	2.1123854
C18_11d_WS3	-0.7178217	0.5333133	1.3883966	0.7337700
C18_11d_WS4	-0.2251164	0.9358776	0.9424986	1.8021784
C18_11d_WW1	0.3197855	0.5681464	1.6497358	0.7709097
C18_11d_WW2	0.1295187	0.4684835	1.2270649	1.6367642
	Myo.Inostol	Isoleucine	Leucine	Lyxonic.acid
C18_11d_WS1	1.0265053	-0.9370969	-1.28454542	0.05849233
C18_11d_WS2	0.9470041	-0.9635587	-1.06278206	0.71877226
C18_11d_WS3	1.2715262	-0.5486835	-0.84548411	0.90820545
C18_11d_WS4	1.8757337	0.5097963	0.09978663	0.02262333
C18_11d_WW1	0.7788075	0.4708175	0.07201951	0.78787203
C18_11d_WW2	1.0130190	-0.1338082	-0.39553971	0.35256857
	Maleic.acid	Malic.acid	Malonic.acid	Mannose.6.phosphate
C18_11d_WS1	-0.07418252	0.4657714	0.1067572	0.18623208
C18_11d_WS2	0.86317474	0.8806769	-0.4627324	0.44188513
C18_11d_WS3	-0.03258706	0.2597052	0.4327085	0.91439372
C18_11d_WS4	-0.09941469	0.4838808	1.2402534	1.36672608
C18_11d_WW1	-0.73913172	-0.3351734	-0.1009585	0.06394756
C18_11d_WW2	0.21591145	-0.4793880	0.1863597	1.59047486
	Melibiose	Phenylalanine	Phosphoric.acid	Proline
C18_11d_WS1	0.10278882	-0.68751145	0.605441967	-0.26080192
C18_11d_WS2	-0.08996645	-0.49153302	-0.009926619	-0.08957458
C18_11d_WS3	-2.04587081	-0.60817138	0.184462362	-0.88207988
C18_11d_WS4	-1.52680105	0.05761733	0.634626673	-0.58721109
C18_11d_WW1	-0.33596066	-0.59869016	1.210003081	-0.88207988
C18_11d_WW2	-0.48376515	-0.06584986	1.207979760	-0.58721109
	Putrescine	Pyroglutamic.acid	Quinic.acid	
C18_11d_WS1	-0.7704912	-0.06071954	1.2968180	
C18_11d_WS2	-1.0928669	-0.97572564	1.0387640	

C18_11d_WS3	0.5472319	1.04425188	0.9915949	
C18_11d_WS4	0.6781973	1.83322698	0.8400808	
C18_11d_WW1	-0.3670343	-0.55353059	1.5456855	
C18_11d_WW2	-0.6268200	-0.27982021	1.2181097	
	X3.caffeoylquinic.acid	Raffinose	Rhamnose	
C18_11d_WS1	1.195696	0.24980597	-0.3189756	
C18_11d_WS2	1.170472	0.66225216	-0.8333650	
C18_11d_WS3	0.966176	0.05815485	-0.6461897	
C18_11d_WS4	1.010474	0.30231675	-0.2136950	
C18_11d_WW1	1.132066	-0.06858858	-0.5741800	
C18_11d_WW2	1.045208	0.11011083	0.1307519	
	Ribonic.acid	Ribose	Serine	Shikimic.acid
C18_11d_WS1	0.4479829	0.5847252	1.5289606	1.883613
C18_11d_WS2	-0.2222956	0.6177648	0.6076439	1.186979
C18_11d_WS3	1.9414142	0.9366934	1.3272393	1.313410
C18_11d_WS4	0.8683185	0.6831970	1.7397064	1.298525
C18_11d_WW1	1.4170629	0.9981112	0.1034093	1.870237
C18_11d_WW2	1.4170629	0.9981112	0.4205826	1.560614
	Succinic.acid	Sucrose	Tartaric.acid	Threitol
C18_11d_WS1	1.12378351	0.9727693	0.88130394	-0.1755182
C18_11d_WS2	1.00592868	1.1753149	-0.05506813	-1.0908375
C18_11d_WS3	-0.06702313	1.6733871	-0.39333848	0.0389203
C18_11d_WS4	0.39986495	1.9385446	0.65385286	0.5980542
C18_11d_WW1	1.14293013	1.1243437	0.48511174	0.5123143
C18_11d_WW2	0.70885137	1.3346676	0.35565954	0.4238521
	Threonic.acid	Threonolactone	Threonine	Uracil
C18_11d_WS1	0.7865698	0.79978675	0.4644569	-1.5690944
C18_11d_WS2	0.8761986	0.52235743	-0.2077896	0.1557937
C18_11d_WS3	0.7974524	-0.00115696	0.1564808	-0.3401481
C18_11d_WS4	0.5707593	0.62051830	0.7574499	0.3822761
C18_11d_WW1	1.5021302	0.95071900	-0.2486532	-0.9071655
C18_11d_WW2	1.5737913	2.07913320	0.1092518	0.3822761
	Valine	Xylose		
C18_11d_WS1	-0.33710795	1.5782722		
C18_11d_WS2	-0.38696882	0.5869536		
C18_11d_WS3	0.15273094	0.9144890		
C18_11d_WS4	1.20476153	1.3174949		
C18_11d_WW1	0.94007550	1.1434431		
C18_11d_WW2	-0.04545139	2.1347616		

----- mldata: Metabolites featuredata 18/19 -----

name	description	class	nrow	ncol
14 mldata	Metabolites featuredata 18/19	data.frame	55	3

Metabolite	Bin
Alanine	5.1001
Arabinose	10.6.1001
Ascorbic.acid	17.5.1001
Aspartic.acid	13.1.1.2.1001
GABA	22.1002
Trans.Caffeic.acid	16.2.1.1006

Alanine
 Arabinose cell wall
 Ascorbic.acid hormone metabol
 Aspartic.acid amino acid metabolism.synthesis.central amino acid metaboli
 GABA pol
 Trans.Caffeic.acid secondary metabolism.phenylpropanoids.lignin

----- intgenes: Names of genes in bins -----

name	description	class	nrow	ncol
15 intgenes	Names of genes in bins	list	8	<NA>

\$`2`

[1] "Vitvi02g01845" "Vitvi01g00025" "Vitvi01g00052" "Vitvi01g00053"
 [5] "Vitvi01g00064" "Vitvi01g00681"

\$`3`

[1] "Vitvi02g01701" "Vitvi10g02295" "Vitvi00g01259" "Vitvi01g00140"
 [5] "Vitvi01g00509" "Vitvi01g00714"

\$`8`

[1] "Vitvi07g03070" "Vitvi01g00274" "Vitvi01g00698" "Vitvi01g00735"
 [5] "Vitvi01g00861" "Vitvi01g01724"

\$`10`

[1] "Vitvi02g01716" "Vitvi07g02990" "Vitvi10g02284" "Vitvi10g02279"
 [5] "Vitvi07g02943" "Vitvi00g02116"

\$`13`

[1] "Vitvi07g03049" "Vitvi00g00869" "Vitvi02g01749" "Vitvi01g00035"
 [5] "Vitvi01g00113" "Vitvi01g00502"

\$`16`

[1] "Vitvi02g01683" "Vitvi02g01722" "Vitvi10g02185" "Vitvi07g03081"
 [5] "Vitvi10g02269" "Vitvi07g03083"

\$`27.3`

[1] "Vitvi02g01753" "Vitvi02g01762" "Vitvi02g01795" "Vitvi02g01721"
 [5] "Vitvi02g01717" "Vitvi02g01790"

\$`35`

[1] "Vitvi07g02832" "Vitvi07g02830" "Vitvi07g02812" "Vitvi07g02811"
 [5] "Vitvi09g02033" "Vitvi09g02034"

----- intmtbs: Names of metabolites in bins -----

name	description	class	nrow	ncol
16 intmtbs	Names of metabolites in bins	list	8	<NA>

```

$`2`
[1] "Fructose" "Glucose" "Sucrose"

$`3`
[1] "Erythronic.acid" "Galactinol" "Galactose"
[4] "Myo.Inostol" "Raffinose" "Threitol"

$`8`
[1] "Fumaric.acid" "Succinic.acid"

$`10`
[1] "Arabinose" "Melibiose" "Rhamnose" "Ribonic.acid"
[5] "Ribose"

$`13`
[1] "Aspartic.acid" "Isoleucine" "Leucine" "Phenylalanine"
[5] "Proline" "Shikimic.acid"

$`16`
[1] "Trans.Caffeic.acid" "Catechin"
[3] "Gallic.acid" "Hydroquinone"
[5] "Quinic.acid" "X3.caffeoylquinic.acid"

$`27.3`
character(0)

$`35`
[1] "Gluconic.acid" "Glucopyranose..H2O." "Lyxonic.acid"
[4] "Maleic.acid" "Malonic.acid" "Phosphoric.acid"

```

----- stat4: Statistics from transcript analysis (50*.Rnw) -----

```

      name                description      class
17 stat4 Statistics from transcript analysis (50*.Rnw) data.frame
      nrow ncol
17 15242  28

```

	X	A	Coef.mc	Coef.c		
Vitvi03g00325	Vitvi03g00325	2.29529737	3.1923214	5.400429		
Vitvi17g00601	Vitvi17g00601	-0.09054575	-1.1866150	4.512791		
Vitvi05g00011	Vitvi05g00011	3.00782492	2.7136299	4.101024		
Vitvi13g02005	Vitvi13g02005	2.33061623	2.9484215	4.078080		
Vitvi02g00695	Vitvi02g00695	-0.45420732	-0.8399601	4.047520		
Vitvi14g01381	Vitvi14g01381	-0.72321946	-1.1530667	3.935431		
	Coef.mf.mc	Coef.f.c	Koef.f	t.mc	t.c	
Vitvi03g00325	-1.7940481	-0.29162541	5.108803	4.801209	2.919406	
Vitvi17g00601	2.1921385	-0.33971975	4.173071	-2.341540	3.200805	
Vitvi05g00011	0.5883901	0.03610049	4.137125	8.336348	4.528348	
Vitvi13g02005	-1.2356105	1.22197743	5.300058	3.618330	1.798857	
Vitvi02g00695	0.7715056	-1.08372917	2.963791	-1.857328	3.216927	
Vitvi14g01381	0.8596946	0.58827898	4.523710	-1.546481	1.897163	

	t.mf.mc	t.f.c	P.value.mc	P.value.c
Vitvi03g00325	-1.9079328	-0.10592036	9.103982e-05	0.0080809276
Vitvi17g00601	3.0587557	-0.16189100	2.892715e-02	0.0042173372
Vitvi05g00011	1.2781316	0.02678234	3.604890e-08	0.0001755321
Vitvi13g02005	-1.0722232	0.36215284	1.571115e-03	0.0861338096
Vitvi02g00695	1.2062964	-0.57870975	7.705364e-02	0.0040613495
Vitvi14g01381	0.8153037	0.19053890	1.366212e-01	0.0713562714
	P.value.mf.mc	P.value.f.c	P.value.adj.mc	P.value.adj.c
Vitvi03g00325	0.069880852	0.9166302	1.006038e-04	0.28511458
Vitvi17g00601	0.005867102	0.8729075	3.029669e-02	0.23762225
Vitvi05g00011	0.214866433	0.9788810	4.329163e-08	0.07644173
Vitvi13g02005	0.295539873	0.7207826	1.694639e-03	0.58897413
Vitvi02g00695	0.240844889	0.5688147	7.988693e-02	0.23288283
Vitvi14g01381	0.423862612	0.8506793	1.407775e-01	0.56306208
	P.value.adj.mf.mc	P.value.adj.f.c	F	
Vitvi03g00325	0.29606841	0.9999488	9.550446	
Vitvi17g00601	0.05151288	0.9999488	37.060841	
Vitvi05g00011	0.56223076	0.9999488	84.152716	
Vitvi13g02005	0.66176271	0.9999488	6.968467	
Vitvi02g00695	0.59836313	0.9999488	35.455608	
Vitvi14g01381	0.76054331	0.9999488	19.816526	
	F.p.value	Type	interestingC	interestingFC
Vitvi03g00325	1.356555e-04	Type5	*	
Vitvi17g00601	2.421190e-09	Type5	*	
Vitvi05g00011	8.630059e-13	Type1	*	
Vitvi13g02005	9.348733e-04	Type1.4		
Vitvi02g00695	3.642503e-09		*	
Vitvi14g01381	5.922355e-07	Type1		
	geneID	BINCODE		
Vitvi03g00325	Vitvi03g00325	27.3.25		
Vitvi17g00601	Vitvi17g00601	17.6.3		
Vitvi05g00011	Vitvi05g00011	35.2		
Vitvi13g02005	Vitvi13g02005	29.5.1		
Vitvi02g00695	Vitvi02g00695	20.1		
Vitvi14g01381	Vitvi14g01381	35.2		
Vitvi03g00325	RNA.regulation of transcription.MYB domain transcription factor			
Vitvi17g00601	hormone metabolism.gibberelin.induced-regulated-responsive-act			
Vitvi05g00011	not assigned.u			
Vitvi13g02005	protein.degradation.subt			
Vitvi02g00695	stress.			
Vitvi14g01381	not assigned.u			
Vitvi03g00325	RAD-like			
Vitvi17g00601	Gibberellin-regulated family prote			
Vitvi05g00011	Pollen Ole e 1 allergen and extensin family pro			
Vitvi13g02005	Subtilisin-like serine endopeptidase family prote			
Vitvi02g00695	Polyketide cyclase/dehydrase and lipid transport superfamily pro			
Vitvi14g01381				

----- statig: Statistics for interesting genes -----

```

name                description                class nrow ncol
18 statig Statistics for interesting genes data.frame 8424 28

```

		X	A	Coef.mc	Coef.c	Coef.mf.mc
NA	<NA>	NA	NA	NA	NA	NA
Vitvi01g00025	Vitvi01g00025	3.057806	3.114662	-1.26413832	-0.1137121	
Vitvi01g00052	Vitvi01g00052	3.349527	3.259138	-0.78711296	0.1807789	
Vitvi01g00053	Vitvi01g00053	4.631552	4.893533	0.19949344	-0.5239621	
Vitvi01g00064	Vitvi01g00064	1.833567	2.504210	-0.03291293	-1.3412870	
Vitvi01g00681	Vitvi01g00681	7.031147	7.124845	-0.23932641	-0.1873968	
		Coef.f.c	Koef.f	t.mc	t.c	t.mf.mc
NA	NA	NA	NA	NA	NA	NA
Vitvi01g00025		0.8379419	-0.4261964	21.759123	-3.17429440	-0.5617227
Vitvi01g00052		1.0025864	0.2154735	14.324978	-1.24351347	0.5618546
Vitvi01g00053		-0.8385976	-0.6391041	16.427027	0.24070623	-1.2437164
Vitvi01g00064		2.0300614	1.9971485	6.239789	-0.02947731	-2.3632291
Vitvi01g00681		0.1691963	-0.0701301	52.405974	-0.63272966	-0.9746584
		t.f.c	P.value.mc	P.value.c	P.value.mf.mc	
NA	NA	NA	NA	NA	NA	NA
Vitvi01g00025		1.4136908	4.211606e-16	0.004486561	0.58012959	
Vitvi01g00052		1.0641997	1.868977e-12	0.227103309	0.58004129	
Vitvi01g00053		-0.6798298	1.255313e-13	0.812069388	0.22703005	
Vitvi01g00064		1.2215701	3.137274e-06	0.976756560	0.02762602	
Vitvi01g00681		0.3005428	3.998197e-24	0.533600377	0.34059797	
		P.value.f.c	P.value.adj.mc	P.value.adj.c		
NA	NA	NA	NA	NA	NA	NA
Vitvi01g00025		0.1718074	8.299069e-16	0.2416402		
Vitvi01g00052		0.2990763	2.640125e-12	0.7466585		
Vitvi01g00053		0.5038863	1.914305e-13	0.9594257		
Vitvi01g00064		0.2351312	3.580556e-06	0.9965809		
Vitvi01g00681		0.7666578	1.819120e-22	0.8804002		
		P.value.adj.mf.mc	P.value.adj.f.c	F		
NA	NA	NA	NA	NA	NA	NA
Vitvi01g00025		0.8515913	0.9999488	1305.9897		
Vitvi01g00052		0.8515913	0.9999488	553.7057		
Vitvi01g00053		0.5797790	0.9999488	599.6382		
Vitvi01g00064		0.1599574	0.9999488	51.0410		
Vitvi01g00681		0.7000202	0.9999488	6549.1885		
		F.p.value	Type	interestingC	interestingFC	
NA	NA	<NA>	<NA>	<NA>	<NA>	<NA>
Vitvi01g00025		2.882747e-25		*		
Vitvi01g00052		2.674051e-21				
Vitvi01g00053		1.147679e-21				
Vitvi01g00064		1.179244e-10	Type3.F			
Vitvi01g00681		9.309219e-33				
		geneID	BINCODE			
NA	NA	<NA>	<NA>			
Vitvi01g00025	Vitvi01g00025	2.2.1.1				
Vitvi01g00052	Vitvi01g00052	2.2.2.6				
Vitvi01g00053	Vitvi01g00053	2.2.2.6				
Vitvi01g00064	Vitvi01g00064	2.2.2.2				
Vitvi01g00681	Vitvi01g00681	35.2				

NAME

```

NA <NA>
Vitvi01g00025 major CHO metabolism.degradation.sucrose.fructokinase
Vitvi01g00052 major CHO metabolism.degradation.starch.transporter
Vitvi01g00053 major CHO metabolism.degradation.starch.transporter
Vitvi01g00064 major CHO metabolism.degradation.starch.starch phosphorylase
Vitvi01g00681 not assigned.unknown

```

```

NA
Vitvi01g00025 fructokinase-like protein | Chr1:2
Vitvi01g00052 Glucose-6-phosphate/phosphate translocator-like protein | Chr5:1
Vitvi01g00053 Glucose-6-phosphate/phosphate translocator-like protein | Chr5:1
Vitvi01g00064 Glycosyl transferase%2C family 35 | Chr3:1
Vitvi01g00681 phosphoglucan%2C water dikinase | Chr4:12

```

Factor for days within the year

```
> pdata1819$day <- factor(as.character(pdata1819$day))
```

4 Preparation of data for mixOmics

Data for **mixOmics** analysis should follow the traditional mathematical orientation: samples in rows, variables in columns. The matrices must have appropriate dimensions. Due to different ranges and variability in years 2018 and 2019, we will analyze them separately.

4.0.1 Selection of samples, transcripts and metaboloms

Common samples in the given year (18).

```
> table(pdata1819$year)

18 19
48 32

> select_year <- pdata1819$year %in% .years
> sum(select_year)

[1] 48

> rownames(pdata1819)[select_year]

 [1] "C18_11d_WS1" "C18_11d_WS2" "C18_11d_WS3" "C18_11d_WS4"
 [5] "C18_11d_WW1" "C18_11d_WW2" "C18_11d_WW3" "C18_11d_WW4"
 [9] "C18_34d_WS1" "C18_34d_WS2" "C18_34d_WS3" "C18_34d_WS4"
[13] "C18_34d_WW1" "C18_34d_WW2" "C18_34d_WW3" "C18_34d_WW4"
[17] "C18_67d_WS1" "C18_67d_WS2" "C18_67d_WS3" "C18_67d_WS4"
[21] "C18_67d_WW1" "C18_67d_WW2" "C18_67d_WW3" "C18_67d_WW4"
[25] "F18_10d_WS1" "F18_10d_WS2" "F18_10d_WS3" "F18_10d_WS4"
[29] "F18_10d_WW1" "F18_10d_WW2" "F18_10d_WW3" "F18_10d_WW4"
[33] "F18_34d_WS1" "F18_34d_WS2" "F18_34d_WS3" "F18_34d_WS4"
[37] "F18_34d_WW1" "F18_34d_WW2" "F18_34d_WW3" "F18_34d_WW4"
[41] "F18_67d_WS1" "F18_67d_WS2" "F18_67d_WS3" "F18_67d_WS4"
[45] "F18_67d_WW1" "F18_67d_WW2" "F18_67d_WW3" "F18_67d_WW4"
```

Name of sample, gene and metabolite names to consider
Samples

```
> sampleNames <- rownames(pdata1819)[select_year]
> head(sampleNames)

[1] "C18_11d_WS1" "C18_11d_WS2" "C18_11d_WS3" "C18_11d_WS4"
[5] "C18_11d_WW1" "C18_11d_WW2"
```

4.0.2 Transcripts matrix: T

Exclude bin 35 for now

```
> intgenesAll <- intgenes
> intgenes <- intgenes[names(intgenes)!="35"]
> names(intgenes)
[1] "2"    "3"    "8"    "10"   "13"   "16"   "27.3"
```

Actually, start with small set of genes, keep just bin 1

```
> names(intgenesAll)
[1] "2"    "3"    "8"    "10"   "13"   "16"   "27.3" "35"
> keepbins <- names(intgenesAll)[.bins]
> intgenes <- intgenesAll[names(intgenesAll)%in% keepbins]
> names(intgenes)
[1] "2"    "3"    "8"    "10"   "13"   "16"   "27.3"
```

```
> geneNames <- unlist(intgenes)
> # any doublets
> which(table(geneNames)>1)
```

```
Vitvi14g02644 Vitvi18g01242
           1650           2037
```

```
> geneNames <- unique(geneNames)
> length(geneNames)
```

```
[1] 2178
```

```
> # keep genes that are measured
> geneNames <- geneNames[geneNames %in% rownames(t1819)]
> length(geneNames)
```

```
[1] 2049
```

```
> head(geneNames)
```

```
[1] "Vitvi01g00025" "Vitvi01g00052" "Vitvi01g00053" "Vitvi01g00064"
[5] "Vitvi01g00681" "Vitvi01g00932"
```

```
> T <- t1819[geneNames, sampleNames]
> head(T)
```

	C18_11d_WS1	C18_11d_WS2	C18_11d_WS3	C18_11d_WS4
Vitvi01g00025	2.525428	2.478706	2.651938	2.423295
Vitvi01g00052	3.372516	3.221593	3.540013	3.532502
Vitvi01g00053	4.733444	4.558951	5.010778	4.990309
Vitvi01g00064	3.204010	2.826364	3.435292	2.918474
Vitvi01g00681	7.006432	7.112737	7.282731	7.034244
Vitvi01g00932	3.902681	3.862343	4.236901	3.985243
	C18_11d_WW1	C18_11d_WW2	C18_11d_WW3	C18_11d_WW4
Vitvi01g00025	2.535710	2.212984	2.361454	2.516753
Vitvi01g00052	2.550469	3.315024	3.315533	3.221392
Vitvi01g00053	4.392540	4.709945	4.821214	4.843719
Vitvi01g00064	2.690307	2.970480	2.995004	3.004581
Vitvi01g00681	6.930850	7.044802	7.057018	7.024871
Vitvi01g00932	3.950284	3.816622	4.079860	3.807215
	C18_34d_WS1	C18_34d_WS2	C18_34d_WS3	C18_34d_WS4
Vitvi01g00025	2.578949	2.311085	2.583815	3.010003
Vitvi01g00052	2.253492	2.757454	2.656510	2.128346
Vitvi01g00053	4.594141	4.939657	4.431605	4.583447
Vitvi01g00064	2.511611	2.502197	2.142444	2.499113
Vitvi01g00681	6.891684	6.678274	6.859723	6.453177
Vitvi01g00932	3.745214	3.507557	4.022424	3.898226
	C18_34d_WW1	C18_34d_WW2	C18_34d_WW3	C18_34d_WW4
Vitvi01g00025	2.726730	2.745623	2.401136	2.614643
Vitvi01g00052	2.245390	1.684681	2.286576	2.679252
Vitvi01g00053	4.304855	4.545437	4.485788	5.121025
Vitvi01g00064	1.681696	2.702874	2.612473	2.295465
Vitvi01g00681	6.164293	6.536588	6.699936	6.371025
Vitvi01g00932	3.644268	3.104288	3.549279	3.684507
	C18_67d_WS1	C18_67d_WS2	C18_67d_WS3	C18_67d_WS4
Vitvi01g00025	3.954792	3.662574	3.702285	3.724124
Vitvi01g00052	2.688860	3.005134	3.291143	3.271692
Vitvi01g00053	3.690792	3.716075	3.626788	3.145972
Vitvi01g00064	1.753158	1.724258	2.113975	1.452874
Vitvi01g00681	6.959409	6.936884	6.836479	6.531912
Vitvi01g00932	4.848866	4.516614	4.411571	4.418600
	C18_67d_WW1	C18_67d_WW2	C18_67d_WW3	C18_67d_WW4
Vitvi01g00025	3.017593	3.1550122	3.098662	3.0632025
Vitvi01g00052	2.482471	2.3852736	2.784489	1.8757093
Vitvi01g00053	3.362112	3.5472244	3.706569	3.0389141
Vitvi01g00064	1.373478	0.9839821	1.378348	0.3759491
Vitvi01g00681	6.877357	6.8713393	6.813021	6.9990242
Vitvi01g00932	3.631923	3.8305964	3.678031	3.6617490
	F18_10d_WS1	F18_10d_WS2	F18_10d_WS3	F18_10d_WS4
Vitvi01g00025	3.190328	3.545382	3.811687	3.705025
Vitvi01g00052	4.033268	4.005980	3.479549	3.335980
Vitvi01g00053	3.076932	3.454593	3.492709	3.455495
Vitvi01g00064	4.645416	4.820738	4.626337	4.620433
Vitvi01g00681	7.047604	7.072526	7.152525	7.190802
Vitvi01g00932	4.871946	4.880086	4.918555	4.949219
	F18_10d_WW1	F18_10d_WW2	F18_10d_WW3	F18_10d_WW4
Vitvi01g00025	3.005951	2.755574	3.247298	2.969490
Vitvi01g00052	3.375932	3.188418	3.082239	3.253917
Vitvi01g00053	2.789974	2.822546	2.923977	2.733084

Vitvi01g00064	4.350841	4.319612	4.355473	4.429152
Vitvi01g00681	7.162528	6.908959	7.144751	6.976919
Vitvi01g00932	5.124985	4.787164	5.143878	5.115840
	F18_34d_WS1	F18_34d_WS2	F18_34d_WS3	F18_34d_WS4
Vitvi01g00025	2.16706262	2.1668879	2.1549196	2.638828
Vitvi01g00052	2.91541049	2.7158399	3.0016969	3.377254
Vitvi01g00053	4.34490150	4.3929206	5.2173197	5.025046
Vitvi01g00064	-0.09883144	0.8748018	0.3448193	1.496935
Vitvi01g00681	6.67898485	6.9279474	6.5777358	6.979445
Vitvi01g00932	3.10628299	3.2370563	3.5527121	3.244341
	F18_34d_WW1	F18_34d_WW2	F18_34d_WW3	F18_34d_WW4
Vitvi01g00025	1.9572744	1.85282095	2.254265	2.4787012
Vitvi01g00052	3.1387492	3.54087695	3.032267	2.9261602
Vitvi01g00053	4.4869625	4.99342458	4.780498	4.4272837
Vitvi01g00064	-0.2468172	-0.09471163	1.035023	0.9078483
Vitvi01g00681	6.3819679	6.48666832	6.767507	6.4394005
Vitvi01g00932	2.8977411	3.17830687	3.446615	3.6090979
	F18_67d_WS1	F18_67d_WS2	F18_67d_WS3	F18_67d_WS4
Vitvi01g00025	3.8714193	3.411367	3.4349493	2.83516476
Vitvi01g00052	2.4624461	2.732211	2.4752948	2.28683118
Vitvi01g00053	3.4543547	3.180826	3.0074054	3.94746773
Vitvi01g00064	0.1012586	-1.021232	-0.2978714	-0.05126174
Vitvi01g00681	6.9261259	6.723329	6.9402200	6.59279764
Vitvi01g00932	3.5053725	3.597451	2.9236761	2.42469999
	F18_67d_WW1	F18_67d_WW2	F18_67d_WW3	F18_67d_WW4
Vitvi01g00025	2.979030	3.007746	2.5105281	2.5763134
Vitvi01g00052	2.450210	2.047726	2.0475561	2.4108593
Vitvi01g00053	2.911310	2.969079	3.5205958	3.5941147
Vitvi01g00064	-2.261285	-1.478158	-0.4146633	-0.4869291
Vitvi01g00681	6.744527	6.736881	6.3439143	6.5238761
Vitvi01g00932	2.705269	2.491469	2.2928427	2.2889762

```
> T <- t(T)
```

```
> dim(T)
```

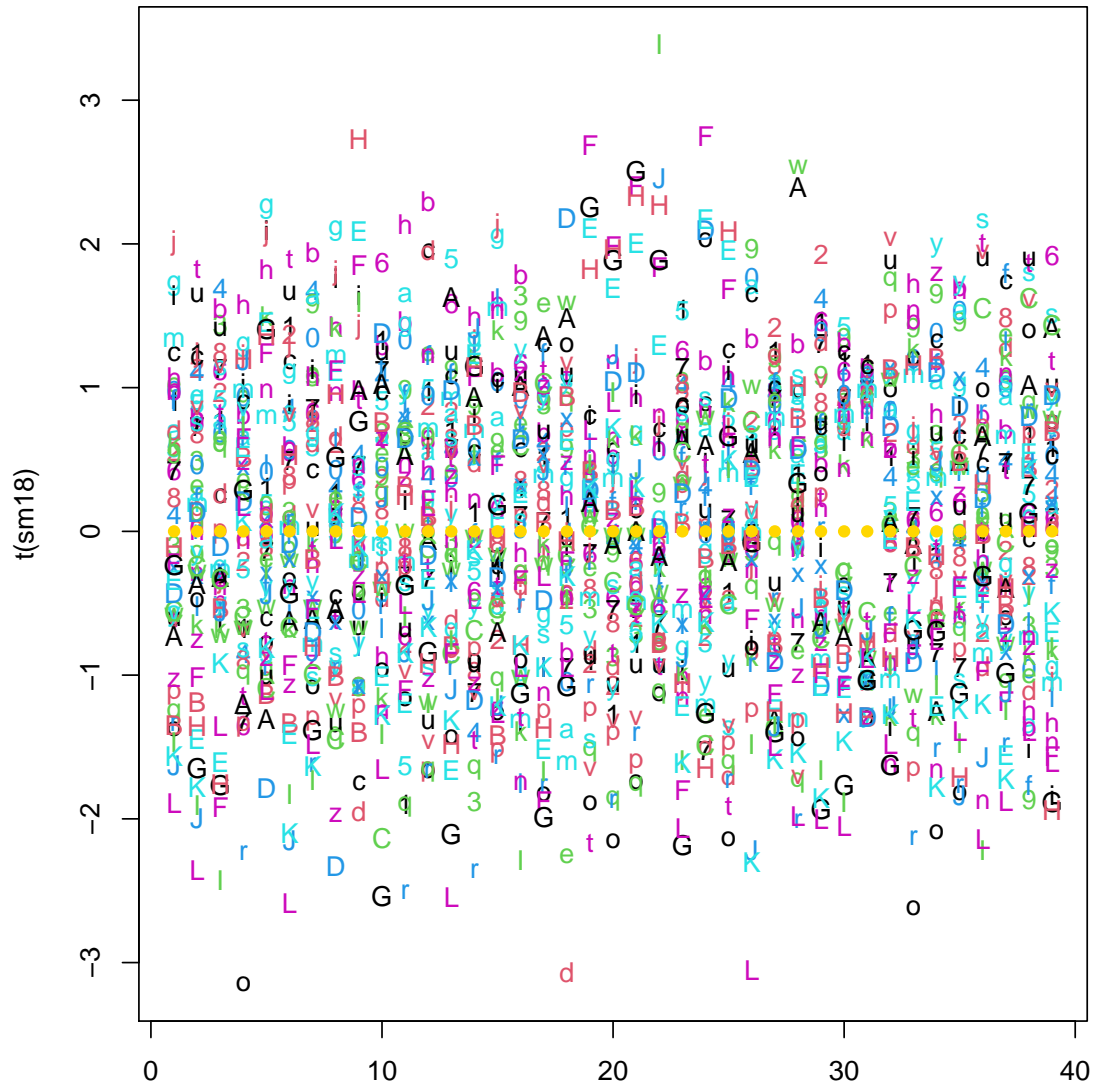
```
[1] 48 2049
```

4.0.3 Metabolites matrix: M

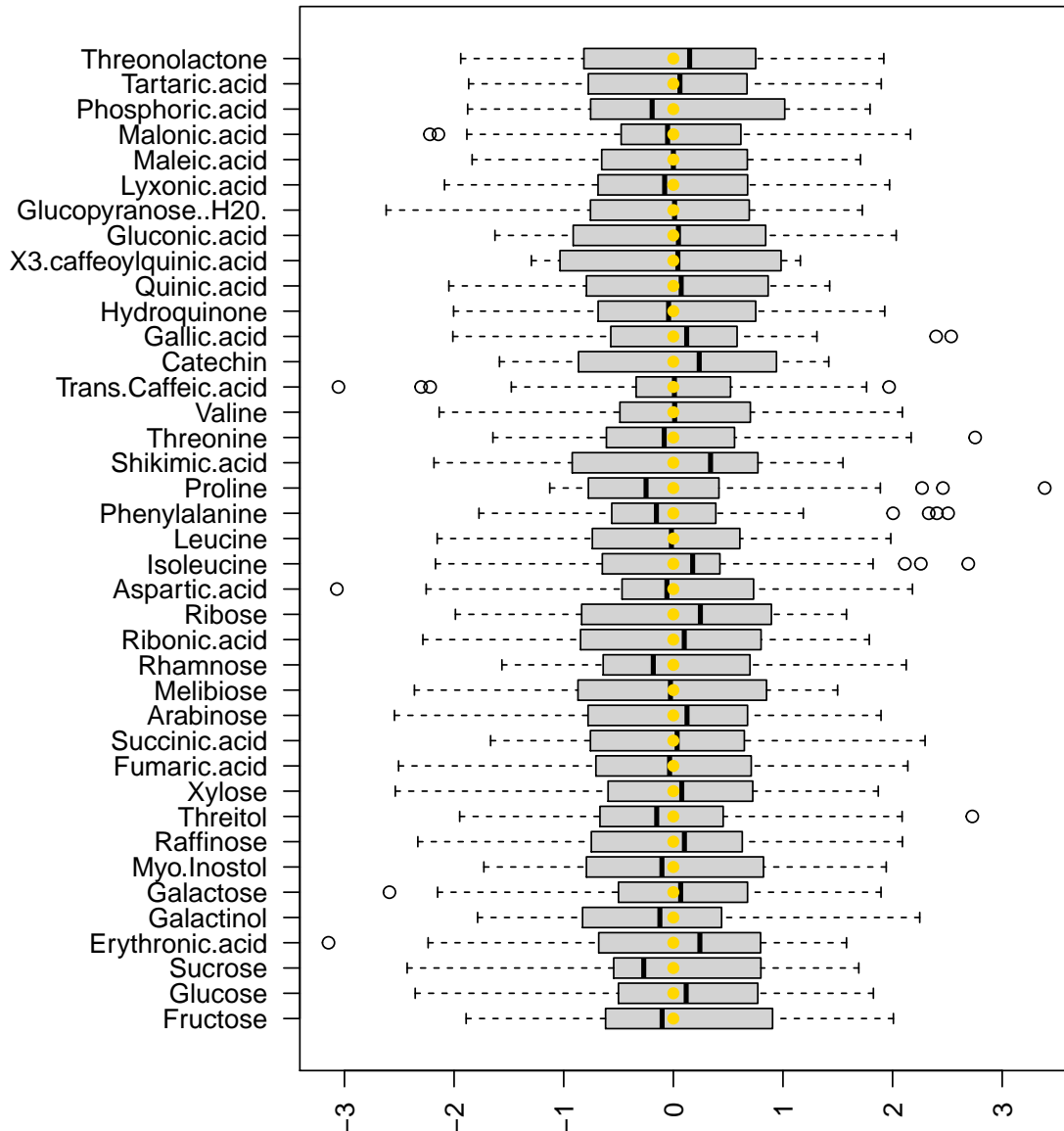
```
> metabNames <- unlist(intmtbs)
> # any doublets
> which(table(metabNames) > 1)
named integer(0)
> metabNames <- unique(metabNames)
> length(metabNames)
[1] 39
```

Standardization across log transformed variables variables, separately for each year.

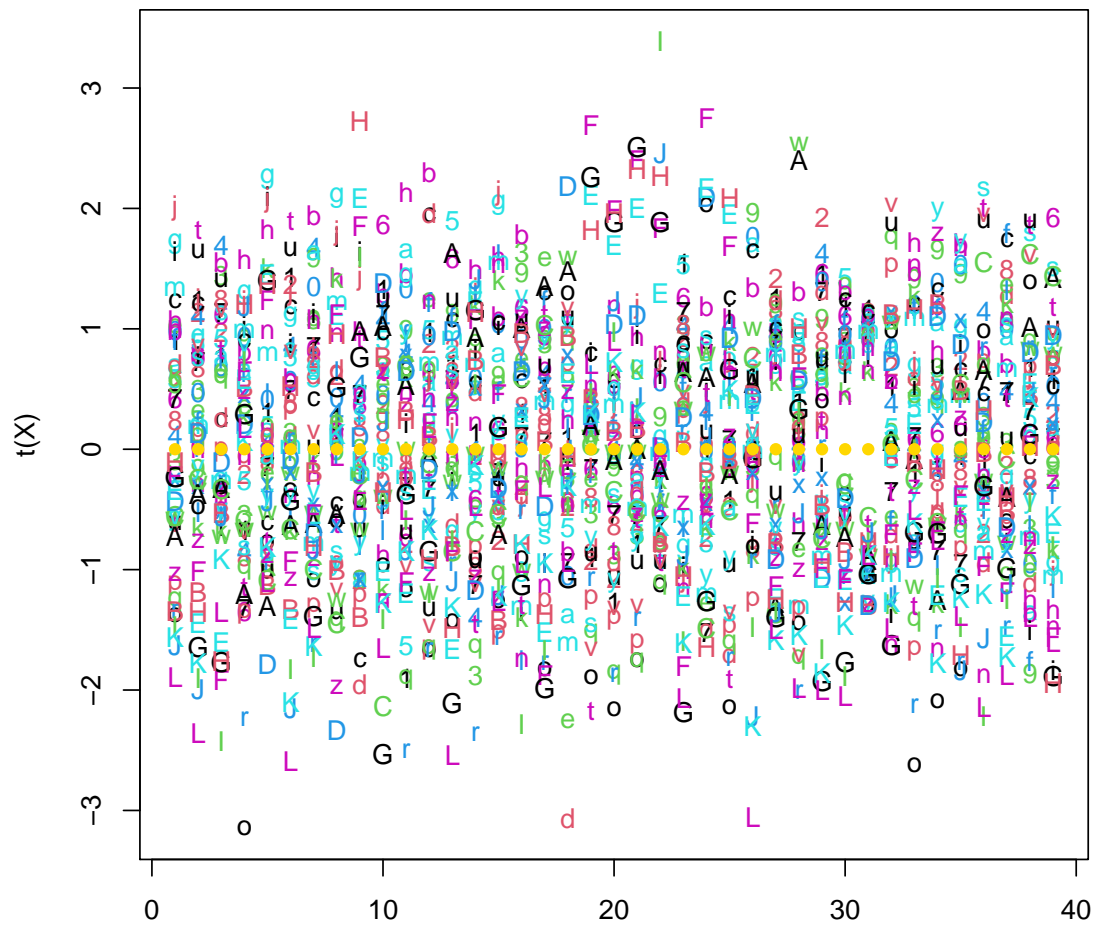
```
> if(18 %in% .years){
+ x <- log2(m18[,m_num_id])
+ dim(x)
+ x <- x[sampleNames,metabNames]
+ dim(x)
+ sx <- scale(x)
+ sm18 <- sx
+ matplot(t(sm18))
+ title("2018")
+ points(1:ncol(sx), apply(sx,2,mean, na.rm=TRUE), col="gold", pch=16)
+ head(apply(sx,2,mean, na.rm=TRUE))
+ head(apply(sx,2,sd, na.rm=TRUE))
+
+ par(mar=c(4,10,3,1))
+ boxplot(sm18, horizontal=TRUE,las=2, main="2018")
+ points( apply(sm18,2,mean, na.rm=TRUE),1:ncol(sm18), col="gold", pch=16)
+ }
```

2018



```
> if( 18 %in% .years) X <- sm18
> if( 19 %in% .years) X <- sm19
> if( sum(.years) > 20) X <- rbind(sm18,sm19)
> dim(X)
[1] 48 39
> matplot(t(X))
> title(paste(.years))
> sx <- X
> points(1:ncol(sx), apply(sx,2,mean, na.rm=TRUE), col="gold", pch=16)
```



```
> head(apply(sx, 2, mean, na.rm=TRUE))
```

Fructose	Glucose	Sucrose	Erythronic.acid
-1.730884e-16	-1.327786e-16	2.866992e-16	1.784814e-16
Galactinol	Galactose		
2.492829e-16	2.022398e-16		

```
> head(apply(sx, 2, sd, na.rm=TRUE))
```

Fructose	Glucose	Sucrose	Erythronic.acid
1	1	1	1
Galactinol	Galactose		
1	1		

```
> head(X)[, 1:4]
```

	Fructose	Glucose	Sucrose	Erythronic.acid
C18_11d_WS1	0.90600272	1.2404116	0.7748822	-0.6776825
C18_11d_WS2	0.55874466	1.1371844	0.9671302	-0.2090150
C18_11d_WS3	-0.09636408	0.4923445	1.4398799	-0.5390021
C18_11d_WS4	0.11667460	1.1130722	1.6915566	-0.8467077
C18_11d_WW1	0.58324074	0.8859467	0.9187504	-0.2594282
C18_11d_WW2	0.41974686	0.7671265	1.1183812	-1.3523351

```
> M <- X[sampleNames,metabNames]
> head(M)
```

	Fructose	Glucose	Sucrose	Erythronic.acid		
C18_11d_WS1	0.90600272	1.2404116	0.7748822	-0.6776825		
C18_11d_WS2	0.55874466	1.1371844	0.9671302	-0.2090150		
C18_11d_WS3	-0.09636408	0.4923445	1.4398799	-0.5390021		
C18_11d_WS4	0.11667460	1.1130722	1.6915566	-0.8467077		
C18_11d_WW1	0.58324074	0.8859467	0.9187504	-0.2594282		
C18_11d_WW2	0.41974686	0.7671265	1.1183812	-1.3523351		
	Galactinol	Galactose	Myo.Inostol	Raffinose		
C18_11d_WS1	0.30792405	1.4289376	0.8369468	0.264789702		
C18_11d_WS2	-0.03758577	1.3719441	0.7581463	0.610057076		
C18_11d_WS3	-0.05379260	0.1400126	1.0798082	0.104354522		
C18_11d_WS4	0.05493535	0.8575332	1.6786907	0.308747581		
C18_11d_WW1	0.19342213	0.7649389	0.5914321	-0.001745064		
C18_11d_WW2	-0.01042427	0.5548684	0.8235793	0.147847971		
	Threitol	Xylose	Fumaric.acid	Succinic.acid		
C18_11d_WS1	-0.204103803	1.3299342	-1.9010578	0.97483839		
C18_11d_WS2	-1.080614750	0.3694248	-0.7822587	0.87936541		
C18_11d_WS3	0.001242802	0.6867807	0.6632605	0.01017827		
C18_11d_WS4	0.536670150	1.0772616	0.7933898	0.38839947		
C18_11d_WW1	0.454565548	0.9086191	-1.6342757	0.99034887		
C18_11d_WW2	0.369854019	1.8691285	-0.7245259	0.63870616		
	Arabinose	Melibiose	Rhamnose	Ribonic.acid	Ribose	
C18_11d_WS1	0.70889487	0.13407439	-0.2695710	0.2999005	0.5227413	
C18_11d_WS2	0.83386822	-0.04685427	-0.7529906	-0.3001772	0.5561878	
C18_11d_WS3	-0.06632987	-1.88275301	-0.5770845	1.6369191	0.8790435	
C18_11d_WS4	0.57953561	-1.39553107	-0.1706291	0.6762127	0.6224256	
C18_11d_WW1	1.89436813	-0.27775536	-0.5094103	1.1674851	0.9412175	
C18_11d_WW2	1.58198139	-0.41649122	0.1530797	1.1674851	0.9412175	
	Aspartic.acid	Isoleucine	Leucine	Phenylalanine		
C18_11d_WS1	0.096814056	-0.8947734	-1.2723131	-0.78196306		
C18_11d_WS2	-0.475849907	-0.9190664	-1.0703230	-0.60007623		
C18_11d_WS3	-0.233583562	-0.5381957	-0.8724001	-0.70832785		
C18_11d_WS4	-0.004531663	0.4335275	-0.0114130	-0.09041192		
C18_11d_WW1	-0.649409392	0.3977435	-0.0367043	-0.69952836		
C18_11d_WW2	-0.445335215	-0.1573250	-0.4625743	-0.20500134		
	Proline	Shikimic.acid	Threonine	Valine		
C18_11d_WS1	-0.2490210	1.5474956	-0.003059961	-0.42502427		
C18_11d_WS2	-0.1040080	0.9256116	-0.732283919	-0.47239304		
C18_11d_WS3	-0.7751834	1.0384760	-0.337139139	0.04033179		
C18_11d_WS4	-0.5254580	1.0251886	0.314766273	1.03978044		
C18_11d_WW1	-0.7751834	1.5355550	-0.776610965	0.78832378		
C18_11d_WW2	-0.5254580	1.2591551	-0.388371084	-0.14794511		
	Trans.Caffeic.acid	Catechin	Gallic.acid	Hydroquinone		
C18_11d_WS1	0.6129458	1.2505486	0.40498086	1.4618442		
C18_11d_WS2	0.7731113	1.4175385	0.23907315	1.9282976		
C18_11d_WS3	-0.1431749	1.0297159	0.15323918	0.5762556		
C18_11d_WS4	0.0206626	0.9664278	0.02880320	1.6240700		
C18_11d_WW1	0.3857998	0.9846931	0.89140544	0.6126795		
C18_11d_WW2	-0.1166319	0.7706672	0.07747038	1.4618442		
	Quinic.acid	X3.caffeoylquinic.acid	Gluconic.acid			
C18_11d_WS1	1.1885212	1.1588285	0.52294016			

```

C18_11d_WS2    0.9424186                1.1347153    0.68732112
C18_11d_WS3    0.8974341                0.9394153    0.13978858
C18_11d_WS4    0.7529372                0.9817628    0.38089797
C18_11d_WW1    1.4258629                1.0980005    0.17958769
C18_11d_WW2    1.1134582                1.0149670   -0.07356123
      Glucopyranose..H2O. Lyxonic.acid Maleic.acid Malonic.acid
C18_11d_WS1    -0.19839007   -0.1502030  -0.04752106  -0.085073907
C18_11d_WS2     0.06406102    0.4915822   0.74164153  -0.701519897
C18_11d_WS3     0.51469519    0.6757093  -0.01250177   0.267753278
C18_11d_WS4     0.02176165   -0.1850673  -0.06876407   1.141883000
C18_11d_WW1     0.34570600    0.5587465  -0.60734287  -0.309916462
C18_11d_WW2     0.08500646    0.1356361   0.19670952   0.001092094
      Phosphoric.acid Tartaric.acid Threonolactone
C18_11d_WS1     0.46459431    0.6786103    0.5836665
C18_11d_WS2    -0.13222378   -0.3040638    0.2939347
C18_11d_WS3     0.05630528   -0.6590611   -0.2527947
C18_11d_WS4     0.49289924    0.4399122    0.3964485
C18_11d_WW1     1.05093073    0.2628271    0.7412918
C18_11d_WW2     1.04896840    0.1269737    1.9197450

```

```
> head(apply(sx, 2, mean, na.rm=TRUE))
```

```

      Fructose      Glucose      Sucrose Erythronic.acid
-1.730884e-16  -1.327786e-16   2.866992e-16   1.784814e-16
      Galactinol      Galactose
 2.492829e-16   2.022398e-16

```

```
> head(apply(sx, 2, sd, na.rm=TRUE))
```

```

      Fructose      Glucose      Sucrose Erythronic.acid
              1              1              1              1
      Galactinol      Galactose
              1              1

```

```
> dim(M)
```

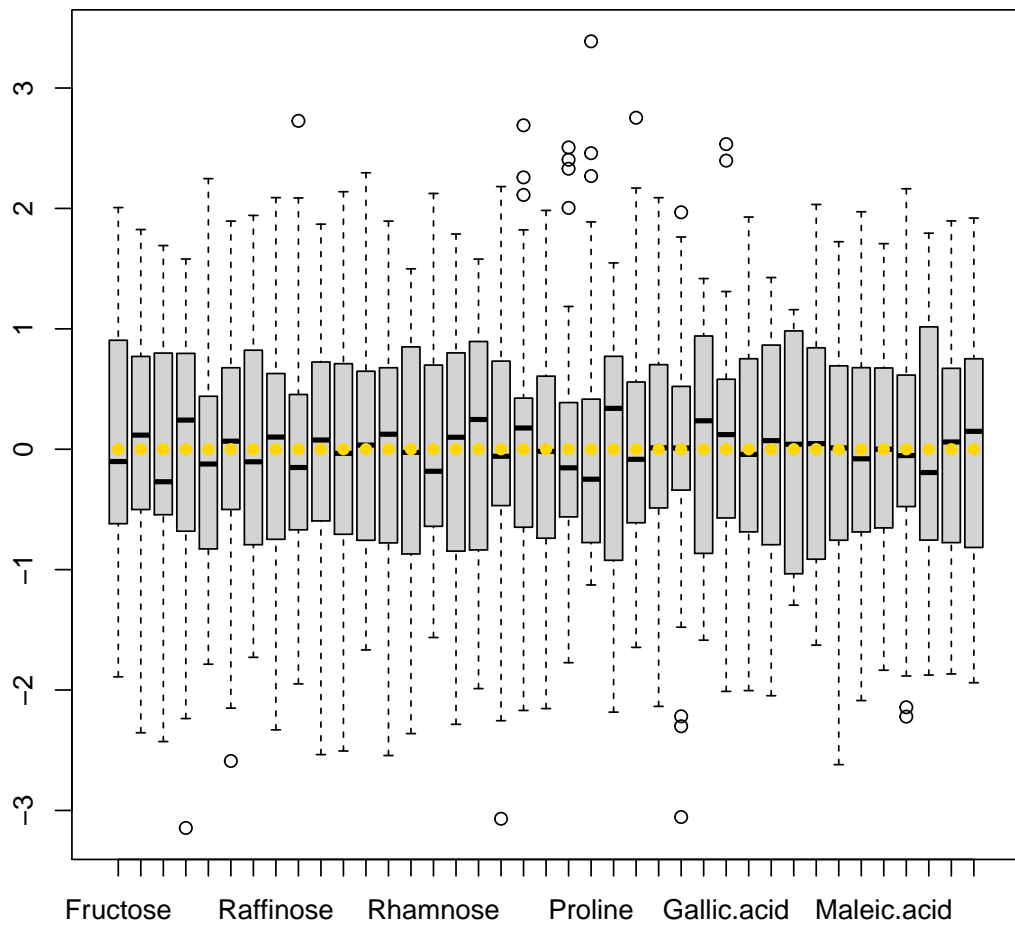
```
[1] 48 39
```

```
> range(M)
```

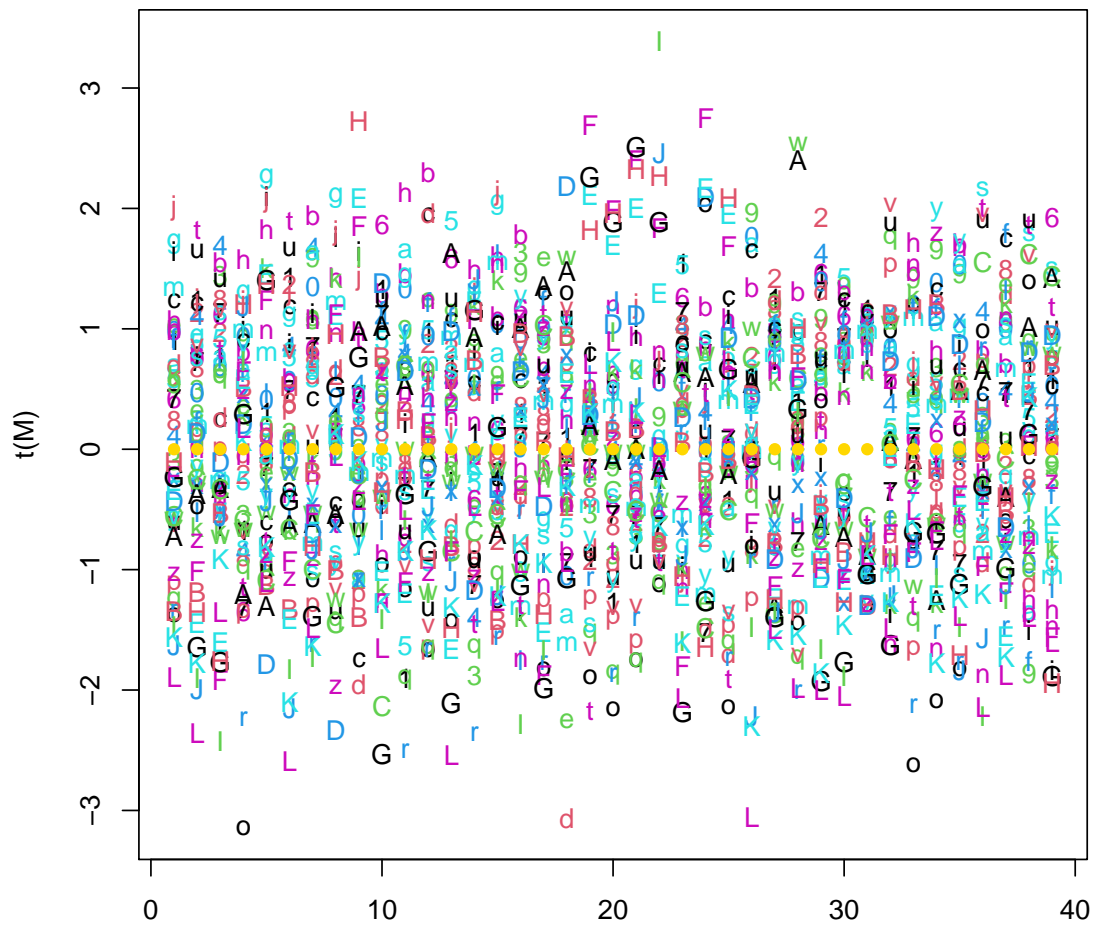
```
[1] -3.145570  3.387852
```

```
> boxplot(M)
```

```
> points(1:ncol(M), apply(M, 2, mean), pch=16, col="gold")
```



```
> matplot(t(M))
> points(1:ncol(M), apply(M, 2, mean), pch=16, col="gold")
```



Standardize transcripts

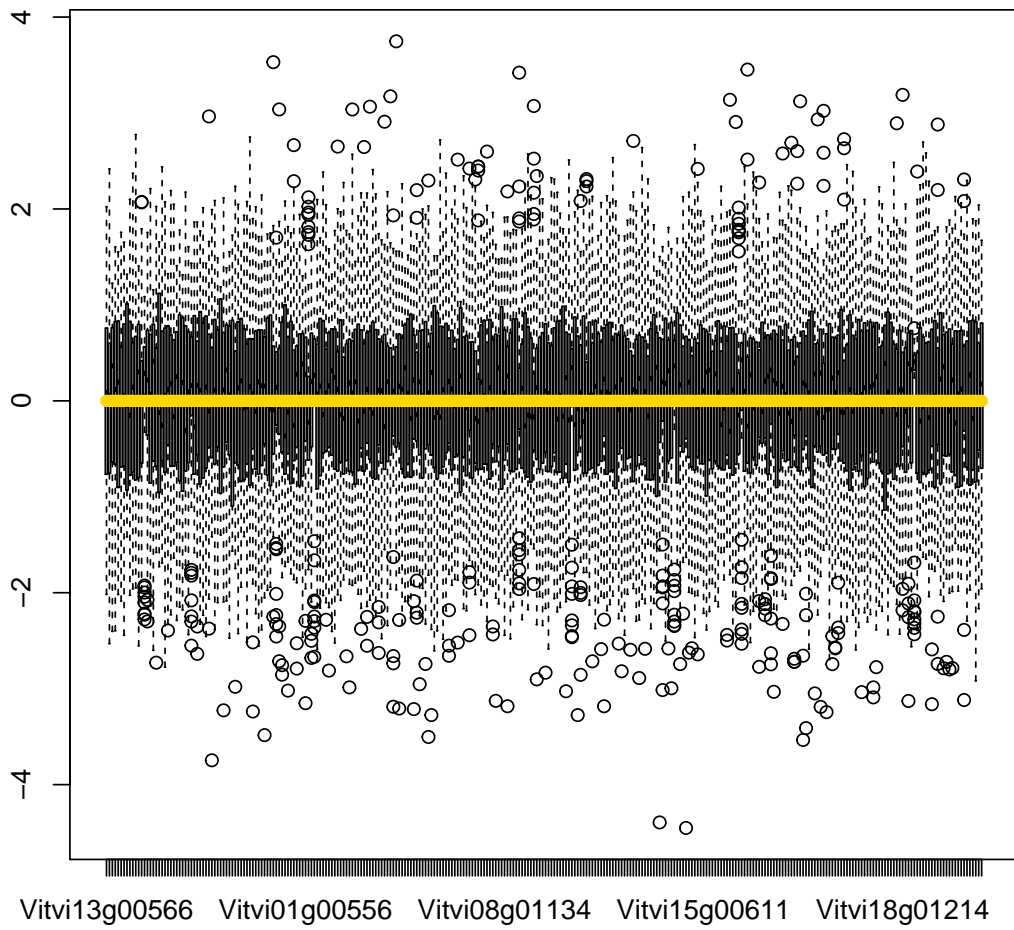
```

> range(T)
[1] -6.427422 12.489044

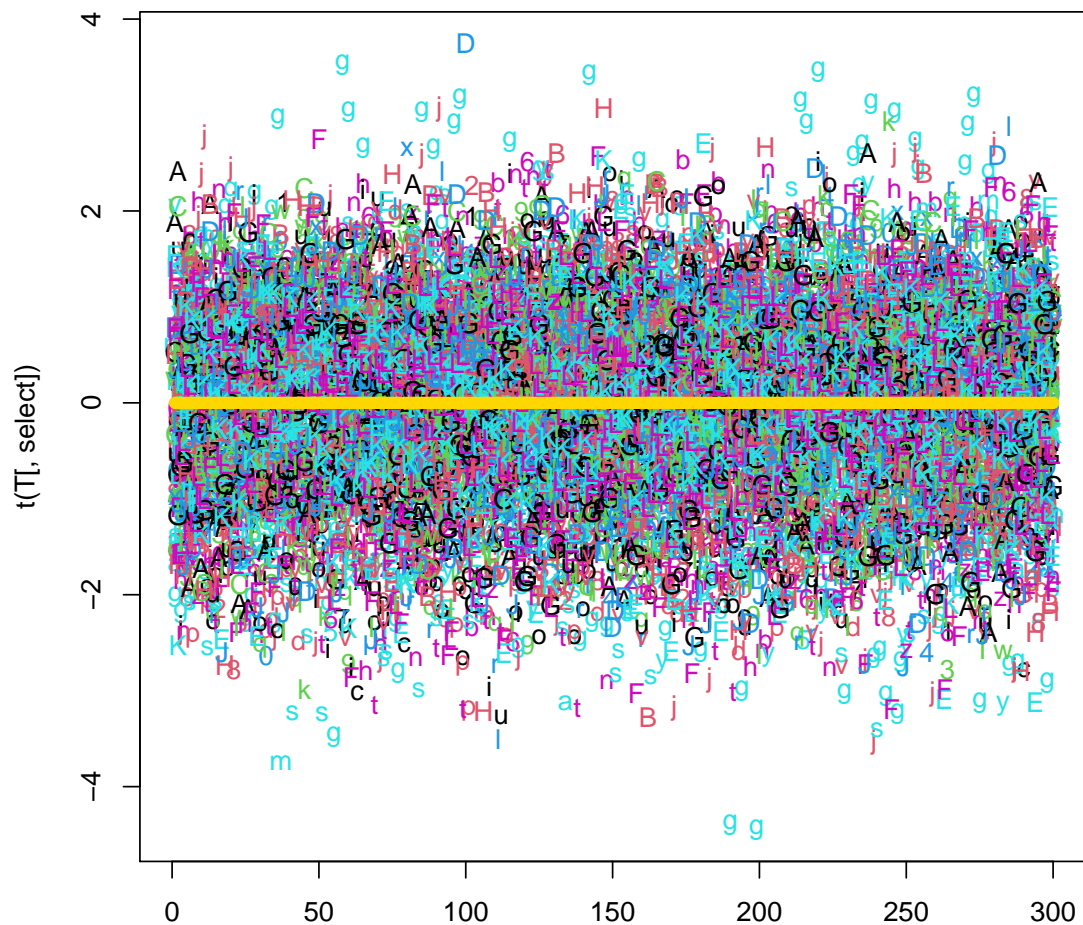
> T <- scale(T)
> range(T)
[1] -4.733874 4.806611

> select <- sample(1:ncol(T), min(ncol(T), 300))
> boxplot(T[,select])
> points(1:ncol(T[,select]), apply(T[,select], 2, mean), pch=16, col="gold")

```



```
> matplot(t(T[,select]))  
> points(1:ncol(T[,select]), apply(T[,select], 2, mean), pch=16, col="gold")
```

Check orientation, dimension names and dimensions:

```
> all(dimnames(M)[[1]]==dimnames(T)[[1]])
[1] TRUE
> dimnames(M[1:5, 1:5])
[[1]]
[1] "C18_11d_WS1" "C18_11d_WS2" "C18_11d_WS3" "C18_11d_WS4"
[5] "C18_11d_WW1"

[[2]]
[1] "Fructose"      "Glucose"      "Sucrose"
[4] "Erythronic.acid" "Galactinol"

> dimnames(T[1:5, 1:5])
[[1]]
[1] "C18_11d_WS1" "C18_11d_WS2" "C18_11d_WS3" "C18_11d_WS4"
[5] "C18_11d_WW1"

[[2]]
[1] "Vitvi01g00025" "Vitvi01g00052" "Vitvi01g00053" "Vitvi01g00064"
[5] "Vitvi01g000681"
```

```

> dim(M)
[1] 48 2049

```

Graphical overview of data

```

> library(gplots) 'gplots' was built under R version 4.0.3

```

```

Attaching package: 'gplots'
The following object is masked from 'package:stats':

```

lowess

```

> lhei <- c(1, 8)

```

```

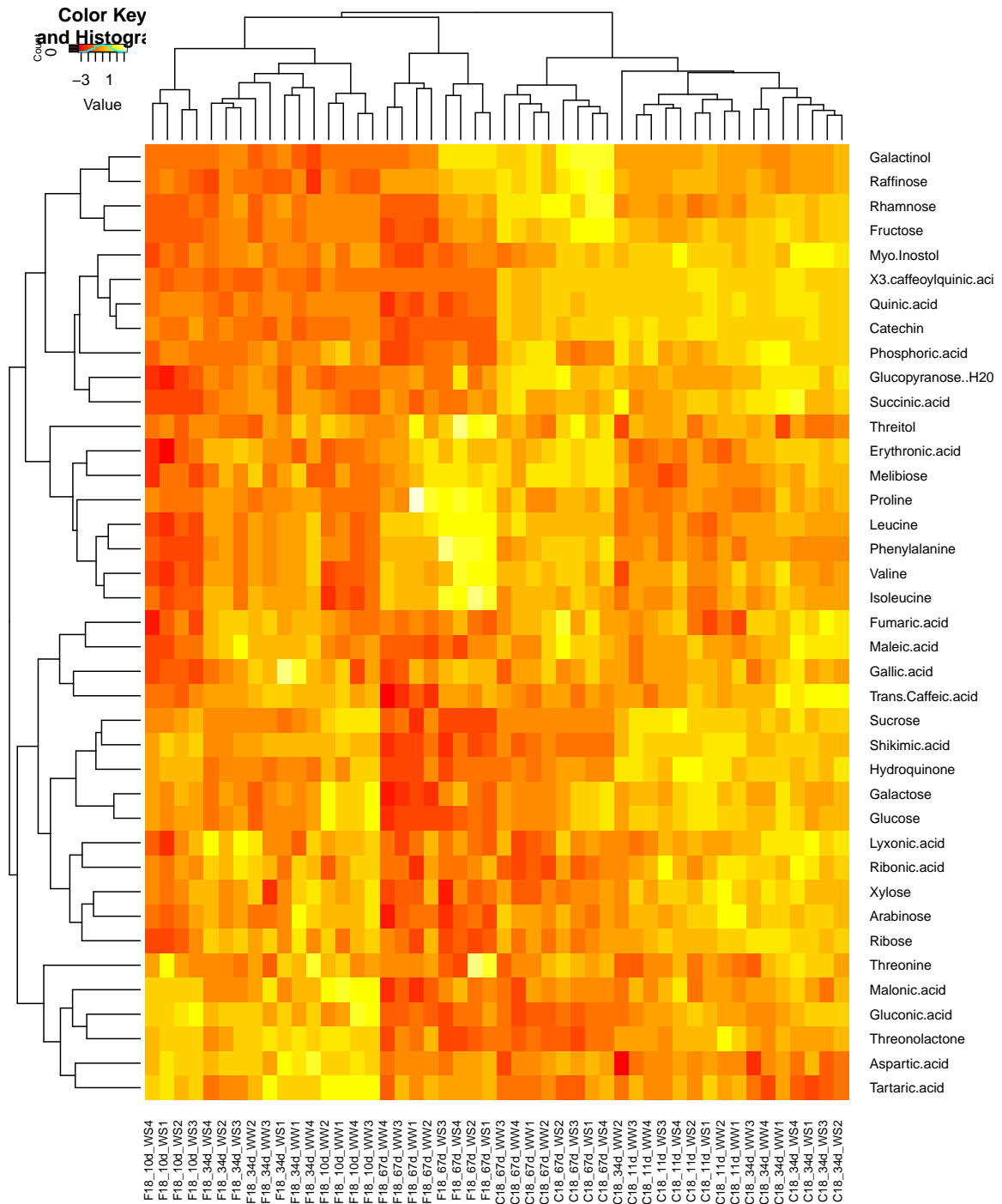
> lwid <- c(1, 6)

```

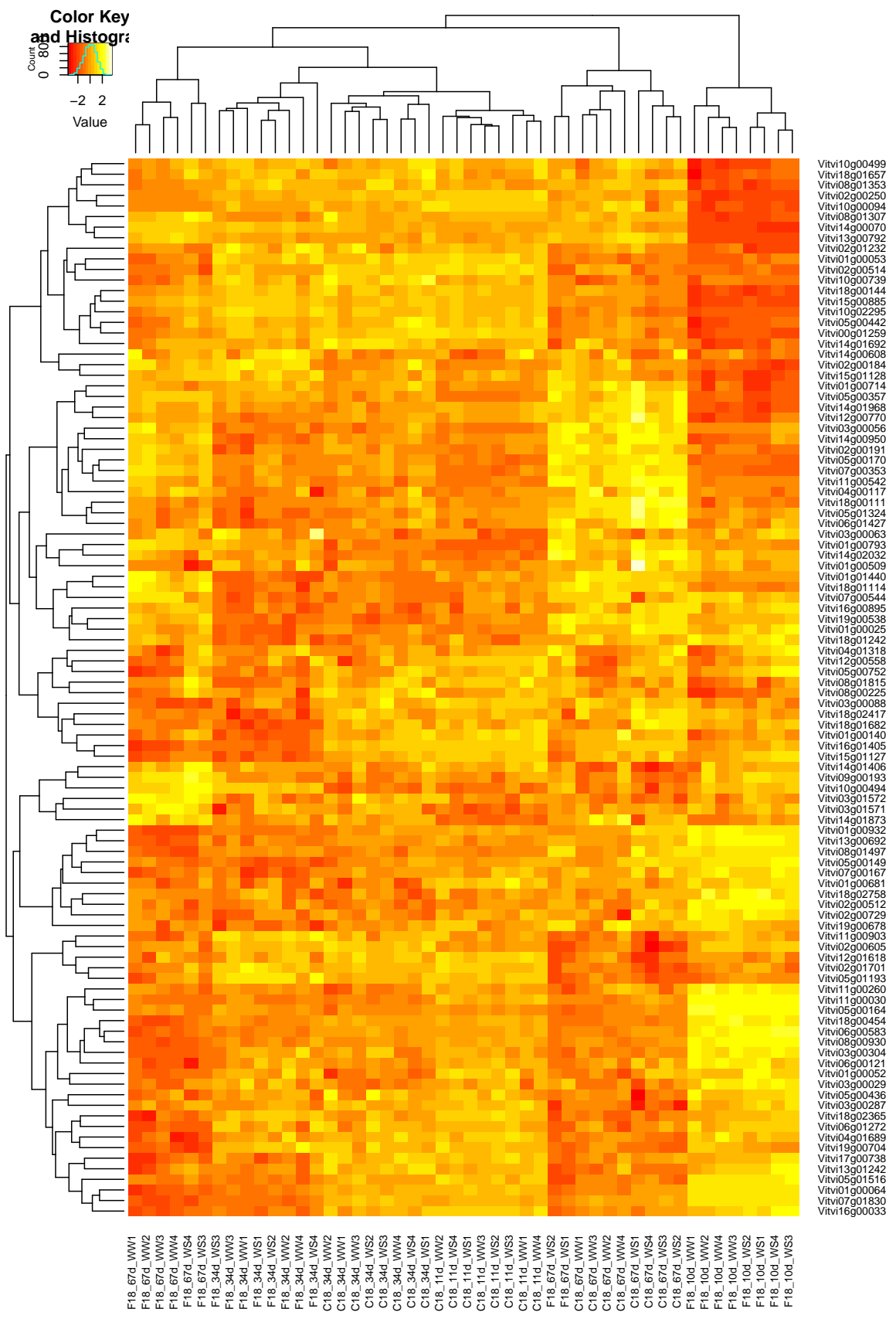
```

> heatmap.2(t(M), lhei=lhei, lwid=lwid, trace="none", margins=c(7, 7), cexCol=0.75)

```



```
> ## nf <- layout(matrix(c(1,2,3,4),2,2,byrow = TRUE), c(1,3), c(1,6), TRUE)
> ## layout.show(nf)
> ## lmat <- matrix(c(4,3,2,1),2,2,byrow = TRUE)
> ## lwid <- c(1,3)
> lhei <- c(1,8)
> lwid <- c(1,6)
> heatmap.2(t(T[,1:min(ncol(T),100)]), lhei=lhei, lwid=lwid,trace="none", marg
```



```
> cat(knit_child(file.path("../doc/", "60b_mixomics.Rnw"), quiet=TRUE))
```

5 Canonical correlation - rCCA

Dimensionality of the problem requires regularized Canonical correlation analysis (rCCA). We will follow the methods implemented in the package **mixOmics**. For explanation see <http://mixomics.org/methods/rcca/>. Quotes below are from this page.

Functions in mixOmics objects are named X and Y, so we will use the same convention and rename by alphabet order:

- X is transcripts/genes data (so far M)
- Y is metabolites data (so far T)

```
> X <- M
```

```
> dim(X)
```

```
[1] 48 39
```

```
> Y <- T
```

```
> dim(Y)
```

```
[1] 48 2049
```

```
> library(mixOmics)
```

Correlation plots

```
> library(CCA)
```

```
Warning: package 'CCA' was built under R version 4.0.3
```

```
Loading required package: fda
```

```
Warning: package 'fda' was built under R version 4.0.3
```

```
Loading required package: splines
```

```
Loading required package: Matrix
```

```
Attaching package: 'Matrix'
```

```
The following object is masked from 'package:spam':
```

```
det
```

```
Loading required package: fds
```

```
Warning: package 'fds' was built under R version 4.0.3
```

```
Loading required package: rainbow
```

```
Warning: package 'rainbow' was built under R version 4.0.3
```

```
Loading required package: pcaPP
```

```
Warning: package 'pcaPP' was built under R version 4.0.3
```

```
Loading required package: RCurl
```

```
Warning: package 'RCurl' was built under R version 4.0.3
```

```
Attaching package: 'fda'
```

The following object is masked from 'package:lattice':

melanoma

The following object is masked from 'package:graphics':

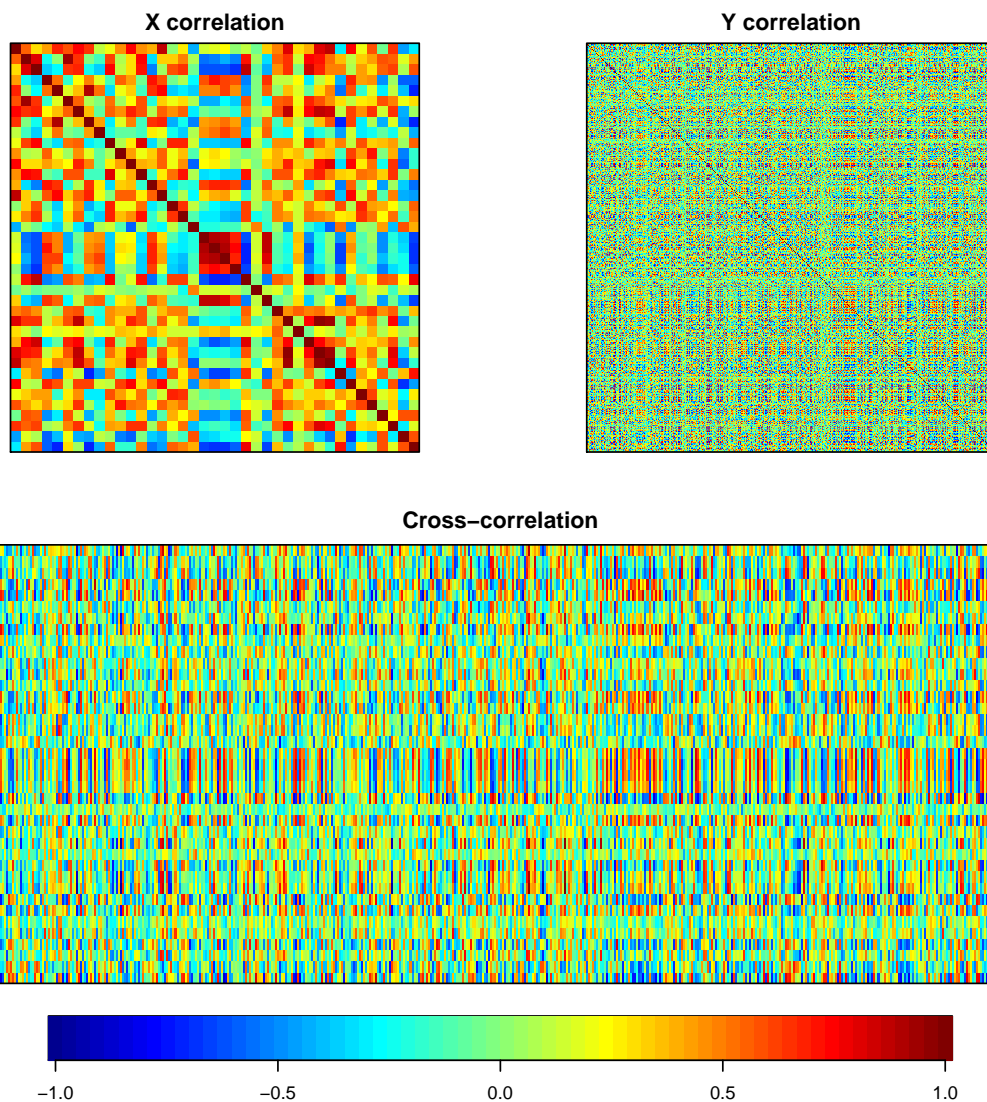
matplot

Attaching package: 'CCA'

The following object is masked from 'package:mixOmics':

rcc

```
> correl <- matcor(X, Y[, sample(1:ncol(Y), min(ncol(Y), 500))])
> img.matcor(correl, type = 2)
```



```
> detach(unload=TRUE)
```

Note: function `rcc()` is in namespaces of packages `CCA` and `mixOmics` - we want to use the latter one.

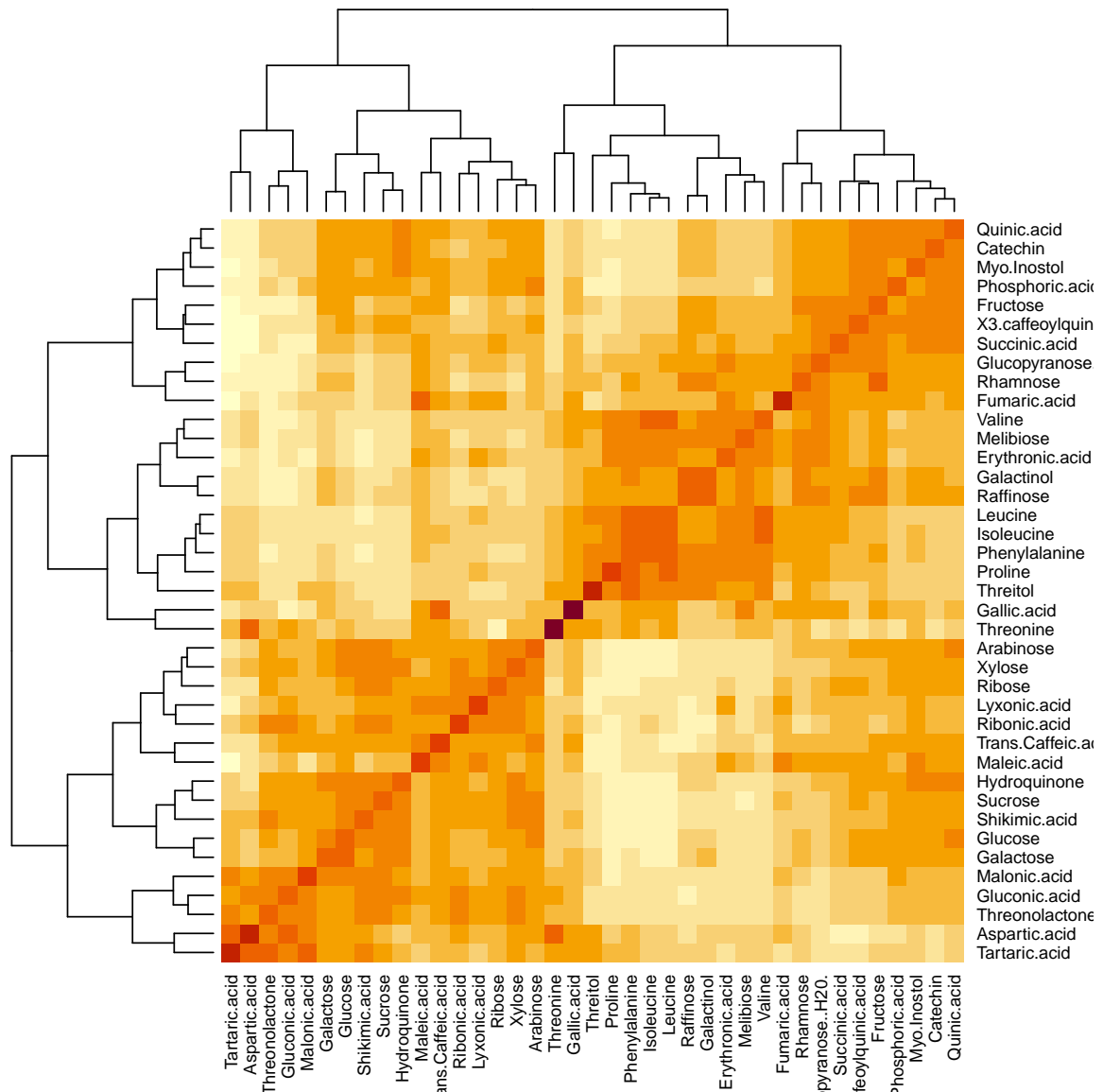
```
> bla <- getAnywhere(rcc)
> str(bla)
```

List of 5

```
$ name : chr "rcc"  
$ objs :List of 2  
..$ package:mixOmics:function (X, Y, ncomp = 2, method = c("ridge", "shrinkage"),  
lambda1 = 0, lambda2 = 0)  
..$ :function (X, Y, ncomp = 2, method = c("ridge", "shrinkage"),  
lambda1 = 0, lambda2 = 0)  
$ where : chr [1:2] "package:mixOmics" "namespace:mixOmics"  
$ visible: logi [1:2] TRUE FALSE  
$ dups : logi [1:2] FALSE TRUE  
- attr(*, "class")= chr "getAnywhere"
```

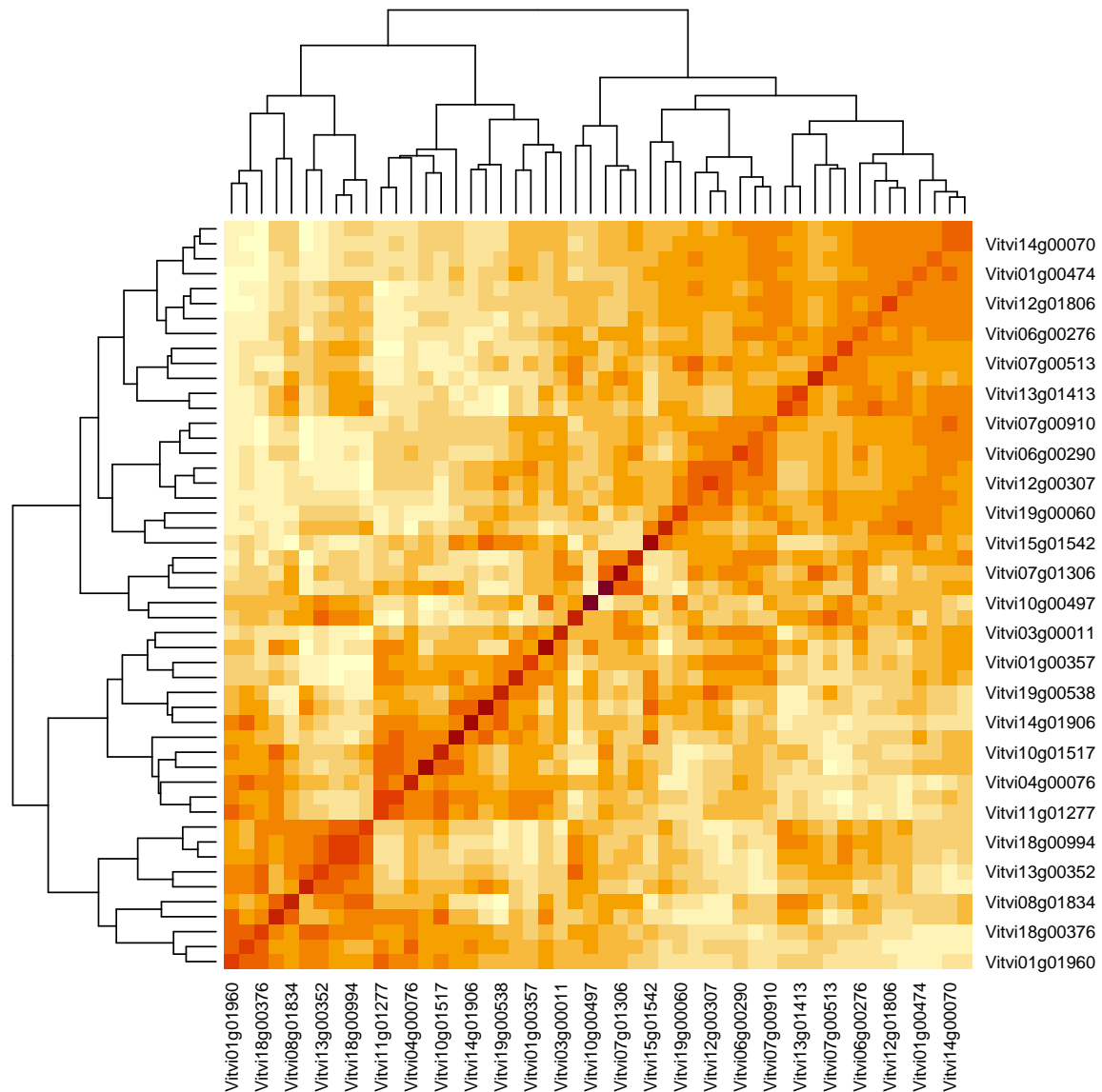
Check co-linearity of covariance matrices
Metabolites

```
> covX <- cov(X)  
> det(covX)  
[1] 6.853797e-38  
> det(cor(X))  
[1] 6.853797e-38  
> heatmap(covX)
```



Transcripts, small submatrix, determinant almost zero

```
> covY <- cov(Y[, sample(1:ncol(Y), min(50, ncol(Y)))])  
> det(covY)  
[1] 1.313543e-93  
> corY <- cor(Y[, sample(1:ncol(Y), min(50, ncol(Y)))])  
> det(corY)  
[1] -3.028298e-93  
> heatmap(covY)
```



5.0.1 Estimation of penalisation parameters

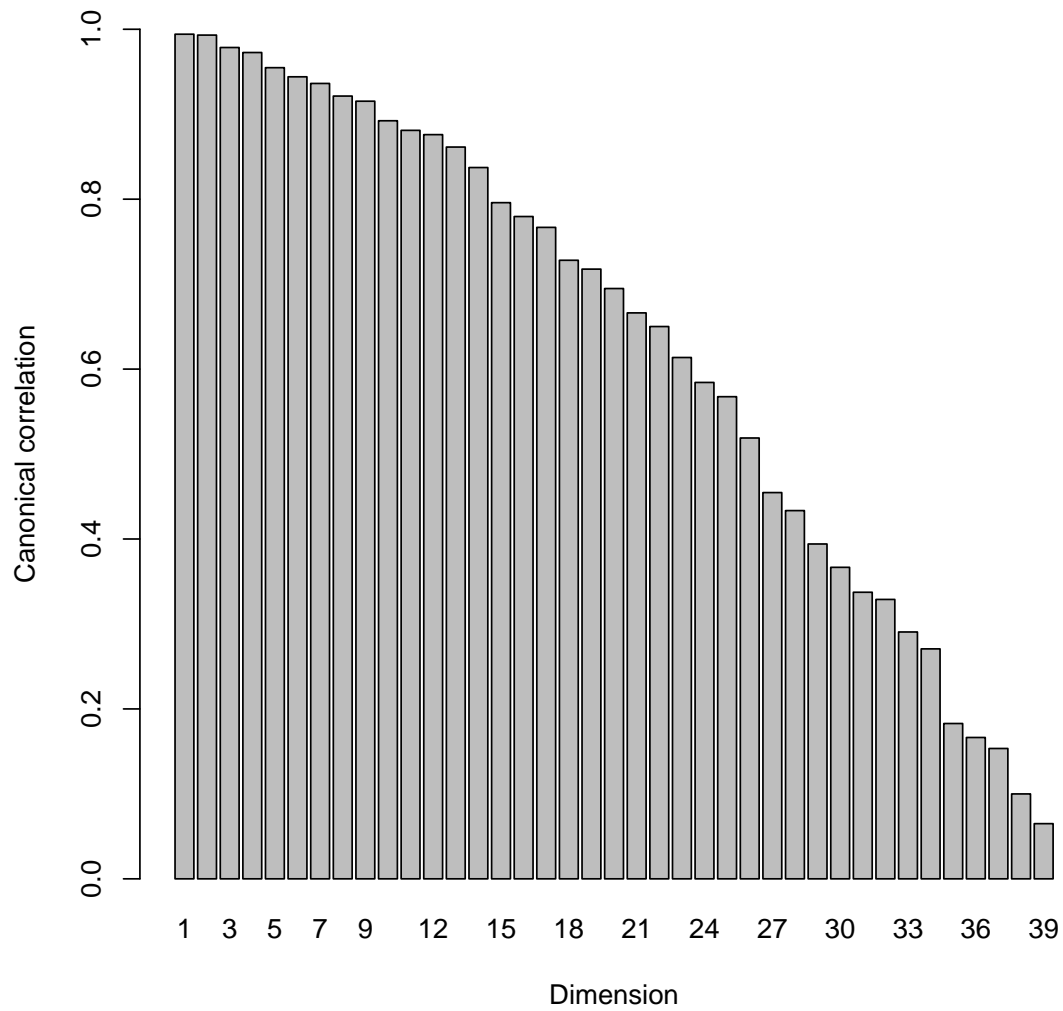
Before running rCCA, we need to tune the regularization parameters λ_1 and λ_2 . We can use the cross-validation procedure (CV), or the shrinkage method which may output different results. The shrinkage estimate method = "shrinkage" can be used to bypass tune.rcc to choose the shrinkage parameters, see ?rcc.

```
> system.time(mt.shrink <- rcc(M,T, ncomp=3, method = 'shrinkage'))
```

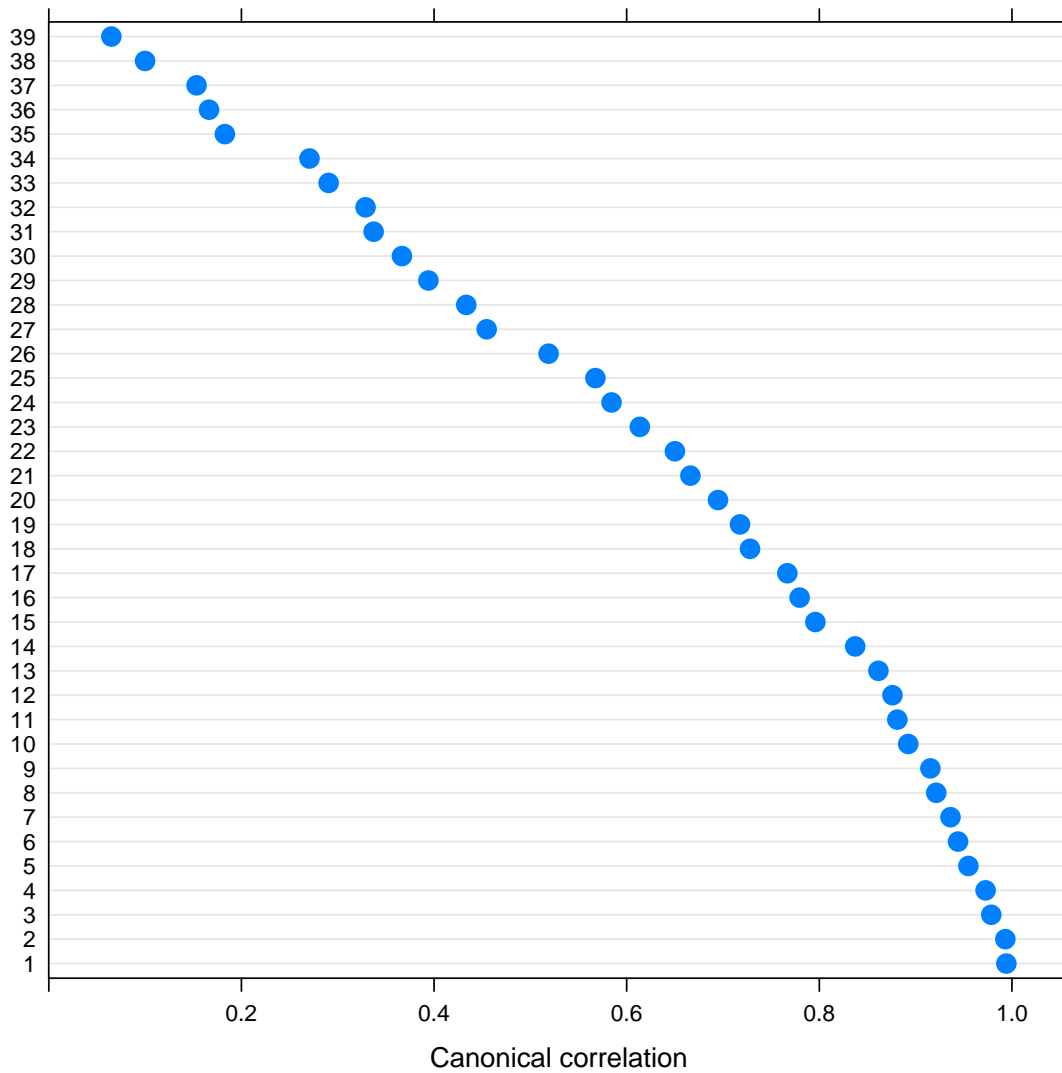


```
user  system elapsed
5.51   0.03   5.56
```

```
> plot(mt.shrink, scree.type="barplot")
```



```
> dotplot(mt.shrink$cor, xlab="Canonical correlation", pch=16, cex=1.5)
```



Parameters λ :

```

> mt.shrink$lambda
  lambda1  lambda2
0.1137600 0.1554872
> det(covX)
[1] 6.853797e-38
> det(covX+diag(mt.shrink$lambda[1],nrow(covX)))
[1] 7.957121e-18
> det(covY)
[1] 1.313543e-93
> det(covY+diag(mt.shrink$lambda[2],nrow(covY)))
[1] 2.459027e-19

```

Cross-validation

```

> if(FALSE){
+ grid1 <- seq(0, 0.2, length = 10)
+ grid2 <- seq(0.0001, 1, length = 10)
+ system.time(cv <- tune.rcc(X, Y,
+ grid1 = grid1, grid2 = grid2, validation = "loo"))
+ #
+
+ image(cv$mat, xlim=range(cv$grid1), ylim=range(cv$grid2))
+ points(mt.shrink$lambda[1],mt.shrink$lambda[2],pch=16,col=4,cex=2)
+ points(cv$opt.lambda1,cv$opt.lambda2,pch=16,col=6,cex=2)
+ legend("topright",col=c(4,6),pch=c(16,16),cex=1.5, legend=c("shrink","CV"),
+
+ #
+
+ cv$opt.lambda1
+ cv$opt.lambda2
+
+
+ # Analysis
+
+ system.time(mt.cv <- rcc(X,Y, ncomp = 3, lambda1 = cv$opt.lambda1, lambda2 =
+ }

```

5.1 Comparison

Function to omit printing small values

```

> print.blank <- function(x, eps=0, zero.print=".", shrink=TRUE, maxln=50, ...
+ nr <- nrow(x)
+ if( shrink ) x <- x[apply(x,1,function(y) any(abs(y) > eps)),]
+ x <- x[1:min(maxln,nrow(x)),]
+ print.table(local({x[abs(x)<eps] <- 0; x}),zero.print=zero.print)
+ if(maxln < nr) cat("\n", maxln, "/", nr, "rows printed\n")
+ }
> #print.blank(rbind(x,x),eps=0.1)

```

Concatenate names and bin ids. mt.shrink has original names, used for printout of the network.

```

> mt <- mt.shrink
> filter <- match(mt$names$colnames$X, rownames(mfdata))
> table(filter)

filter
 2  4  6  7  9 11 13 14 15 16 17 18 19 23 24 25 26 27 28 30 32 33 34
 1  1  1  1  1  1  1  1  1  1  1  1  1  1  1  1  1  1  1  1  1  1  1
35 38 39 40 41 42 43 45 46 47 48 49 51 52 54 55
 1  1  1  1  1  1  1  1  1  1  1  1  1  1  1

> binm <- mfdata$Bin[filter]
> head(cbind(mt$names$colnames$X,binm,mfdata[filter,1:2]))

```

	mt\$names\$colnames\$X	binm	Metabolite
Fructose	Fructose	2.2.1.1001	Fructose
Glucose	Glucose	2.2.1.1002	Glucose
Sucrose	Sucrose	2.1.1.1006	Sucrose
Erythronic.acid	Erythronic.acid	3.99.1007	Erythronic acid
Galactinol	Galactinol	3.1.1001	Galactinol
Galactose	Galactose	3.99.1010	Galactose

	Bin
Fructose	2.2.1.1001
Glucose	2.2.1.1002
Sucrose	2.1.1.1006
Erythronic.acid	3.99.1007
Galactinol	3.1.1001
Galactose	3.99.1010

```
> mt$names$colnames$X=paste( mt$names$colnames$X, binm,sep=" | ")
> #
> filter <- match(mt$names$colnames$Y, fdata$geneID)
> bing <- fdata$BINCODE[filter]
> head(cbind(mt$names$colnames$Y,bing, fdata[filter, 2:3]))
```

	mt\$names\$colnames\$Y	bing	geneID	BINCODE
478	Vitvi01g00025	2.2.1.1	Vitvi01g00025	2.2.1.1
502	Vitvi01g00052	2.2.2.6	Vitvi01g00052	2.2.2.6
503	Vitvi01g00053	2.2.2.6	Vitvi01g00053	2.2.2.6
514	Vitvi01g00064	2.2.2.2	Vitvi01g00064	2.2.2.2
981	Vitvi01g00681	35.2	Vitvi01g00681	35.2
1128	Vitvi01g00932	2.2.2.1.1	Vitvi01g00932	2.2.2.1.1

```
> mt$names$colnames$Y=paste( mt$names$colnames$Y, bing ,sep=" | ")
> head(mt$names$colnames$Y)
```

```
[1] "Vitvi01g00025 | 2.2.1.1" "Vitvi01g00052 | 2.2.2.6"
[3] "Vitvi01g00053 | 2.2.2.6" "Vitvi01g00064 | 2.2.2.2"
[5] "Vitvi01g00681 | 35.2" "Vitvi01g00932 | 2.2.2.1.1"
```

```
> #
> dimnames(mt$loadings$X) <- list(mt$names$colnames$X, paste0("Comp", 1:ncol(m
> dimnames(mt$loadings$Y) <- list(mt$names$colnames$Y, paste0("Comp", 1:ncol(m
```

5.1.1 Loadings

Metabolites

Genes with absolute nonzero loadings above the third quartile are listed
Shrink

```
> x <- cbind(mt$loadings$X)
> quantile(abs(x))
      0%      25%      50%      75%     100%
0.0003736096 0.0244578371 0.0470114282 0.0719138780 0.1689235159
> (eps <- quantile(abs(x), 0.75))
      75%
0.07191388
> print.blank(x, eps=eps)
```

	Comp1	Comp2	Comp3
Fructose 2.2.1.1001	.	0.09022347	.
Sucrose 2.1.1.1006	-0.07338038	.	.
Erythronic.acid 3.99.1007	.	.	.
Galactinol 3.1.1001	.	.	.
Myo.Inostol 3.4.1001	.	0.07672716	.
Raffinose 3.1.1002	.	.	.
Fumaric.acid 8.1.1004	.	.	.
Succinic.acid 8.1.1008	.	0.08559219	.
Rhamnose 10.6.1013	.	0.07727833	.
Ribonic.acid 10.6.1014	.	.	.
Ribose 10.6.1015	.	.	.
Aspartic.acid 13.1.1.2.1001	.	.	.
Isoleucine 13.1.4.5.1001	.	.	.
Leucine 13.1.4.4.1001	.	.	.
Phenylalanine 13.1.6.3.1001	.	.	.
Shikimic.acid 13.1.5.3.1005	-0.08005842	.	.
Threonine 13.1.3.2.1001	.	.	.
Trans.Caffeic.acid 16.2.1.1006	.	.	.
Catechin 16.8.7.1001	.	0.08468161	.
Gallic.acid 16.2.99.1045	.	.	.
Quinic.acid 16.2.99.1044	.	0.08243532	.
X3.caffeoylquinic.acid 16.2.99.1008	.	0.09393144	.
Gluconic.acid 35.1.1012	.	.	.
Glucopyranose..H2O. 35.1.1038	.	0.08224903	.
Lyxonic.acid 35.1.1023	.	.	.
Maleic.acid 35.1.1024	.	.	.
Threonolactone 35.1.1034	.	.	.
	Comp3		
Fructose 2.2.1.1001	.	.	.
Sucrose 2.1.1.1006	.	.	.
Erythronic.acid 3.99.1007	0.10364431	.	.
Galactinol 3.1.1001	-0.08426424	.	.
Myo.Inostol 3.4.1001	.	.	.
Raffinose 3.1.1002	-0.13509452	.	.
Fumaric.acid 8.1.1004	0.14852308	.	.
Succinic.acid 8.1.1008	.	.	.
Rhamnose 10.6.1013	.	.	.
Ribonic.acid 10.6.1014	0.14306573	.	.

Ribose 10.6.1015	0.08026277
Aspartic.acid 13.1.1.2.1001	0.14086677
Isoleucine 13.1.4.5.1001	0.10581865
Leucine 13.1.4.4.1001	0.09751821
Phenylalanine 13.1.6.3.1001	0.07755901
Shikimic.acid 13.1.5.3.1005	.
Threonine 13.1.3.2.1001	0.12182140
Trans.Caffeic.acid 16.2.1.1006	0.15174279
Catechin 16.8.7.1001	-0.10065233
Gallic.acid 16.2.99.1045	0.16778455
Quinic.acid 16.2.99.1044	.
X3.caffeoylquinic.acid 16.2.99.1008	-0.10141446
Gluconic.acid 35.1.1012	0.10217538
Glucopyranose..H2O. 35.1.1038	.
Lyxonic.acid 35.1.1023	0.14803593
Maleic.acid 35.1.1024	0.16892352
Threonolactone 35.1.1034	0.07266242

```
> nettip <- "txt"
> sufix <- paste(addArgs, collapse="-")
> (netfn<- gsub("\\.", "_", paste0("loadings-metabolite-", sufix)))
```

```
[1] "loadings-metabolite-18-1-1-2-3-4-5-6-7"
```

```
> pth <- .oroot
> (netpath<- file.path(pth, paste(netfn, nettip, sep=". ")))
```

```
[1] "../..output/60_Trans-x-Meta-18-1-1-2-3-4-5-6-7/loadings-metabolite-18-1-
```

```
> my.write.table(cbind(mt$loadings$X), file=netpath,
+ label="Loadings for metabolites"
+ )
```

```
Warning in write.table(x, file = file, col.names = col.names, sep = sep, : app
```

```
Object: cbind(mt$loadings$X) \\
Label: Loadings for metabolites \\
File :\\
```

```
\href{run:D:\\DEJAVNOSTI\\OMIKE\\pISA-projects\\_p_VinskaTrta\\_I_EnViRoS\\_S_
```

Transcripts

Shrink

```
> y <- cbind(mt$loadings$Y)
> quantile(abs(y))
      0%      25%      50%      75%     100%
2.217286e-07 8.133859e-04 1.770633e-03 3.178184e-03 2.152267e-02
> (eps <- quantile(abs(y), 0.75))
      75%
0.003178184
> print.blank(y, eps=eps)

              Comp1              Comp2              Comp3
Vitvi01g00052 | 2.2.2.6          . -0.003350023          .
Vitvi01g00053 | 2.2.2.6          .          . 0.004255208
Vitvi01g00681 | 35.2          . -0.003856349 -0.005365578
Vitvi02g00184 | 2.2.2.1.2        .          . 0.008052410
Vitvi02g00512 | 2.2.1.3          . -0.003430956 -0.005015523
Vitvi02g00605 | 2.2.2.1.2        .          . -0.004869284
Vitvi03g00088 | 2.2.1.3.1 -0.003363853          . 0.003472197
Vitvi03g01571 | 2.2.2.1.1 0.004350121          .          .
Vitvi03g01572 | 2.2.2.1.1          .          . -0.004062375
Vitvi05g00357 | 2.2.2.1.2          .          . 0.003630840
Vitvi05g01516 | 2.2.1.1 -0.003834300          .          .
Vitvi06g01272 | 2.2.1.4 -0.003554102          .          .
Vitvi07g00167 | 2.1.2.4          .          . -0.006022954
Vitvi07g00353 | 2.2.1.5          .          . 0.003775482
Vitvi08g01815 | 2.1.2.60          . 0.008074499 0.015648271
Vitvi09g00193 | 2.2.1.3          .          . -0.004337168
Vitvi10g00494 | 2.2.2.2          . -0.003892783          .
Vitvi10g00739 | 2.1.2.2          . 0.004546677 0.003187458
Vitvi11g00260 | 2.2.1.4          .          . -0.005000296
Vitvi12g00770 | 2.1.2.1          .          . -0.004022522
Vitvi12g01618 | 2.1.1.2          .          . -0.007902341
Vitvi13g01242 | 2.2.2.10          .          . -0.003529797
Vitvi14g00608 | 2.1.2          . 0.004068240 0.021522670
Vitvi14g00950 | 2.2.1.1          .          . -0.006501129
Vitvi14g01406 | 2.2.2.1.1          .          . 0.003800408
Vitvi14g01873 | 2.1.2.1          .          . -0.006629932
Vitvi15g01127 | 2.2.2.1.2          .          . -0.005451282
Vitvi16g00033 | 2.2.2.6 -0.004039838          .          .
Vitvi16g01405 | 2.1.2.2          . 0.004219532          .
Vitvi17g00738 | 2.1.2.1          . 0.004253792 0.003523087
Vitvi18g00144 | 2.2.2.1.1          .          . -0.005729033
Vitvi18g02758 | 2.1.2.1          .          . -0.003396870
Vitvi18g01242 | 2.1.2          .          . -0.005519885
Vitvi18g01682 | 2.2.1.3.1 -0.003249522          .          .
Vitvi18g02365 | 2.1.1.1          . -0.003353421          .
Vitvi18g02417 | 2.2.1.1          . 0.006777687          .
Vitvi19g00538 | 2.1.2.3          .          . -0.004318830
Vitvi19g00678 | 2.2.2.1.2          . 0.004108663 0.003824114
Vitvi02g01701 | 3.2.2          .          . -0.004176305
Vitvi01g00140 | 3.4.5          .          . -0.004752220
```

```

Vitvi01g00509 | 3.2.3      -0.003659919      .      .
Vitvi01g00793 | 3.2.3      .      .      0.004384227
Vitvi01g01440 | 3.5      .      .      -0.006530247
Vitvi03g00063 | 3.5      .      -0.005466330      -0.005381996
Vitvi03g00287 | 3.4.5      -0.004730244      .      0.006777955
Vitvi04g00117 | 3.5      .      0.005233474      0.005169947
Vitvi04g01318 | 3.5      0.003658369      .      0.008259410
Vitvi04g01689 | 3.6      -0.006910607      .      .
Vitvi05g00170 | 3.1.1.1      .      .      -0.004110894
Vitvi05g00436 | 35.2      .      .      0.003720687

```

50 / 2049 rows printed

```

> nettip <- "txt"
> suffix <- paste(addArgs, collapse="-")
> (netfn<- gsub("\\.", "_", paste0("loadings-metabolite-", suffix)))

```

```
[1] "loadings-metabolite-18-1-1-2-3-4-5-6-7"
```

```

> pth <- .oroot
> (netpath<- file.path(pth, paste(netfn, nettip, sep=".")))

```

```
[1] ".././output/60_Trans-x-Meta-18-1-1-2-3-4-5-6-7/loadings-metabolite-18-1-
```

```

> my.write.table(cbind(mt$loadings$X), file=netpath,
+   label="Loadings for genes"
+   )

```

Warning in write.table(x, file = file, col.names = col.names, sep = sep, : app

```
Object: cbind(mt$loadings$X) \\
```

```
Label: Loadings for genes \\
```

```
File :\\
```

```
\href{run:D:\\DEJAVNOSTI\\OMIKE\\pISA-projects\\_p_VinskaTrta\\_I_EnViRoS\\_S_
```


Upper 5%

```
> y <- cbind(mt$loadings$Y)
> quantile(abs(y))
      0%      25%      50%      75%     100%
2.217286e-07 8.133859e-04 1.770633e-03 3.178184e-03 2.152267e-02
> (eps <- quantile(abs(y), 0.95))
      95%
0.006414837
> print.blank(y, eps=eps)
```

		Comp1	Comp2	Comp3
Vitvi02g00184	2.2.2.1.2	.	.	0.008052410
Vitvi08g01815	2.1.2.60	.	0.008074499	0.015648271
Vitvi12g01618	2.1.1.2	.	.	-0.007902341
Vitvi14g00608	2.1.2	.	.	0.021522670
Vitvi14g00950	2.2.1.1	.	.	-0.006501129
Vitvi14g01873	2.1.2.1	.	.	-0.006629932
Vitvi18g02417	2.2.1.1	.	0.006777687	.
Vitvi01g01440	3.5	.	.	-0.006530247
Vitvi03g00287	3.4.5	.	.	0.006777955
Vitvi04g01318	3.5	.	.	0.008259410
Vitvi04g01689	3.6	-0.006910607	.	.
Vitvi07g00431	3.1.2.2	.	.	-0.007021599
Vitvi08g00789	3.5	.	.	-0.006623847
Vitvi08g01260	3.5	.	.	0.008773947
Vitvi08g01536	3.5	.	.	0.011641046
Vitvi11g01211	3.5	.	-0.010144953	-0.006666015
Vitvi18g00384	3.2.2	.	.	-0.007224775
Vitvi18g02757	3.5	.	.	0.009667680
Vitvi19g00016	3.5	.	.	0.007296604
Vitvi10g01386	8.1.6	.	.	-0.007017027
Vitvi11g00147	8.1.1.2	.	.	0.009145702
Vitvi13g00265	8.1.5	.	.	-0.007138941
Vitvi19g01739	8.2.99	.	.	-0.008816341
Vitvi19g01771	8.1.4	.	.	-0.009697472
Vitvi02g01716	10.5.4	0.006659027	.	.
Vitvi07g02943	10.2.1	.	.	0.006600524
Vitvi01g00784	10.7	.	.	-0.006668965
Vitvi02g01349	10.2.1	.	.	0.007512734
Vitvi02g00642	10.1	.	.	-0.009345031
Vitvi02g01662	10.8.2	.	.	-0.008712110
Vitvi03g00209	10.7	.	.	-0.007884942
Vitvi05g00276	10.8.2	.	.	-0.008275578
Vitvi05g01266	10.6.2	.	-0.006891997	.
Vitvi06g00481	10.7	.	.	-0.012035399
Vitvi07g00401	10.6.2	.	-0.008567279	-0.008997301
Vitvi07g02226	10.6.3	.	.	-0.007924185
Vitvi08g00235	10.5.1.1	.	.	-0.008514575
Vitvi08g00240	10.5.1.1	.	.	-0.007719087
Vitvi08g01070	10.3.2	.	-0.007057781	-0.009629198
Vitvi08g01098	10.1.5	.	-0.008006948	-0.011620103
Vitvi09g00442	10.2	.	.	-0.008379189
Vitvi11g00013	10.8.1	.	.	0.009815691

Vitvi11g00809		10.1.8	.	.	0.010239731
Vitvi12g00024		10.5	.	.	0.008361529
Vitvi12g00131		10.6.3	.	-0.006882861	-0.006492061
Vitvi12g00586		10.6.2	.	.	-0.007340986
Vitvi12g00721		10.5.1.1	.	.	-0.006708768
Vitvi14g01635		10.6.3	.	.	-0.012469507
Vitvi14g01995		10.8.1	.	0.007145193	.
Vitvi15g01565		10.5.3	.	.	-0.008993780

50 / 2049 rows printed

5.2 Summary

```
> mt$explained_variance
```

```
$X
```

	comp 1	comp 2	comp 3
	0.32152633	0.28496470	0.08874357

```
$Y
```

	comp 1	comp 2	comp 3
	0.21421573	0.16082685	0.08676566

5.3 Plots

```
> variety <- factor(substr(mt$names$sample, 1, 1))
> status <- as.numeric(factor(substr(mt$names$sample, 10, 10)))
> day <- as.numeric(factor(substr(mt$names$sample, 5, 5)))

> my.plotIndiv <- function(x, comp=c(1,2), col=c(4,2), pch=c(16,1), grid=TRUE,
+   variety <- factor(substr(x$names$sample, 1, 1))
+   statusf <- factor(substr(x$names$sample, 10, 10))
+   status <- as.numeric(statusf)
+   dayf <- factor(substr(x$names$sample, 5, 5))
+   day2 <- substr(x$names$sample, 5, 6)
+   day2[day2=="11"] <- "10"
+   day2 <- factor(day2)
+   day <- as.numeric(dayf)
+   #
+   cols <- col[as.numeric(variety)]
+   pchs <- pch[status]
+   cexs <- day/2
+   #
+   if(missing(title)) title <- x$names$data
+   #
+   if(length(comp)==2 & !shift){
+     par(mfrow=c(1,2), mar=c(5,3,2,0.5))
+     for(i in 1:2){
+       xy <- x$variates[[i]][,comp]
+       xlab <- paste("Variate", comp[1])
+       ylab <- paste("Variate", comp[2])
+       plot(xy[,1], xy[,2], type="n", xlab="", ylab="", axes=FALSE, ann=FALSE, asp=
+         title(title[i])
+         if(grid) abline(h=seq(-4,4,0.5), v=seq(-4,4,0.5), col="grey90", lwd=0.1)
+         box()
+         axis(1, cex.axis=0.75)
+         axis(2, las=2, cex.axis=0.75)
+         if(i ==1) mtext(ylab, 2, line=1.5)
+         if(i ==1) {
+ legend( par("usr")[1]*1.3, par("usr")[3]*1.2
+       , pch=c(NA, NA, unique(pchs), 1, 1, 1)
+       , text.col=c(unique(cols), 1, 1, 1, 1, 1)
+       , pt.cex=c(rep(1,4), unique(cexs))
+       , legend=c(levels(variety), levels(statusf), levels(day2))
+       , bty="n"
+       , xpd=TRUE
+       , horiz=TRUE
+       )
+     }
+     mtext(xlab, 1, line=-4, outer=TRUE, adj=0.55)
+
+     points(xy[,1], xy[,2], cex=cexs, pch=pchs, col=cols, ...)
+   }
+ }
+ #
+ if(length(comp)==3 & shift){
```

```

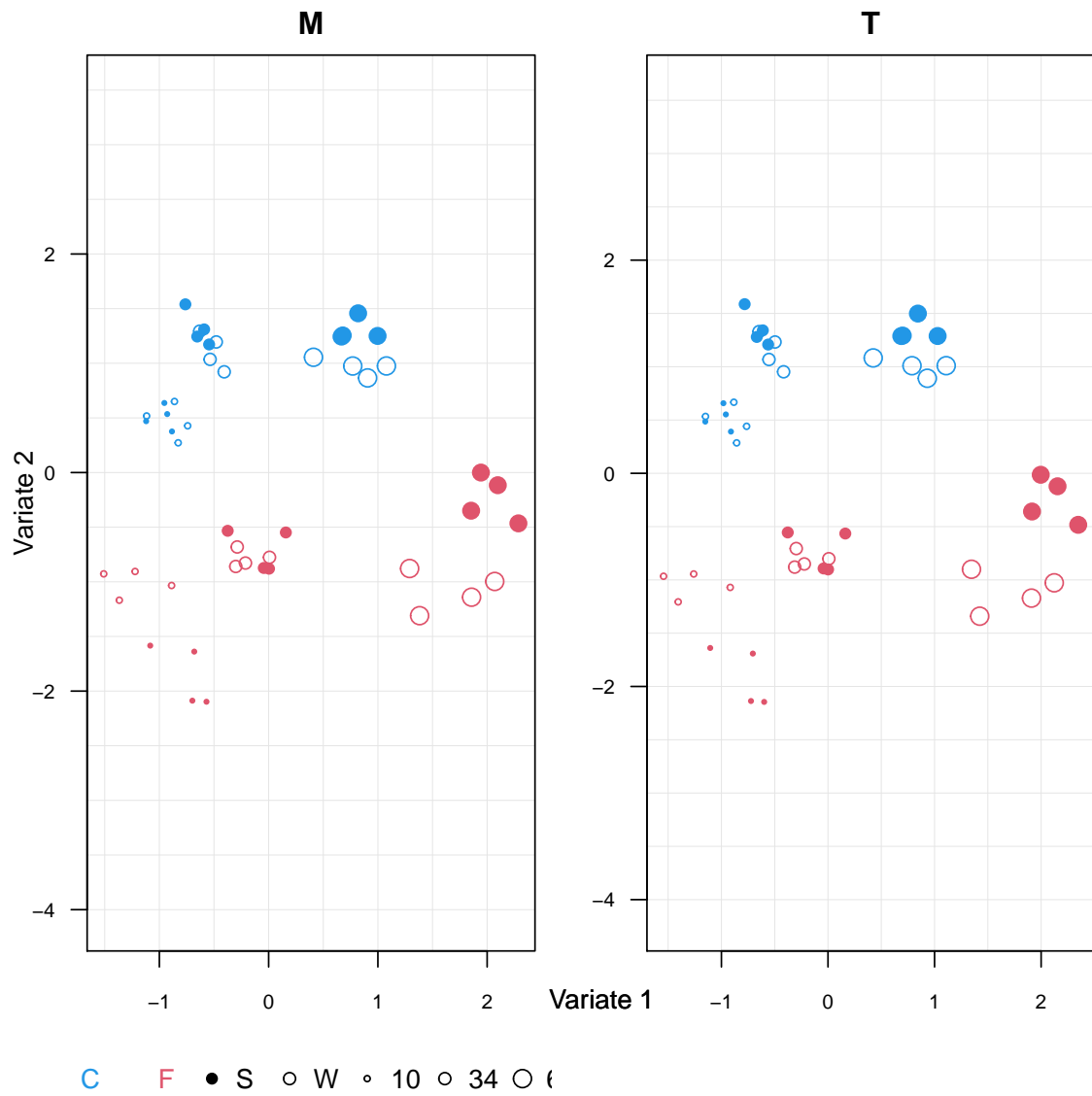
+   par(mfrow=c(1,2), mar=c(5.5,3,2,0.5))
+   for(i in 1:2){
+     xy <- x$variates[[i]][,comp]
+     xlab <- paste("Variate",comp[1])
+     ylab <- paste("Variate",comp[2])
+     plot(xy[,1],xy[,2],type="n",xlab="",ylab="",axes=FALSE,ann=FALSE, asp=
+     title(title[i])
+     if(grid) abline(h=seq(-4,4,0.5),v=seq(-4,4,0.5),col="grey90",lwd=0.1)
+     box()
+     axis(1,cex.axis=0.75)
+     axis(2, las=2,cex.axis=0.75)
+     if(i ==1) mtext(ylab,2,line=1.5)
+     if(i ==1) {
+ #       legend("topright"
+ legend( par("usr")[1]*1.3,par("usr")[3]*1.2
+       , pch=c(NA,NA,unique(pchs),1,1,1)
+       , text.col=c(unique(cols),1,1,1,1,1)
+       , pt.cex=c(rep(1,4),unique(cexs))
+       , legend=c(levels(variety),levels(statusf), levels(day2))
+       , bty="n"
+       , xpd=TRUE
+       , horiz=TRUE
+       )
+     }
+     mtext(xlab,1,line=-4,outer=TRUE, adj=0.55)
+
+     points(xy[,1],xy[,2],cex=cexs,pch=pchs,col=cols, ...)
+     segments(xy[,1],xy[,2],xy[,1],xy[,3],col=cols)
+   }
+ }
+
+ #
+ if(length(comp)==2 & shift){
+   par(mfrow=c(1,1), mar=c(6,4,2,0.5))
+   xy <- x$variates[[1]][,comp]
+   xy2 <- x$variates[[2]][,comp]
+   xlab <- paste("Variate",comp[1])
+   ylab <- paste("Variate",comp[2])
+   plot(xy[,1],xy[,2],type="n",xlab="",ylab="",axes=FALSE,ann=FALSE, asp=
+   title(title[i])
+   if(grid) abline(h=seq(-4,4,0.5),v=seq(-4,4,0.5),col="grey90",lwd=0.1)
+   box()
+   axis(1,cex.axis=0.75)
+   axis(2, las=2,cex.axis=0.75)
+   mtext(ylab,2,line=2)
+   if(1 ==1) {
+ legend( par("usr")[1]*1.05,par("usr")[3]*1.3
+     , pch=c(NA,NA,unique(pchs),1,1,1)
+     , text.col=c(unique(cols),1,1,1,1,1)
+     , pt.cex=c(rep(1,4),unique(cexs))
+     , legend=c(levels(variety),levels(statusf), levels(day2))
+     , bty="n"

```

```

+         , xpd=TRUE
+         , horiz=TRUE
+       )
+     }
+     mtext(xlab, 1, line=-3, outer=TRUE)
+     legend("topright", pch=c(1, NA), lwd=c(NA, 2), legend=c("M", "T"), bty="n")
+
+     points(xy[, 1], xy[, 2], cex=cexs, pch=pchs, col=cols, ...)
+     segments(xy[, 1], xy[, 2], xy2[, 1], xy2[, 2], col=cols)
+   }
+
+
+
+
+ #
+ if(length(comp)==3 & !shift){
+   par(mfrow=c(1, 2), mar=c(5, 3, 2, 0.5))
+   for(i in 1:2){
+     persp.xyz(x$variates[[i]], cex=cexs, pch=pchs, col=cols, ...)
+     title(title[i])
+   }
+ }
+ }
+ }
> my.plotIndiv(mt, comp=1:2)

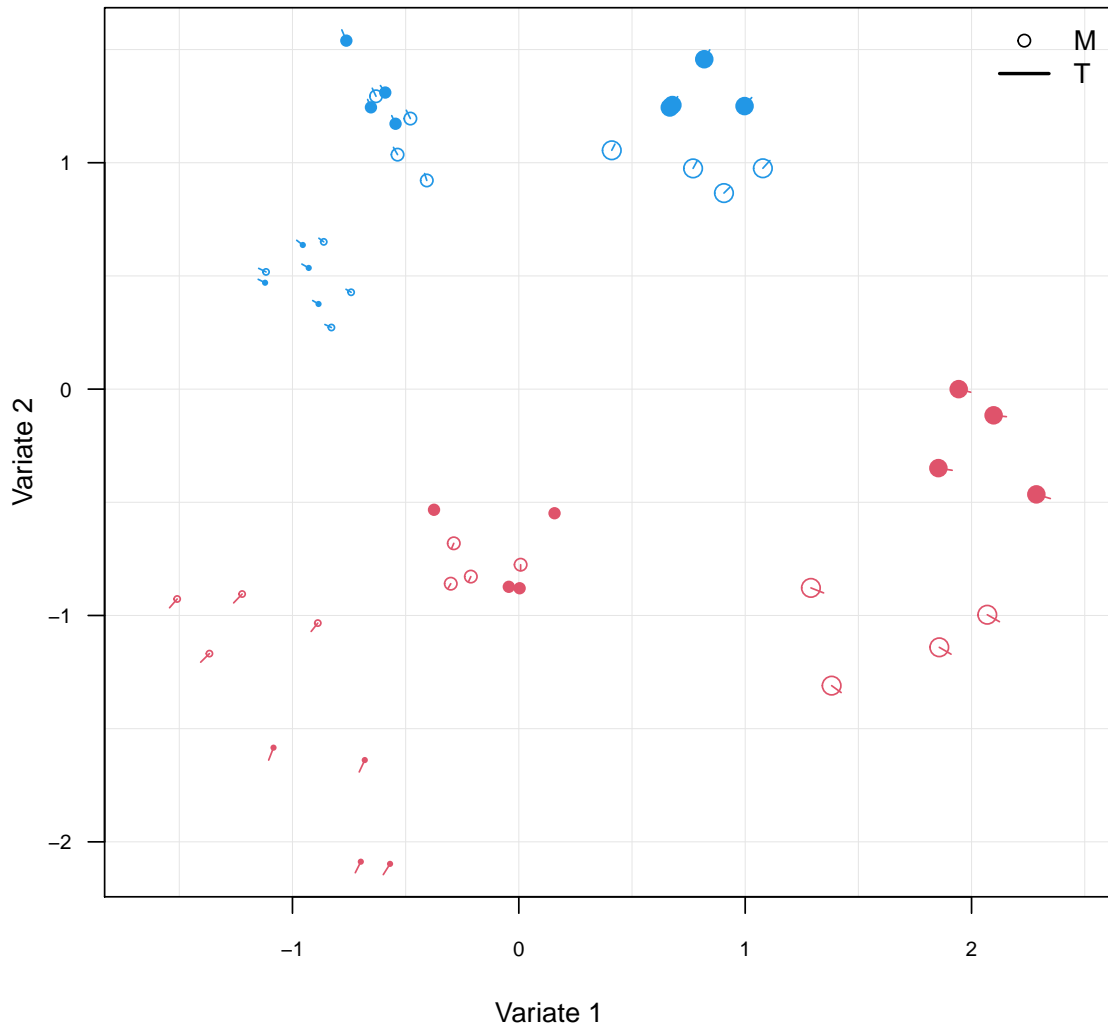
```



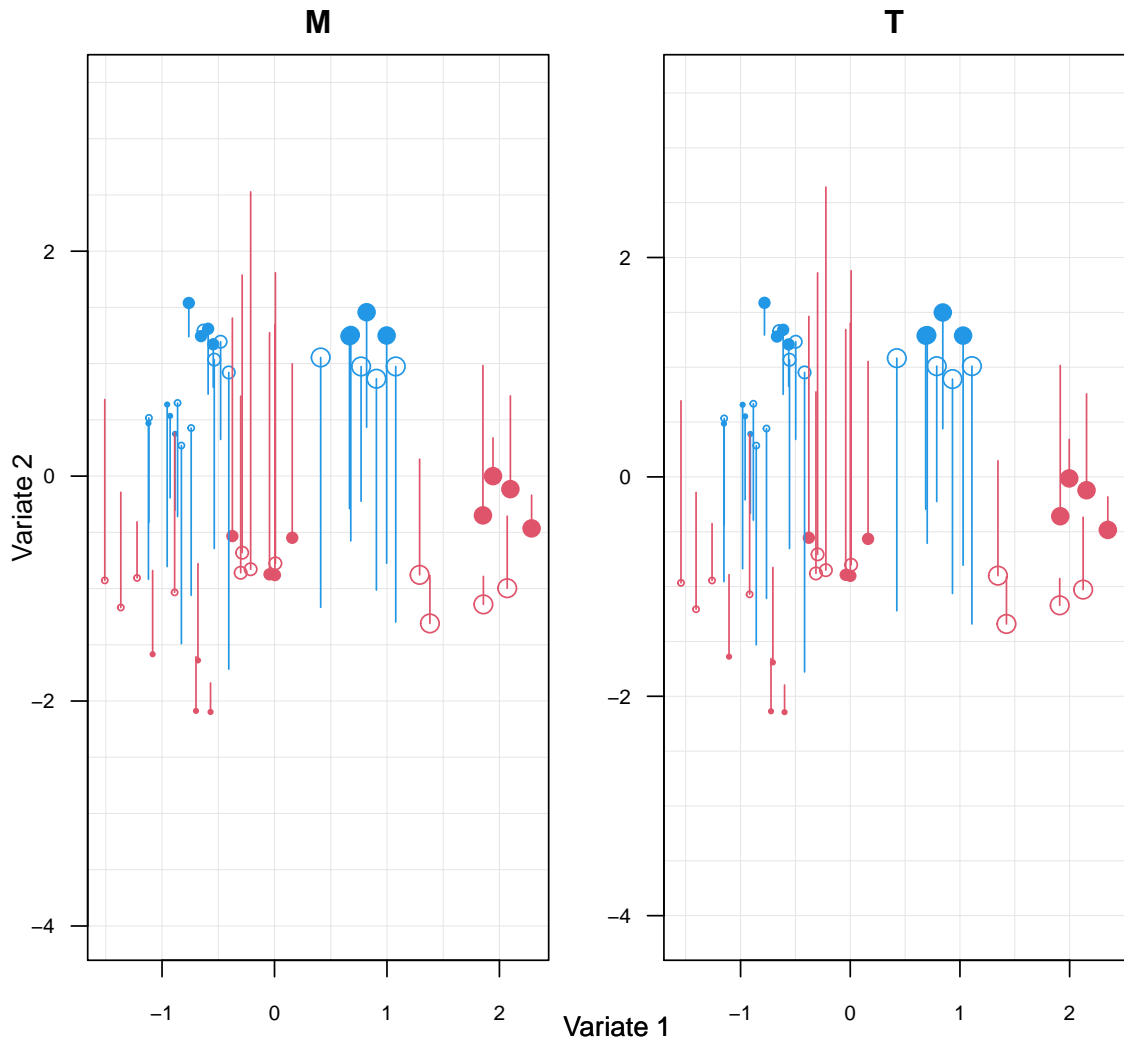
@

```

> # cabernet blue
> my.plotIndiv(mt, comp=1:2, shift=TRUE)
  
```



```
> my.plotIndiv(C, S, W, 10, 34, 67, mt, comp=1:3, shift=TRUE)
```



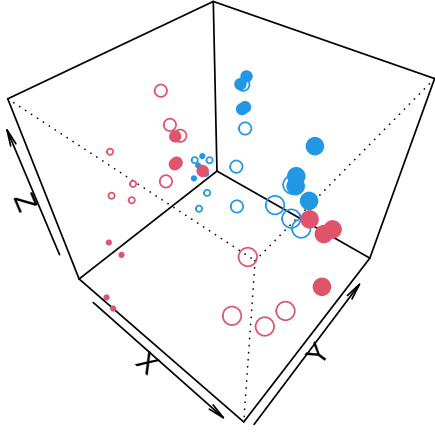
C F • S ○ W ◦ 10 ○ 34 ○ (

```
> my.plotIndiv(mt, comp=1:3, side="", phi=45, theta=40)
```

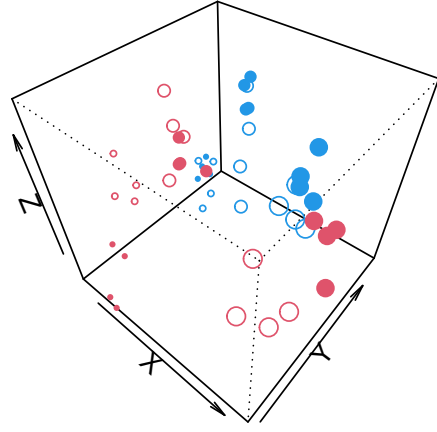
```
[1] 3
```

```
[1] 3
```


M



T

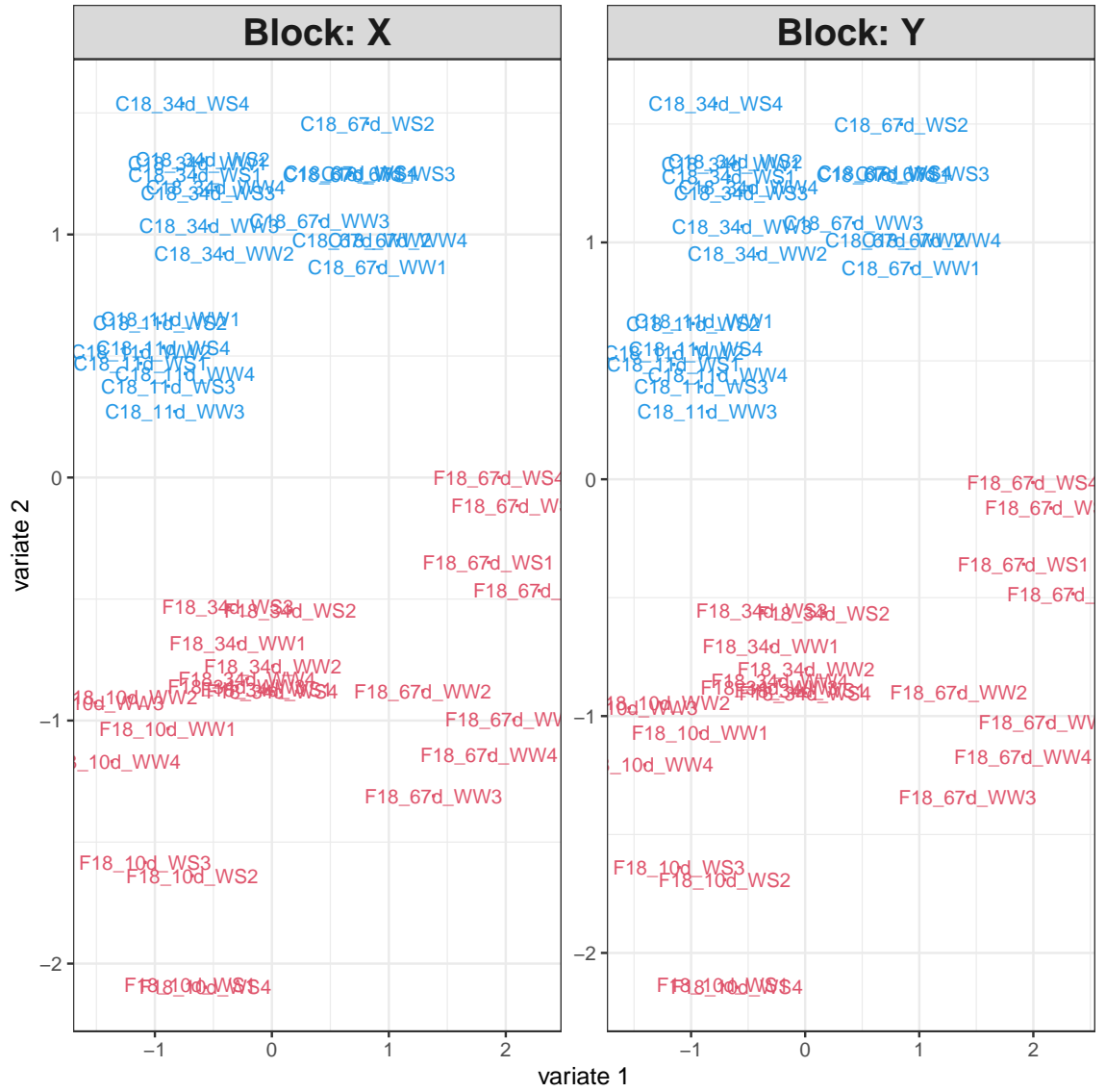


```
> stereo(mt, turn=-40, elevation=30)
```

```

> plotIndiv(mt, comp=c(1,2)
+ , group=variety
+ , col.per.group=c(4,2)
+ )

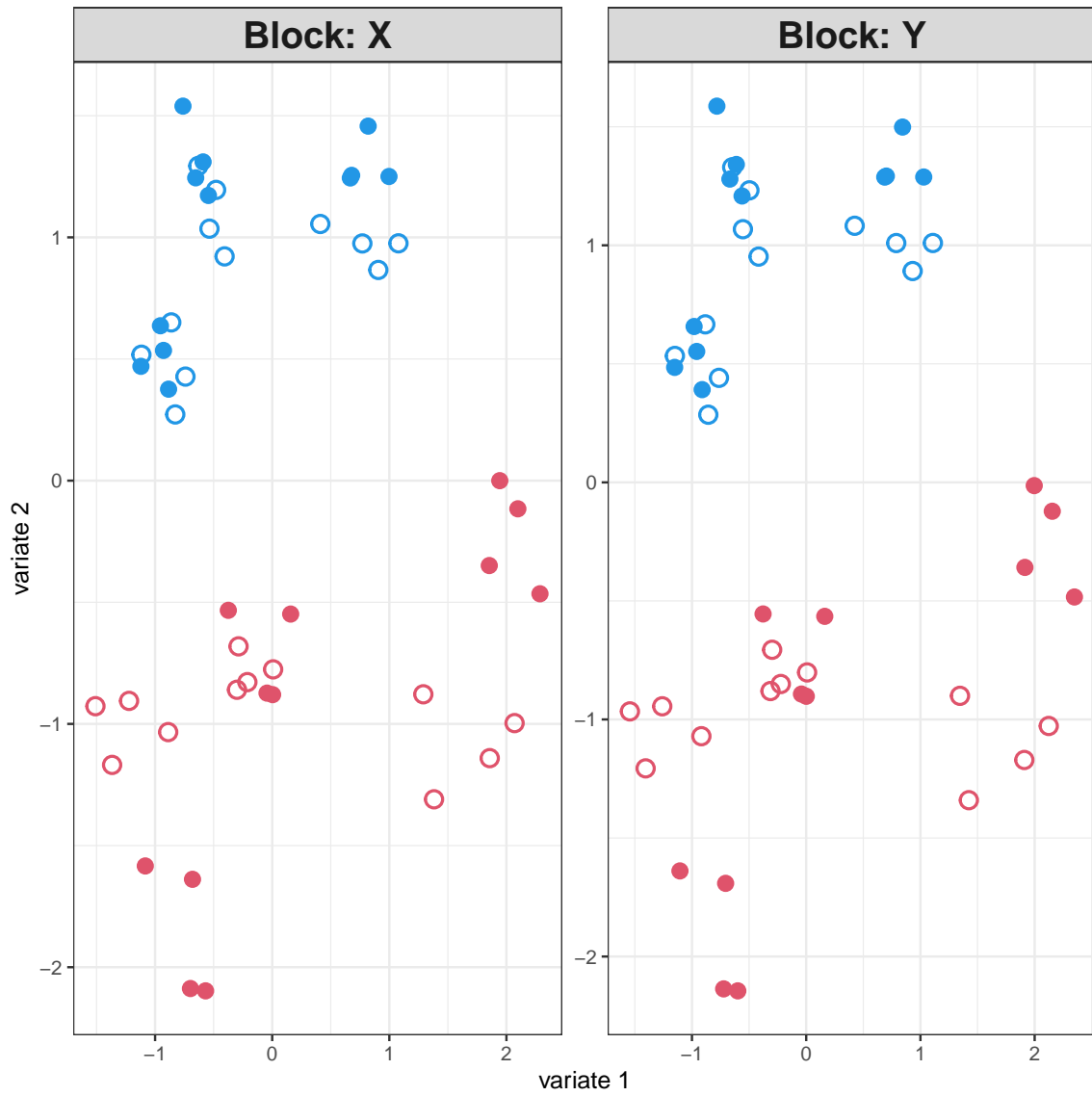
```



```

> plotIndiv(mt, comp=c(1,2)
+ , ind.names=FALSE
+ , group=variety
+ , col.per.group=c(4,2)
+ , pch=c(16,1)[status]
+ #, cex=c(1,1.5,2)[day]
+ #, star=TRUE
+ #, style="graphics"
+ )

```



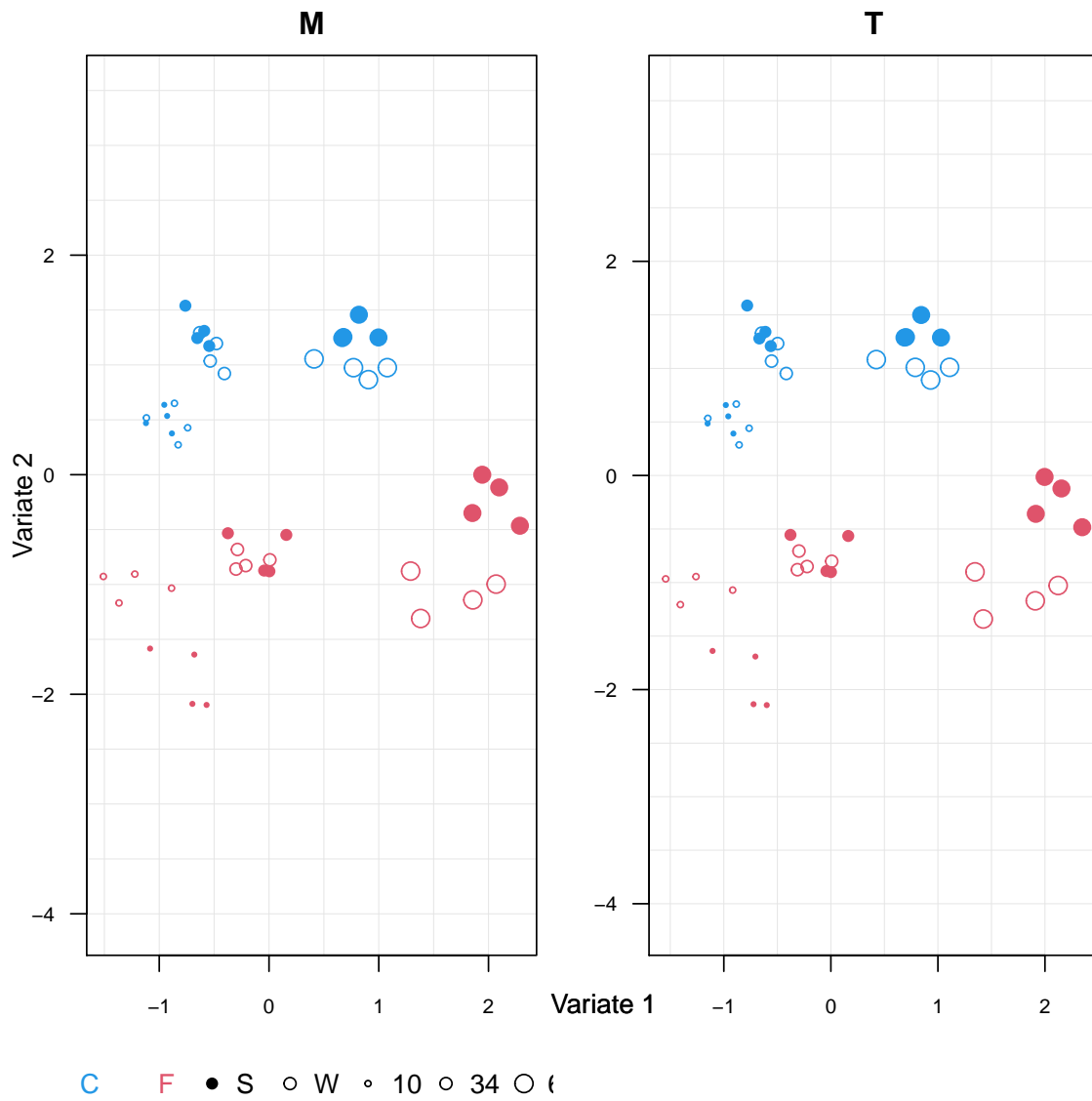


Figure 5: Samples plot for Components 1 and 2.

```
> my.plotIndiv(mt, comp=c(1,2)
+ )
```

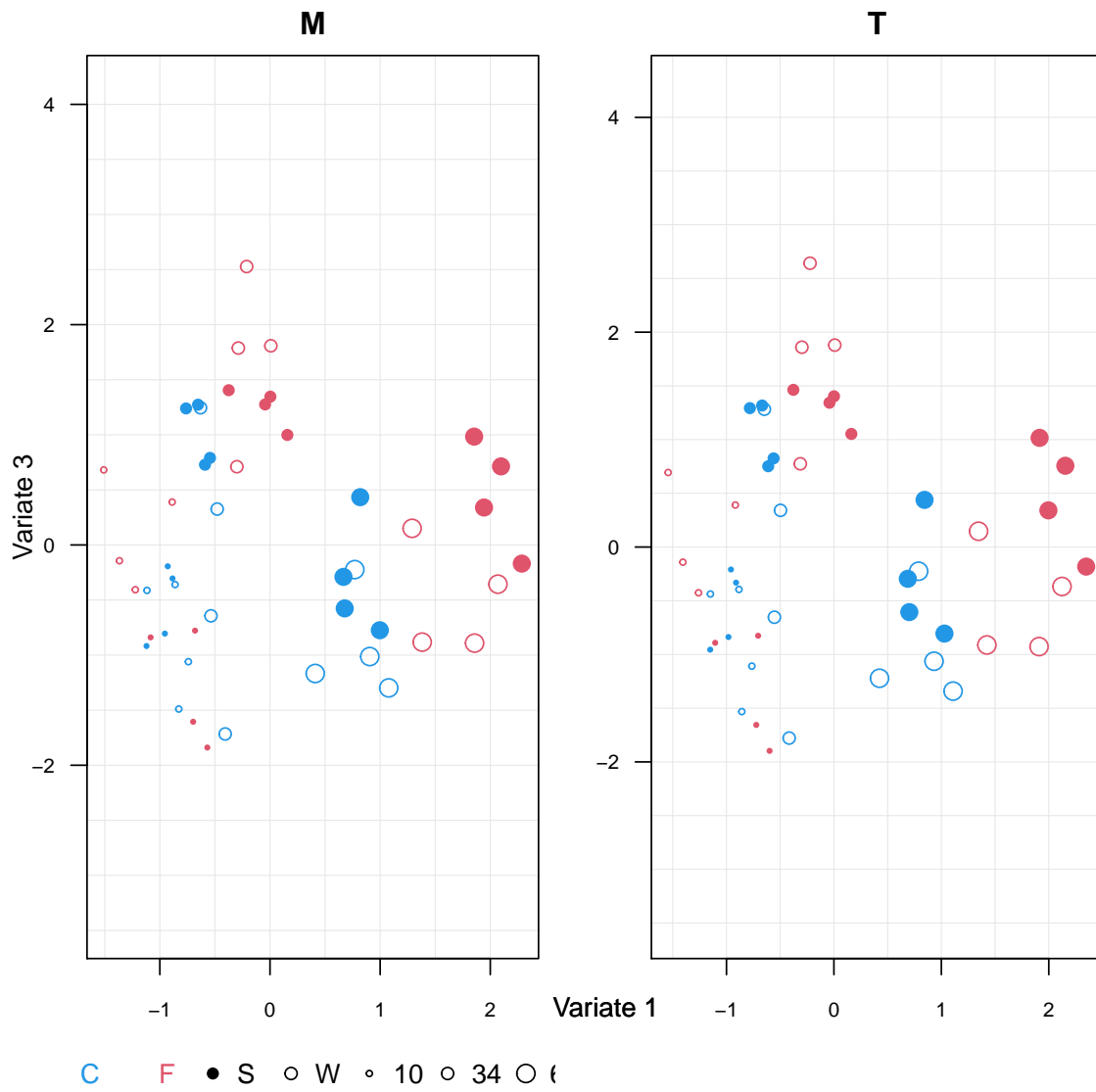


Figure 6: Samples plot for Components 1 and 3.

```
> my.plotIndiv(mt, comp=c(1,3)
+ )
```

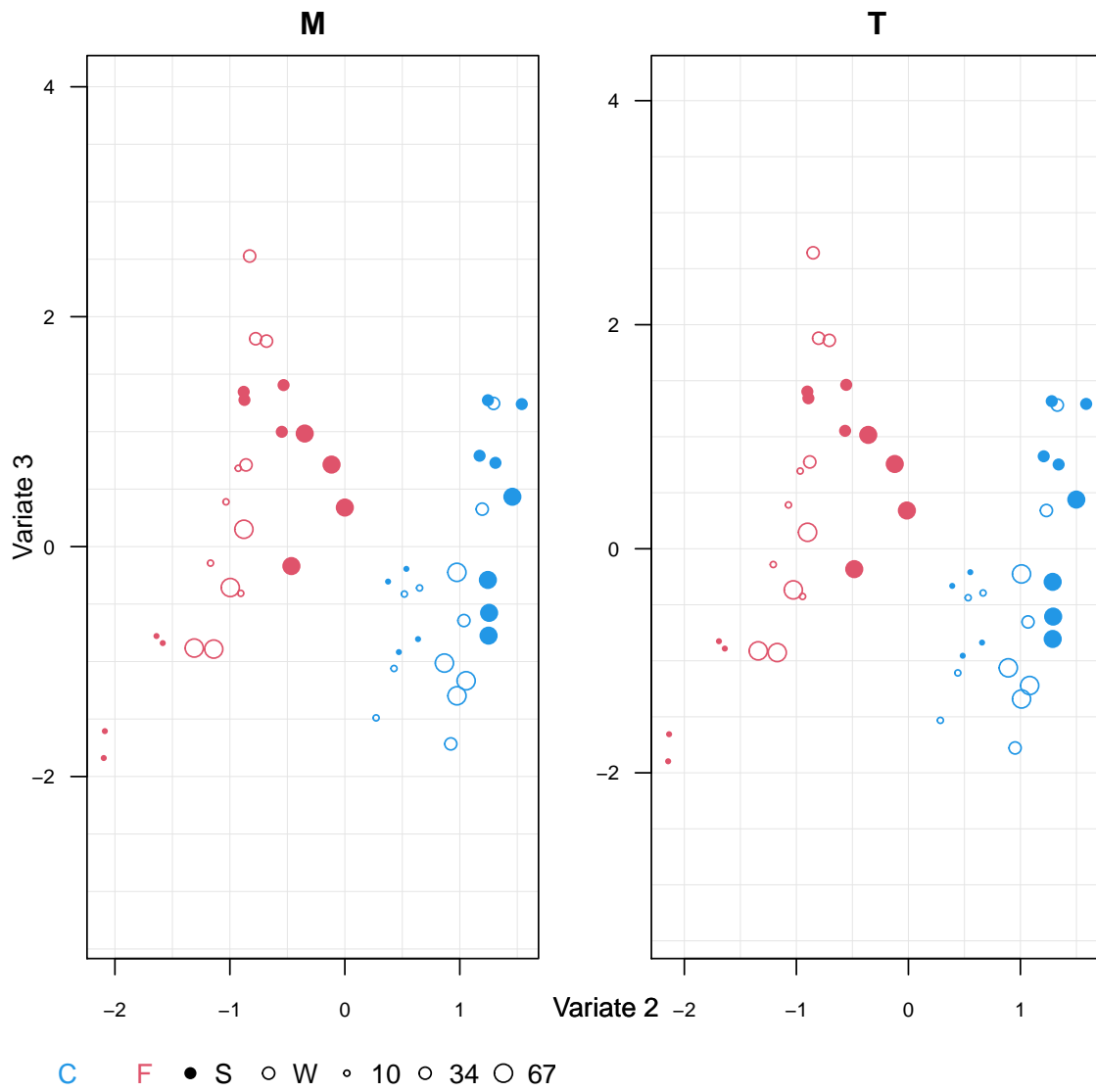
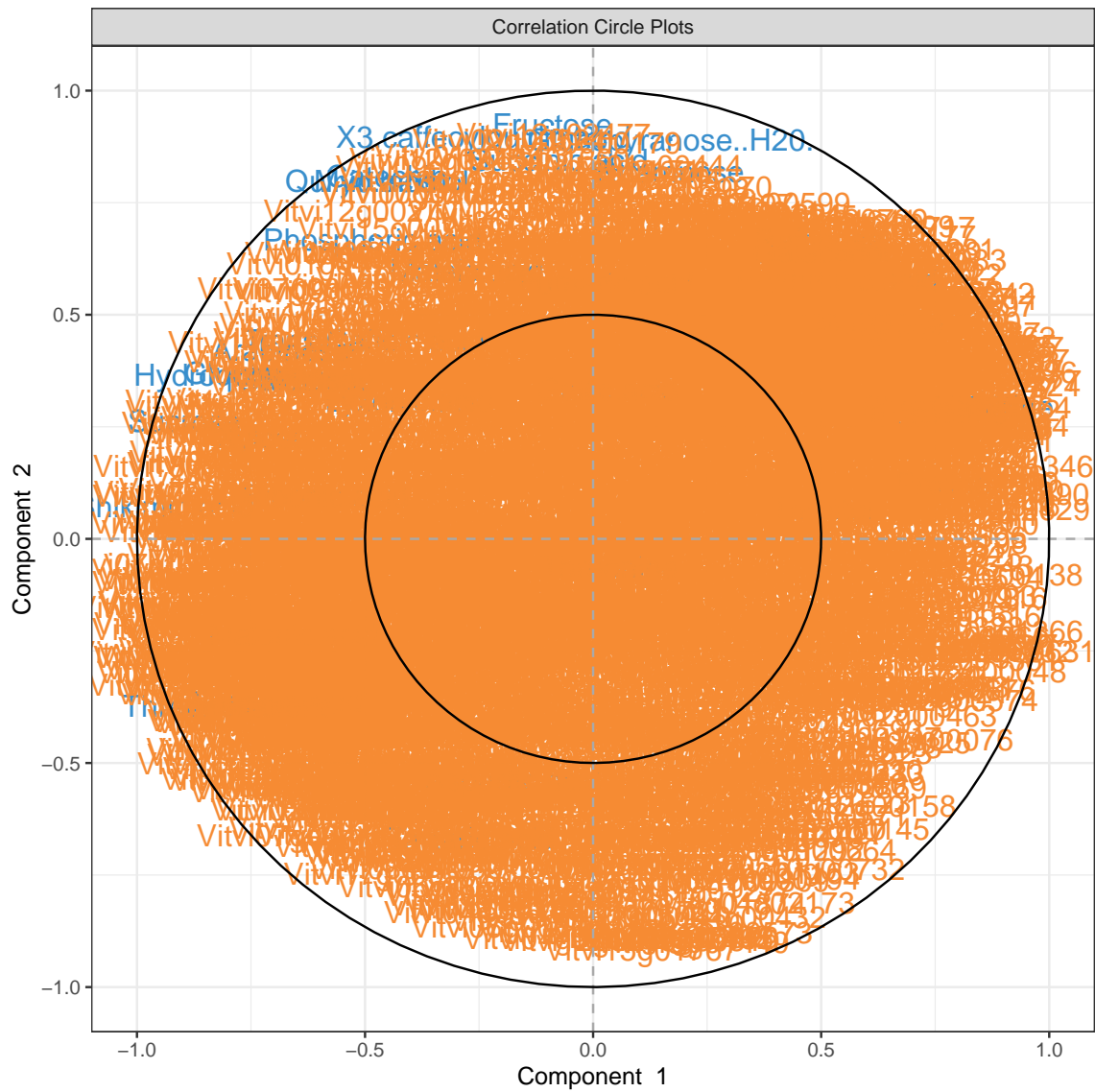


Figure 7: Samples plot for Components 2 and 3.

```
> my.plotIndiv(mt, comp=c(2,3)
+ )
```

```
> plotVar(mt)
```



```
> if(interactive()) graphics.off()
```

First three components, interactive 3d rgl image

```
> #plotVar(mt, style="3d")
```

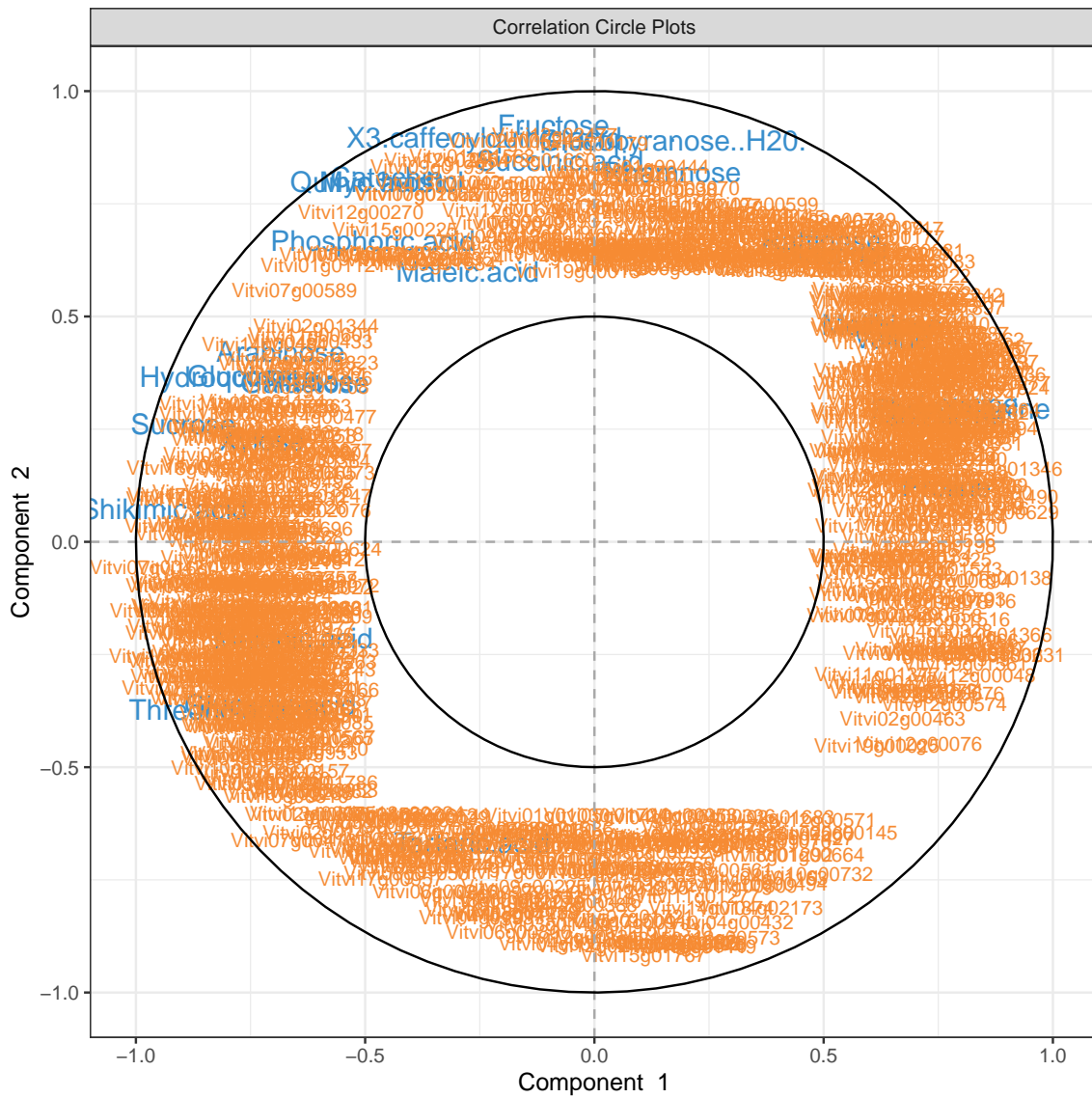
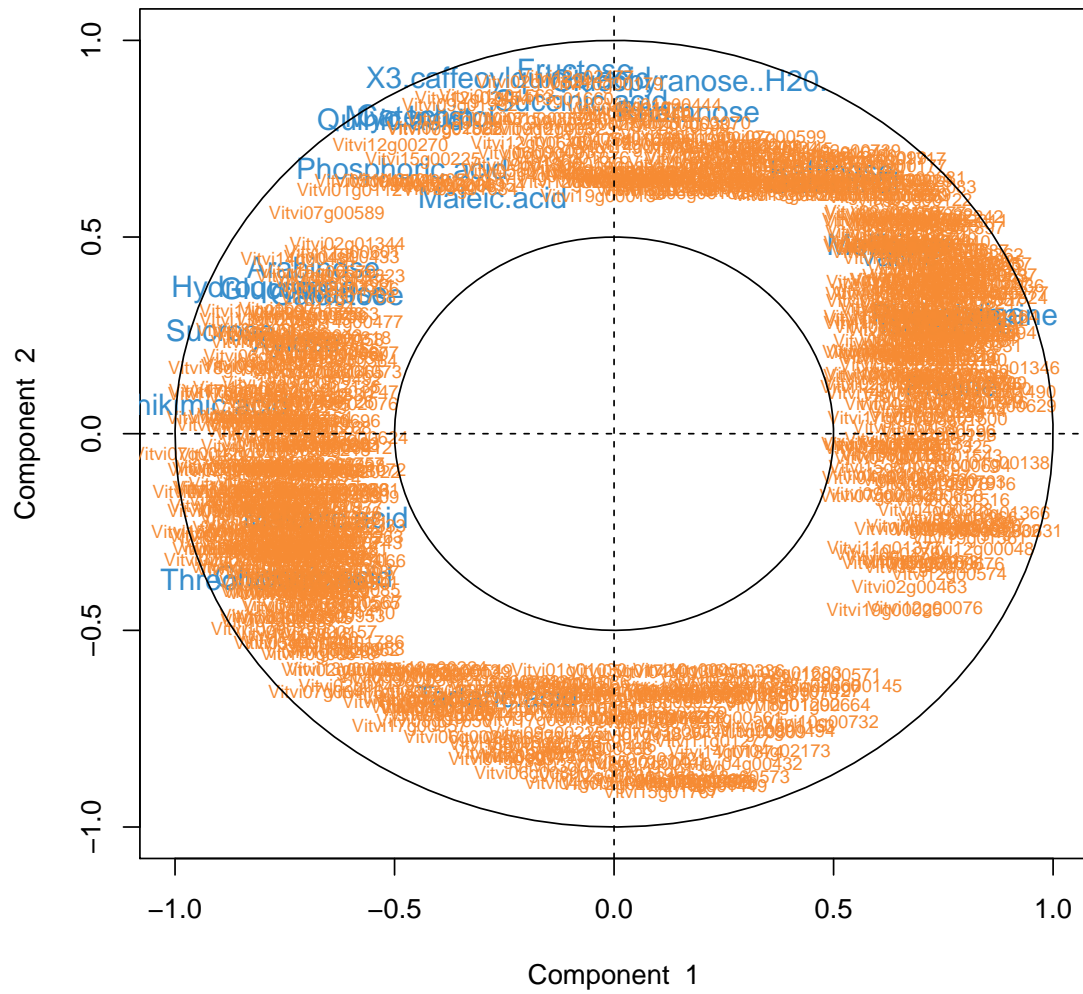



Figure 8: Correlation Circle plot; Cutoff: 0.6.

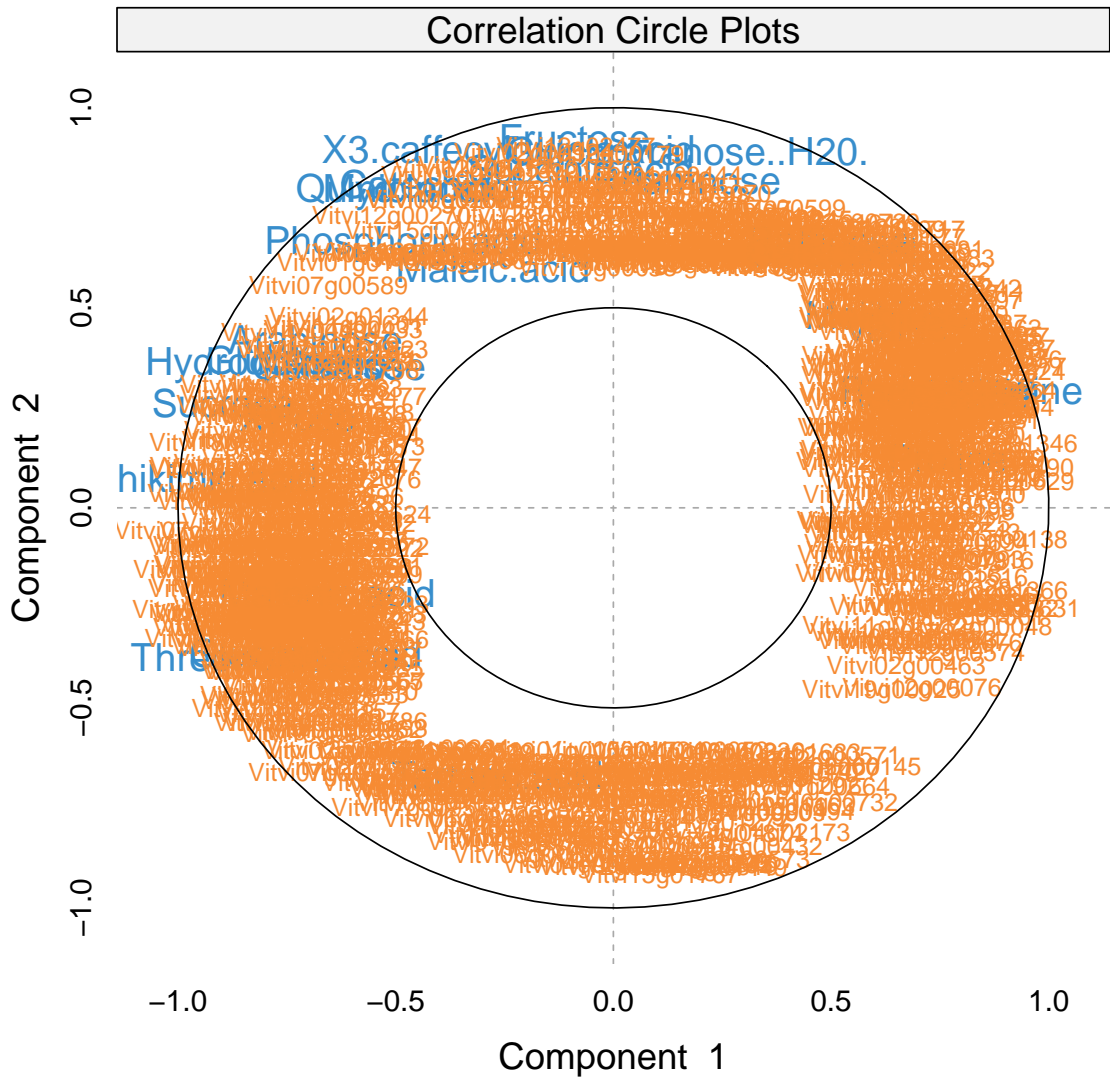
```
> plotVar(mt, cutoff=0.6, cex=c(1.5,1)*3, style="ggplot2")
```

```
> plotVar(mt, cutoff=0.6, cex=c(1.5,1)*0.7, style="graphics")
```

Correlation Circle Plots

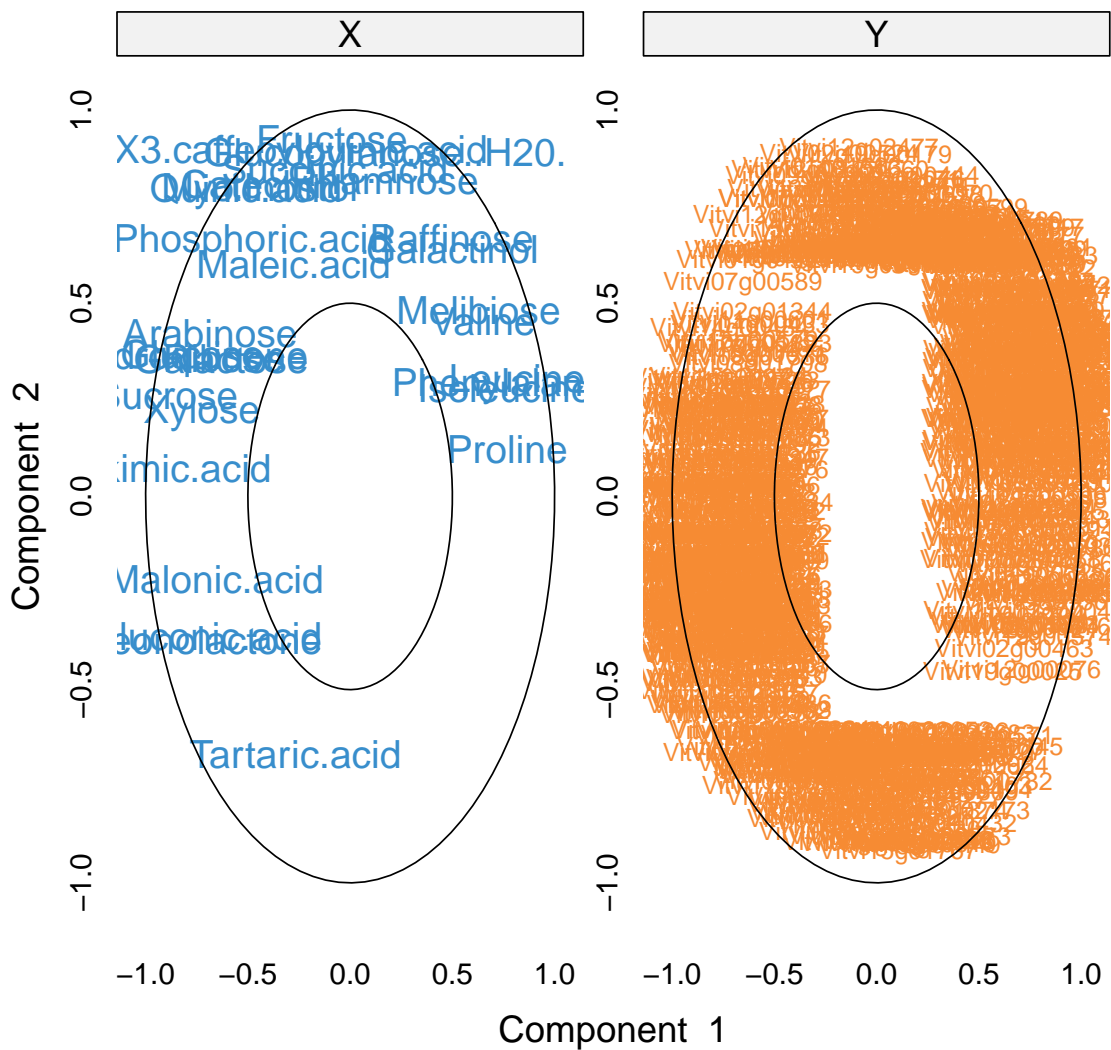


```
> plotVar(mt, cutoff=0.6, cex=c(1.5,1)*0.7, style="lattice")
```



```
> plotVar(mt, cutoff=0.6, cex=c(1.5,1)*0.7, style="lattice", overlap=FALSE)
```

Correlation Circle Plots



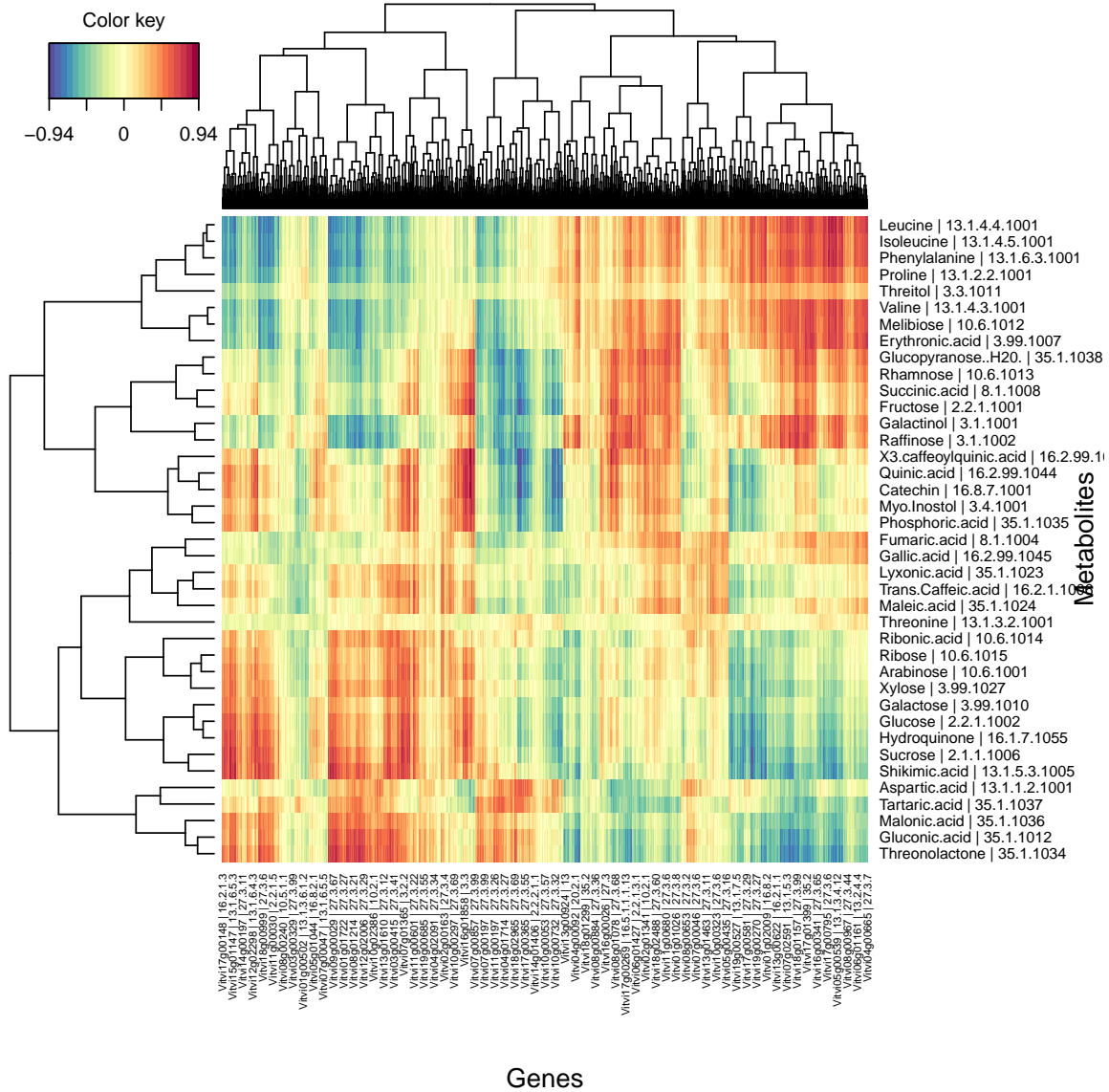


Figure 9: Heat map of genes and metabolites.

Additional plots

```
> cim(mt, margins=c(10,10), xlab="Genes", ylab="Metabolites"
+ )
```

Select rectangles to zoom

```
> if(interactive()) cim(mt, margins=c(6,8), xlab="Genes", ylab="Metabolites",
```

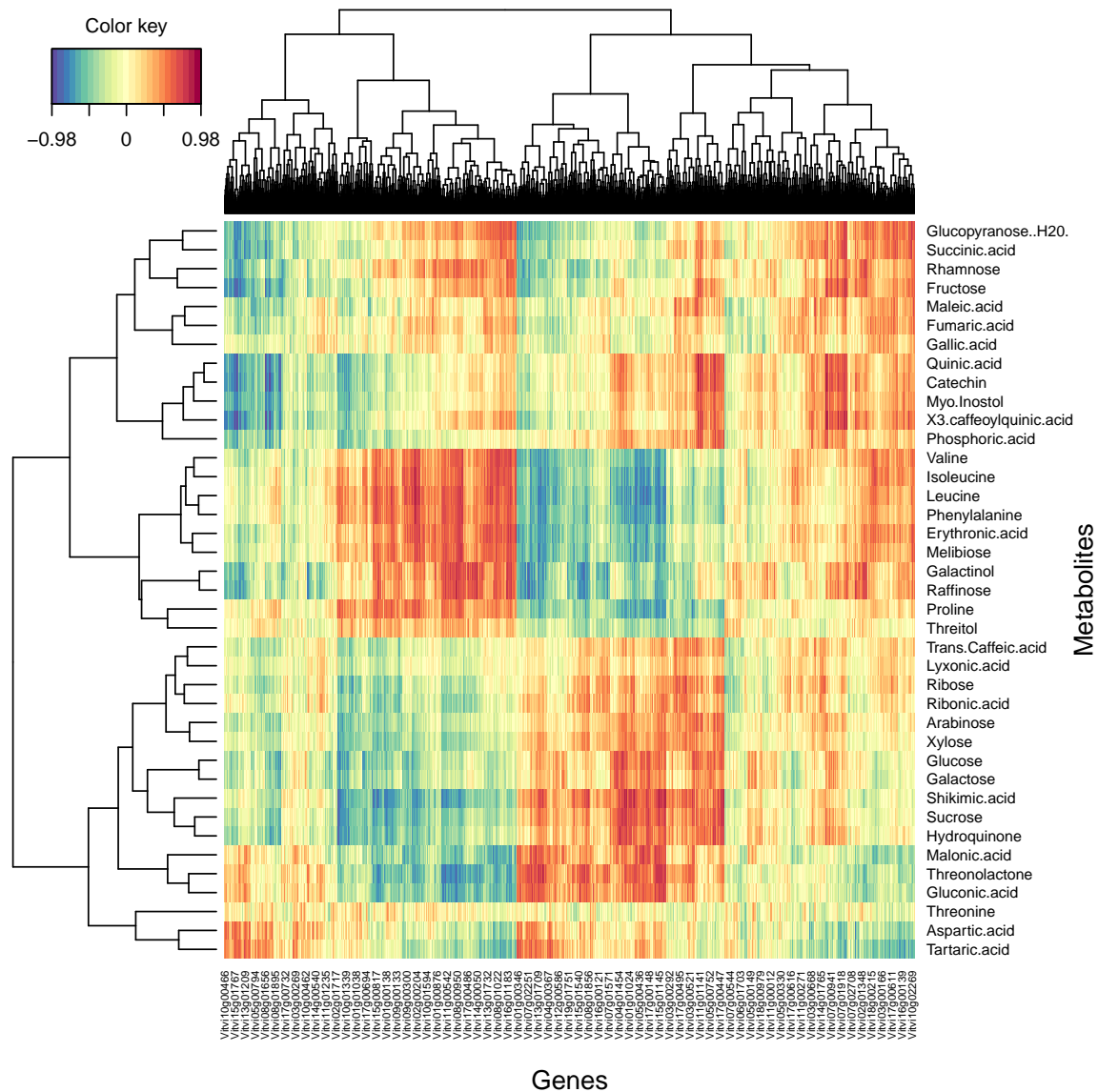


Figure 10: Heat map of correlations between genes and metabolites.

```
> cim(cor(M,T), margins=c(6,8), xlab="Genes", ylab="Metabolites")
```

6 Relevance network

Customized network function

```
> cutoff <- 0.8
```

6.0.1 Network with high cutoff: 0.8

```
> cutoff
```

```
[1] 0.8
```

```
> nettip <- "pdf"
```

```
> suffix <- paste(addArgs, collapse="-")
```

```
> (netfn <- gsub("\\.", "_", paste0("network-", cutoff, "_", suffix)))
```

```
[1] "network-0_8_18-1-1-2-3-4-5-6-7"
```

```
> pth <- .oroot
```

```
> (netpath <- file.path(pth, paste(netfn, nettip, sep=".")))
```

```
[1] "../..output/60_Trans-x-Meta-18-1-1-2-3-4-5-6-7/network-0_8_18-1-1-2-3-4-
```

```
> #jpeg(netpath)
```

```
> net <- my.network(mt, cutoff=cutoff, lwd=4
```

```
+   , shape.node=c("rectangle", "rectangle")
```

```
+   , color.node=c( 2, 4)
```

```
+   , show.edge.labels=TRUE
```

```
+ #   , interactive=interactive()
```

```
+   , name.save=file.path(pth, netfn)
```

```
+   , save=nettip
```

```
+   , main=paste0("Bins :", suffix, paste("\ncutoff =", cutoff))
```

```
+ )
```

```
Warning: package 'igraph' was built under R version 4.0.3
```

```
Attaching package: 'igraph'
```

```
The following objects are masked from 'package:lubridate':
```

```
  %--%, union
```

```
The following objects are masked from 'package:BiocGenerics':
```

```
  normalize, path, union
```

```
The following objects are masked from 'package:stats':
```

```
  decompose, spectrum
```

```
The following object is masked from 'package:base':
```

```
  union
```

```
> #dev.off()
```

> str(net)

List of 4

\$ gR :List of 10

..\$:List of 1

.. ..\$ Fructose | 2.2.1.1001: 'igraph.vs' Named int [1:27] 34 36 37 38 42 57

..- attr(*, "names")= chr [1:27] "Vitvi07g00376 | 10.2.1" "Vitvi10g001

..- attr(*, "env")=<weakref>

..- attr(*, "graph")= chr "aa1516c0-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Sucrose | 2.1.1.1006: 'igraph.vs' Named int [1:14] 25 26 39 40 65 77

..- attr(*, "names")= chr [1:14] "Vitvi18g00994 | 3.4.2" "Vitvi13g0183

..- attr(*, "env")=<weakref>

..- attr(*, "graph")= chr "aa1516c0-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Myo.Inostol | 3.4.1001: 'igraph.vs' Named int [1:3] 62 67 71

..- attr(*, "names")= chr [1:3] "Vitvi09g01992 | 16.1.5" "Vitvi12g0057

..- attr(*, "env")=<weakref>

..- attr(*, "graph")= chr "aa1516c0-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Raffinose | 3.1.1002: 'igraph.vs' Named int [1:7] 21 24 32 56 99 103

..- attr(*, "names")= chr [1:7] "Vitvi07g02242 | 3.1.1.1" "Vitvi14g017

..- attr(*, "env")=<weakref>

..- attr(*, "graph")= chr "aa1516c0-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Isoleucine | 13.1.4.5.1001: 'igraph.vs' Named int [1:4] 16 105 132 14

..- attr(*, "names")= chr [1:4] "Vitvi05g00357 | 2.2.2.1.2" "Vitvi07g0

..- attr(*, "env")=<weakref>

..- attr(*, "graph")= chr "aa1516c0-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Leucine | 13.1.4.4.1001: 'igraph.vs' Named int [1:34] 16 17 23 28 29

..- attr(*, "names")= chr [1:34] "Vitvi05g00357 | 2.2.2.1.2" "Vitvi07g

..- attr(*, "env")=<weakref>

..- attr(*, "graph")= chr "aa1516c0-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Phenylalanine | 13.1.6.3.1001: 'igraph.vs' Named int [1:12] 16 29 33

..- attr(*, "names")= chr [1:12] "Vitvi05g00357 | 2.2.2.1.2" "Vitvi03g

..- attr(*, "env")=<weakref>

..- attr(*, "graph")= chr "aa1516c0-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Shikimic.acid | 13.1.5.3.1005: 'igraph.vs' Named int [1:25] 25 26 28

..- attr(*, "names")= chr [1:25] "Vitvi18g00994 | 3.4.2" "Vitvi13g0183

..- attr(*, "env")=<weakref>

..- attr(*, "graph")= chr "aa1516c0-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Catechin | 16.8.7.1001: 'igraph.vs' Named int [1:37] 18 19 22 34 41 4

..- attr(*, "names")= chr [1:37] "Vitvi10g00494 | 2.2.2.2" "Vitvi16g01

..- attr(*, "env")=<weakref>

..- attr(*, "graph")= chr "aa1516c0-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Hydroquinone | 16.1.7.1055: 'igraph.vs' Named int [1:2] 118 124

..- attr(*, "names")= chr [1:2] "Vitvi11g00097 | 27.3.25" "Vitvi13g006

..- attr(*, "env")=<weakref>

..- attr(*, "graph")= chr "aa1516c0-80ce-11eb-b933-cf429f65e4a5"


```

..- attr(*, "class")= chr "igraph"
$ M      : num [1:39, 1:2049] 0 0 0 0 0 0 0 0 0 0 ...
..- attr(*, "dimnames")=List of 2
.. ..$ : chr [1:39] "Fructose" "Glucose" "Sucrose" "Erythronic.acid" ...
.. ..$ : chr [1:2049] "Vitvi01g00025" "Vitvi01g00052" "Vitvi01g00053" "Vitvi
$ cutoff: num 0.8
$ E      : 'igraph.es' int [1:262] 1 2 3 4 5 6 7 8 9 10 ...
..- attr(*, "vnames")= chr [1:262] "Fructose | 2.2.1.1001|Vitvi07g00376 | 10
..- attr(*, "env")=<weakref>
..- attr(*, "graph")= chr "aa1516c0-80ce-11eb-b933-cf429f65e4a5"

```

> *sapply(net\$gR, print)*

```

$`Fructose | 2.2.1.1001`
+ 27/147 vertices, named, from aa1516c:
 [1] Vitvi07g00376 | 10.2.1      Vitvi10g00179 | 10.6.2
 [3] Vitvi12g00131 | 10.6.3      Vitvi12g02477 | 10.6.3
 [5] Vitvi01g01563 | 13.2.2.3     Vitvi02g00524 | 16.5.1.1.1.10
 [7] Vitvi04g02192 | 16.4.1      Vitvi09g01992 | 16.1.5
 [9] Vitvi11g01330 | 16.4.1      Vitvi12g00573 | 16.1.5
[11] Vitvi12g02177 | 16.1.5      Vitvi12g02445 | 16.8.1.21
[13] Vitvi12g02451 | 16.8.1.21   Vitvi12g02454 | 16.8.1.21
[15] Vitvi13g01298 | 16.8.3.1     Vitvi13g01660 | 16.8.3.1
[17] Vitvi15g00510 | 16.5.1.1.1.11 Vitvi15g01504 | 16.8.3.1
[19] Vitvi15g01767 | 27.3.4      Vitvi04g00432 | 27.3.58
+ ... omitted several vertices

```

```

$`Sucrose | 2.1.1.1006`
+ 14/147 vertices, named, from aa1516c:
 [1] Vitvi18g00994 | 3.4.2      Vitvi13g01832 | 8.1.2
 [3] Vitvi12g02167 | 10.6.1     Vitvi17g00480 | 10.2.2
 [5] Vitvi10g01366 | 16.2      Vitvi17g00889 | 16.5.99.1
 [7] Vitvi07g00421 | 27.3.32   Vitvi09g00112 | 27.3.25
 [9] Vitvi11g00097 | 27.3.25   Vitvi12g00048 | 27.3.32
[11] Vitvi13g00088 | 27.3.25   Vitvi13g00631 | 27.3.57
[13] Vitvi18g00902 | 27.3.22   Vitvi19g01561 | 27.3.27

```

```

$`Myo.Inositol | 3.4.1001`
+ 3/147 vertices, named, from aa1516c:
 [1] Vitvi09g01992 | 16.1.5     Vitvi12g00573 | 16.1.5
 [3] Vitvi12g02454 | 16.8.1.21

```

```

$`Raffinose | 3.1.1002`
+ 7/147 vertices, named, from aa1516c:
 [1] Vitvi07g02242 | 3.1.1.1 Vitvi14g01717 | 3.1.2.2
 [3] Vitvi05g00681 | 10.2.1 Vitvi03g01897 | 16.7
 [5] Vitvi06g00304 | 27.3.11 Vitvi06g01552 | 27.3.18
 [7] Vitvi08g01050 | 27.3.11

```

```

$`Isoleucine | 13.1.4.5.1001`
+ 4/147 vertices, named, from aa1516c:
 [1] Vitvi05g00357 | 2.2.2.1.2 Vitvi07g00515 | 27.3.25
 [3] Vitvi15g00912 | 27.3.35 Vitvi18g00036 | 27.3.35

```

\$`Leucine | 13.1.4.4.1001`

+ 34/147 vertices, named, from aal516c:

[1]	Vitvi05g00357		2.2.2.1.2		Vitvi07g00353		2.2.1.5
[3]	Vitvi14g00063		3.1.1.1		Vitvi02g01230		10.8.2
[5]	Vitvi03g00537		10.5.3		Vitvi05g00675		10.2.1
[7]	Vitvi05g01353		10.8.1		Vitvi07g00525		10.6.3
[9]	Vitvi04g00870		13.1.1.1.1		Vitvi07g00101		13.1.3.4.12
[11]	Vitvi13g00597		13.1.6.1.7		Vitvi03g00564		16.2.1.10
[13]	Vitvi18g02476		16.8.3.1		Vitvi01g00918		27.3.35
[15]	Vitvi01g01796		27.3.99		Vitvi04g00845		27.3.25
[17]	Vitvi06g00360		27.3.3		Vitvi06g01480		27.3.35
[19]	Vitvi07g00421		27.3.32		Vitvi07g00515		27.3.25

+ ... omitted several vertices

\$`Phenylalanine | 13.1.6.3.1001`

+ 12/147 vertices, named, from aal516c:

[1]	Vitvi05g00357		2.2.2.1.2		Vitvi03g00537		10.5.3
[3]	Vitvi05g01353		10.8.1		Vitvi13g00597		13.1.6.1.7
[5]	Vitvi18g02476		16.8.3.1		Vitvi06g00360		27.3.3
[7]	Vitvi07g00421		27.3.32		Vitvi07g00515		27.3.25
[9]	Vitvi08g01164		27.3.63		Vitvi13g02031		27.3.50
[11]	Vitvi15g00912		27.3.35		Vitvi18g00036		27.3.35

\$`Shikimic.acid | 13.1.5.3.1005`

+ 25/147 vertices, named, from aal516c:

[1]	Vitvi18g00994		3.4.2		Vitvi13g01832		8.1.2
[3]	Vitvi02g01230		10.8.2		Vitvi04g00735		10.2.1
[5]	Vitvi07g00525		10.6.3		Vitvi12g02167		10.6.1
[7]	Vitvi17g00480		10.2.2		Vitvi04g01558		13.1.5.3.2
[9]	Vitvi15g01490		13.1.6.1.6		Vitvi10g01366		16.2
[11]	Vitvi17g00889		16.5.99.1		Vitvi01g00138		27.3.11
[13]	Vitvi02g00421		27.3.29		Vitvi03g00666		27.3.50
[15]	Vitvi04g00629		27.3.99		Vitvi06g00360		27.3.3
[17]	Vitvi07g00421		27.3.32		Vitvi07g00515		27.3.25
[19]	Vitvi09g00029		27.3.67		Vitvi10g01346		27.3.36

+ ... omitted several vertices

\$`Catechin | 16.8.7.1001`

+ 37/147 vertices, named, from aal516c:

[1]	Vitvi10g00494		2.2.2.2		Vitvi16g01405		2.1.2.2
[3]	Vitvi11g01277		3.3		Vitvi07g00376		10.2.1
[5]	Vitvi01g00664		13.1.6.5.2		Vitvi01g01563		13.2.2.3
[7]	Vitvi12g00158		13.1.1.2.1		Vitvi12g00270		13.1.6.4.3
[9]	Vitvi18g01292		13.1.7.11		Vitvi18g02060		13.1.3.5.2
[11]	Vitvi02g00524		16.5.1.1.1.10		Vitvi04g02192		16.4.1
[13]	Vitvi09g01992		16.1.5		Vitvi09g01413		16.1.5
[15]	Vitvi10g01862		16.1.5		Vitvi11g01330		16.4.1
[17]	Vitvi12g00573		16.1.5		Vitvi12g02445		16.8.1.21
[19]	Vitvi12g02451		16.8.1.21		Vitvi12g02454		16.8.1.21

+ ... omitted several vertices

\$`Hydroquinone | 16.1.7.1055`

```

+ 2/147 vertices, named, from aa1516c:
[1] Vitvi11g00097 | 27.3.25 Vitvi13g00631 | 27.3.57
$`Fructose | 2.2.1.1001`
+ 27/147 vertices, named, from aa1516c:
  [1] Vitvi07g00376 | 10.2.1          Vitvi10g00179 | 10.6.2
  [3] Vitvi12g00131 | 10.6.3          Vitvi12g02477 | 10.6.3
  [5] Vitvi01g01563 | 13.2.2.3         Vitvi02g00524 | 16.5.1.1.1.10
  [7] Vitvi04g02192 | 16.4.1          Vitvi09g01992 | 16.1.5
  [9] Vitvi11g01330 | 16.4.1          Vitvi12g00573 | 16.1.5
 [11] Vitvi12g02177 | 16.1.5          Vitvi12g02445 | 16.8.1.21
 [13] Vitvi12g02451 | 16.8.1.21       Vitvi12g02454 | 16.8.1.21
 [15] Vitvi13g01298 | 16.8.3.1       Vitvi13g01660 | 16.8.3.1
 [17] Vitvi15g00510 | 16.5.1.1.1.11 Vitvi15g01504 | 16.8.3.1
 [19] Vitvi15g01767 | 27.3.4          Vitvi04g00432 | 27.3.58
+ ... omitted several vertices

$`Sucrose | 2.1.1.1006`
+ 14/147 vertices, named, from aa1516c:
  [1] Vitvi18g00994 | 3.4.2          Vitvi13g01832 | 8.1.2
  [3] Vitvi12g02167 | 10.6.1       Vitvi17g00480 | 10.2.2
  [5] Vitvi10g01366 | 16.2          Vitvi17g00889 | 16.5.99.1
  [7] Vitvi07g00421 | 27.3.32      Vitvi09g00112 | 27.3.25
  [9] Vitvi11g00097 | 27.3.25      Vitvi12g00048 | 27.3.32
 [11] Vitvi13g00088 | 27.3.25      Vitvi13g00631 | 27.3.57
 [13] Vitvi18g00902 | 27.3.22      Vitvi19g01561 | 27.3.27

$`Myo.Inositol | 3.4.1001`
+ 3/147 vertices, named, from aa1516c:
[1] Vitvi09g01992 | 16.1.5       Vitvi12g00573 | 16.1.5
[3] Vitvi12g02454 | 16.8.1.21

$`Raffinose | 3.1.1002`
+ 7/147 vertices, named, from aa1516c:
[1] Vitvi07g02242 | 3.1.1.1 Vitvi14g01717 | 3.1.2.2
[3] Vitvi05g00681 | 10.2.1 Vitvi03g01897 | 16.7
[5] Vitvi06g00304 | 27.3.11 Vitvi06g01552 | 27.3.18
[7] Vitvi08g01050 | 27.3.11

$`Isoleucine | 13.1.4.5.1001`
+ 4/147 vertices, named, from aa1516c:
[1] Vitvi05g00357 | 2.2.2.1.2 Vitvi07g00515 | 27.3.25
[3] Vitvi15g00912 | 27.3.35 Vitvi18g00036 | 27.3.35

$`Leucine | 13.1.4.4.1001`
+ 34/147 vertices, named, from aa1516c:
  [1] Vitvi05g00357 | 2.2.2.1.2 Vitvi07g00353 | 2.2.1.5
  [3] Vitvi14g00063 | 3.1.1.1 Vitvi02g01230 | 10.8.2
  [5] Vitvi03g00537 | 10.5.3 Vitvi05g00675 | 10.2.1
  [7] Vitvi05g01353 | 10.8.1 Vitvi07g00525 | 10.6.3
  [9] Vitvi04g00870 | 13.1.1.1.1 Vitvi07g00101 | 13.1.3.4.12
 [11] Vitvi13g00597 | 13.1.6.1.7 Vitvi03g00564 | 16.2.1.10
 [13] Vitvi18g02476 | 16.8.3.1 Vitvi01g00918 | 27.3.35
 [15] Vitvi01g01796 | 27.3.99 Vitvi04g00845 | 27.3.25

```

```
[17] Vitvi06g00360 | 27.3.3      Vitvi06g01480 | 27.3.35
[19] Vitvi07g00421 | 27.3.32      Vitvi07g00515 | 27.3.25
+ ... omitted several vertices
```

```
$`Phenylalanine | 13.1.6.3.1001`
```

```
+ 12/147 vertices, named, from aal516c:
```

```
[1] Vitvi05g00357 | 2.2.2.1.2  Vitvi03g00537 | 10.5.3
[3] Vitvi05g01353 | 10.8.1      Vitvi13g00597 | 13.1.6.1.7
[5] Vitvi18g02476 | 16.8.3.1    Vitvi06g00360 | 27.3.3
[7] Vitvi07g00421 | 27.3.32     Vitvi07g00515 | 27.3.25
[9] Vitvi08g01164 | 27.3.63     Vitvi13g02031 | 27.3.50
[11] Vitvi15g00912 | 27.3.35     Vitvi18g00036 | 27.3.35
```

```
$`Shikimic.acid | 13.1.5.3.1005`
```

```
+ 25/147 vertices, named, from aal516c:
```

```
[1] Vitvi18g00994 | 3.4.2      Vitvi13g01832 | 8.1.2
[3] Vitvi02g01230 | 10.8.2     Vitvi04g00735 | 10.2.1
[5] Vitvi07g00525 | 10.6.3     Vitvi12g02167 | 10.6.1
[7] Vitvi17g00480 | 10.2.2     Vitvi04g01558 | 13.1.5.3.2
[9] Vitvi15g01490 | 13.1.6.1.6 Vitvi10g01366 | 16.2
[11] Vitvi17g00889 | 16.5.99.1  Vitvi01g00138 | 27.3.11
[13] Vitvi02g00421 | 27.3.29     Vitvi03g00666 | 27.3.50
[15] Vitvi04g00629 | 27.3.99     Vitvi06g00360 | 27.3.3
[17] Vitvi07g00421 | 27.3.32     Vitvi07g00515 | 27.3.25
[19] Vitvi09g00029 | 27.3.67     Vitvi10g01346 | 27.3.36
```

```
+ ... omitted several vertices
```

```
$`Catechin | 16.8.7.1001`
```

```
+ 37/147 vertices, named, from aal516c:
```

```
[1] Vitvi10g00494 | 2.2.2.2      Vitvi16g01405 | 2.1.2.2
[3] Vitvi11g01277 | 3.3          Vitvi07g00376 | 10.2.1
[5] Vitvi01g00664 | 13.1.6.5.2  Vitvi01g01563 | 13.2.2.3
[7] Vitvi12g00158 | 13.1.1.2.1  Vitvi12g00270 | 13.1.6.4.3
[9] Vitvi18g01292 | 13.1.7.11    Vitvi18g02060 | 13.1.3.5.2
[11] Vitvi02g00524 | 16.5.1.1.1.10 Vitvi04g02192 | 16.4.1
[13] Vitvi09g01992 | 16.1.5      Vitvi09g01413 | 16.1.5
[15] Vitvi10g01862 | 16.1.5      Vitvi11g01330 | 16.4.1
[17] Vitvi12g00573 | 16.1.5      Vitvi12g02445 | 16.8.1.21
[19] Vitvi12g02451 | 16.8.1.21  Vitvi12g02454 | 16.8.1.21
```

```
+ ... omitted several vertices
```

```
$`Hydroquinone | 16.1.7.1055`
```

```
+ 2/147 vertices, named, from aal516c:
```

```
[1] Vitvi11g00097 | 27.3.25  Vitvi13g00631 | 27.3.57
```

```
> library(igraph)
```

```
> nettip <- "gml"
```

```
> (netfn<- gsub("\\.", "_", paste0("network-", cutoff, "_", suffix)))
```

```
[1] "network-0_8_18-1-1-2-3-4-5-6-7"
```

```
> (netpath <- file.path(.oroot, paste(netfn, nettip, sep=".")))
```

```
[1] ".././output/60_Trans-x-Meta-18-1-1-2-3-4-5-6-7/network-0_8_18-1-1-2-3-4-
```

```
> write.graph(net$gR, file=netpath, format=nettip)
```

```

> #jpeg(netpath)
> net <- my.network(mt.shrink, cutoff=cutoff, lwd=4
+   , shape.node=c("rectangle", "rectangle")
+   , color.node=c( 2, 4)
+   , show.edge.labels=TRUE
+ #   , interactive=interactive()
+   , name.save=file.path(pth, netfn)
+   , save=nettip
+   , main=paste0("Bins :", suffix, paste("\ncutoff =", cutoff))
+ )
Error: 'save' must be one of 'jpeg', 'png', 'tiff' or 'pdf'.
> #dev.off()
> #sapply(net$gR, print)

```

Extract edges from the graph and prepare the data.frame with names, bin codes, descriptions ...

```

> edg <- matrix(unlist(sapply(attr(E(net$gR), "vnames"), strsplit, split="\\|")),
> colnames(edg)=c("Metabolite", "geneID")
> medg <- mldata[edg[,1], 1:3 ]
> tedg <- fdata[match(edg[,2], fdata$geneID), -1]
> edges <- data.frame(edg, weight=E(net$gR)$weight, medg, tedg )
> str(edges)
'data.frame':      524 obs. of  10 variables:
 $ Metabolite : chr  "Fructose " "Vitvi07g00376 " "Fructose " "Vitvi10g00179
 $ geneID      : chr  " 2.2.1.1001" " 10.2.1" " 2.2.1.1001" " 10.6.2" ...
 $ weight      : num  -0.863 0.841 -0.815 0.847 0.845 ...
 $ Metabolite.1: chr  NA NA NA NA ...
 $ Bin         : chr  NA NA NA NA ...
 $ Description : chr  NA NA NA NA ...
 $ geneID.1    : chr  NA NA NA NA ...
 $ BINCODE     : chr  NA NA NA NA ...
 $ NAME        : chr  NA NA NA NA ...
 $ DESCRIPTION : chr  NA NA NA NA ...
> dim(edges)
[1] 524 10

```

Export edges file

```

> cutoff
[1] 0.8
> nettip <- "txt"
> suffix <- paste(addArgs, collapse="-")
> pth <- .oroot
> (netfn <- gsub("\\.", "_", paste0("edges_table-", cutoff, "_", suffix)))
[1] "edges_table-0_8_18-1-1-2-3-4-5-6-7"
> (netpath <- file.path(.oroot, paste(netfn, nettip, sep=". ")))
[1] "../..output/60_Trans-x-Meta-18-1-1-2-3-4-5-6-7/edges_table-0_8_18-1-1-2-
> my.write.table(edges, file=netpath,
+   label=paste("Edges table, cutoff =", cutoff)
+   )
Warning in write.table(x, file = file, col.names = col.names, sep = sep, : app
Object: edges \\
Label: Edges table, cutoff = 0.8 \\
File :\\
\\href{run:D:\\DEJAVNOSTI\\OMIKE\\pISA-projects\\_p_VinskaTrta\\_I_EnViRoS\\_S_
> cutoff <- 0.5

```

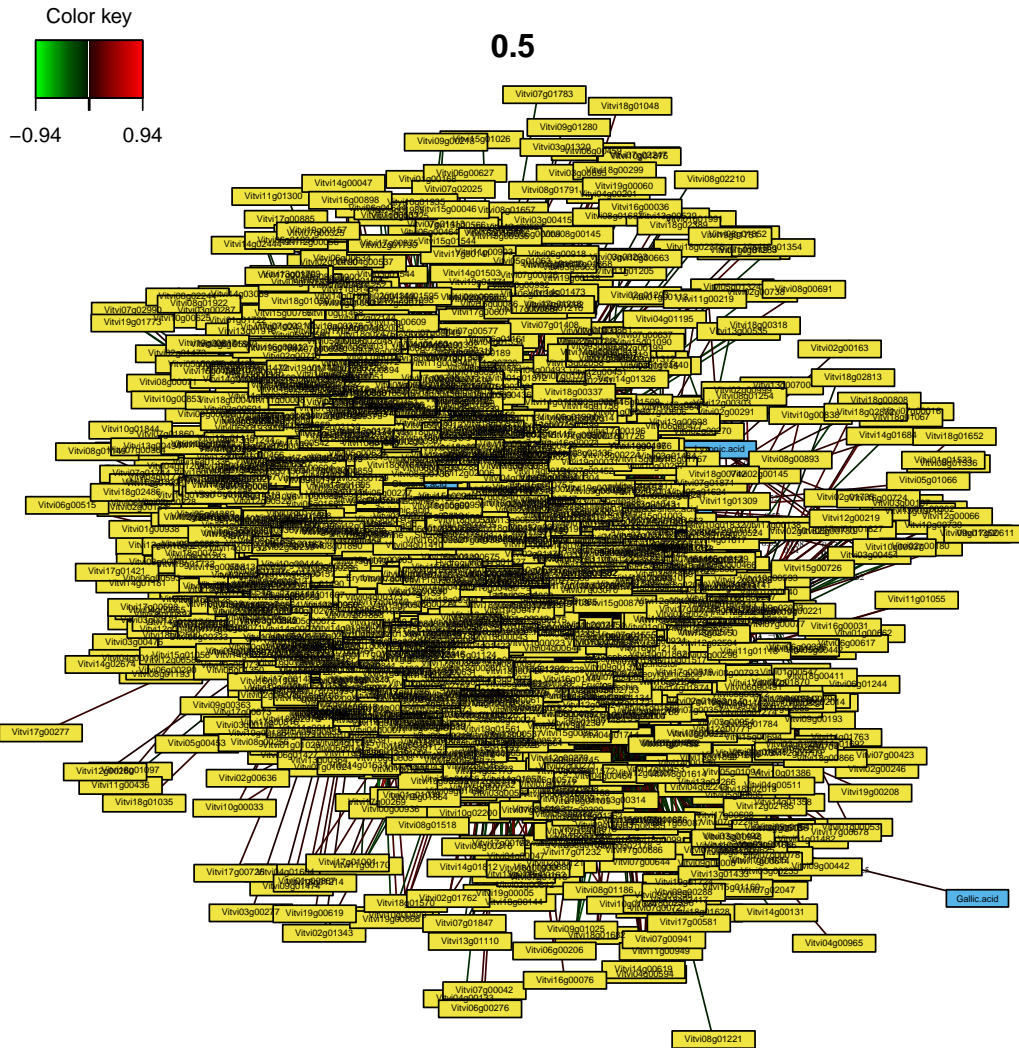
6.0.2 Network with low cutoff: 0.5

```
> cutoff
[1] 0.5
> nettip <- "pdf"
> sufix <- paste(addArgs, collapse="-")
> (netfn<- gsub("\\.", "_", paste0("network-", cutoff, "_", sufix)))
[1] "network-0_5_18-1-1-2-3-4-5-6-7"
> pth <- .oroot
> (netpath<- file.path(pth, paste(netfn, nettip, sep=".")))
[1] ".././output/60_Trans-x-Meta-18-1-1-2-3-4-5-6-7/network-0_5_18-1-1-2-3-4-5-6-7.pdf"

> #
> net <- my.network(mt, cutoff=cutoff, lwd=4,
+   shape.node=c("rectangle", "rectangle"),
+   color.node=c( 2, 4)
+   , name.save=file.path(pth, netfn)
+   , save=nettip
+   , main=paste0("Bins :", sufix, paste("\ncutoff =", cutoff))
+   )
```

To prepare the edges table, we will use cutoff 0.5. For visual inspection, filter on column weight can be used.

```
> #
> cutoff
[1] 0.5
> net <-my.network(mt.shrink, cutoff=cutoff,
+   shape.node=c("rectangle", "rectangle"),
+   color.node=c( 2, 4),
+   show.edge.labels=TRUE
+   )
```



> str(net)

List of 4

\$ gR :List of 10

..\$:List of 1

.. ..\$ Fructose: 'igraph.vs' Named int [1:365] 43 46 48 50 58 60 61 62 63 64
- attr(*, "names")= chr [1:365] "Vitvi02g00250" "Vitvi03g00088" "Vitvi03g00089"
- attr(*, "env")=<weakref>
- attr(*, "graph")= chr "ac77ef8c-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Glucose: 'igraph.vs' Named int [1:352] 40 46 49 52 54 57 64 74 80 82
- attr(*, "names")= chr [1:352] "Vitvi01g00064" "Vitvi03g00088" "Vitvi03g00089"
- attr(*, "env")=<weakref>
- attr(*, "graph")= chr "ac77ef8c-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Sucrose: 'igraph.vs' Named int [1:535] 40 41 46 47 49 52 53 54 56 57
- attr(*, "names")= chr [1:535] "Vitvi01g00064" "Vitvi01g00932" "Vitvi01g00933"
- attr(*, "env")=<weakref>
- attr(*, "graph")= chr "ac77ef8c-80ce-11eb-b933-cf429f65e4a5"

..\$:List of 1

.. ..\$ Erythronic.acid: 'igraph.vs' Named int [1:555] 38 40 42 44 47 48 49 50
- attr(*, "names")= chr [1:555] "Vitvi01g00052" "Vitvi01g00064" "Vitvi01g00065"
- attr(*, "env")=<weakref>

```

.. .. ..- attr(*, "graph")= chr "ac77ef8c-80ce-11eb-b933-cf429f65e4a5"
..$ :List of 1
.. ..$ Galactinol: 'igraph.vs' Named int [1:529] 45 47 48 49 55 56 59 60 69
.. .. ..- attr(*, "names")= chr [1:529] "Vitvi02g00605" "Vitvi03g00304" "Vit
.. .. ..- attr(*, "env")=<weakref>
.. .. ..- attr(*, "graph")= chr "ac77ef8c-80ce-11eb-b933-cf429f65e4a5"
..$ :List of 1
.. ..$ Galactose: 'igraph.vs' Named int [1:149] 40 57 64 80 82 83 84 97 105
.. .. ..- attr(*, "names")= chr [1:149] "Vitvi01g00064" "Vitvi07g01830" "Vit
.. .. ..- attr(*, "env")=<weakref>
.. .. ..- attr(*, "graph")= chr "ac77ef8c-80ce-11eb-b933-cf429f65e4a5"
..$ :List of 1
.. ..$ Myo.Inostol: 'igraph.vs' Named int [1:272] 43 46 50 62 63 64 83 84 94
.. .. ..- attr(*, "names")= chr [1:272] "Vitvi02g00250" "Vitvi03g00088" "Vit
.. .. ..- attr(*, "env")=<weakref>
.. .. ..- attr(*, "graph")= chr "ac77ef8c-80ce-11eb-b933-cf429f65e4a5"
..$ :List of 1
.. ..$ Raffinose: 'igraph.vs' Named int [1:584] 45 47 48 49 51 55 56 60 69 7
.. .. ..- attr(*, "names")= chr [1:584] "Vitvi02g00605" "Vitvi03g00304" "Vit
.. .. ..- attr(*, "env")=<weakref>
.. .. ..- attr(*, "graph")= chr "ac77ef8c-80ce-11eb-b933-cf429f65e4a5"
..$ :List of 1
.. ..$ Xylose: 'igraph.vs' Named int [1:228] 54 76 88 90 92 93 97 98 102 105
.. .. ..- attr(*, "names")= chr [1:228] "Vitvi06g01272" "Vitvi14g00950" "Vit
.. .. ..- attr(*, "env")=<weakref>
.. .. ..- attr(*, "graph")= chr "ac77ef8c-80ce-11eb-b933-cf429f65e4a5"
..$ :List of 1
.. ..$ Fumaric.acid: 'igraph.vs' Named int [1:30] 61 146 216 286 300 326 406
.. .. ..- attr(*, "names")= chr [1:30] "Vitvi08g01353" "Vitvi18g02757" "Vitv
.. .. ..- attr(*, "env")=<weakref>
.. .. ..- attr(*, "graph")= chr "ac77ef8c-80ce-11eb-b933-cf429f65e4a5"
..- attr(*, "class")= chr "igraph"
$ M      : num [1:39, 1:2049] 0 0 0 0 0 0 0 0 0 0 0 ...
..- attr(*, "dimnames")=List of 2
.. ..$ : chr [1:39] "Fructose" "Glucose" "Sucrose" "Erythronic.acid" ...
.. ..$ : chr [1:2049] "Vitvi01g00025" "Vitvi01g00052" "Vitvi01g00053" "Vitvi
$ cutoff: num 0.5
$ E      : 'igraph.es' int [1:12972] 1 2 3 4 5 6 7 8 9 10 ...
..- attr(*, "vnames")= chr [1:12972] "Fructose|Vitvi02g00250" "Fructose|Vitv
..- attr(*, "env")=<weakref>
..- attr(*, "graph")= chr "ac77ef8c-80ce-11eb-b933-cf429f65e4a5"

```

Extract edges from the graph and prepare the data.frame with names, bin codes, descriptions ...

```

> edg <- matrix(unlist(sapply(attr(E(net$gR), "vnames"), strsplit, split="\\|")),
> colnames(edg)=c("Metabolite", "geneID")
> medg <- mldata[edg[,1], 1:3 ]
> tedg <- fdata[match(edg[,2], fdata$geneID), -1]
> edges <- data.frame(edg, weight=E(net$gR)$weight, medg, tedg )
> str(edges)

'data.frame':      12972 obs. of  10 variables:
 $ Metabolite : chr "Fructose" "Fructose" "Fructose" "Fructose" ...

```



```

$ geneID      : chr  "Vitvi02g00250" "Vitvi03g00088" "Vitvi05g00164" "Vitvi05g00164" "Vitvi05g00164" ...
$ weight      : num  0.592 0.592 -0.516 0.521 0.515 ...
$ Metabolite.1: chr  "Fructose" "Fructose" "Fructose" "Fructose" ...
$ Bin         : chr  "2.2.1.1001" "2.2.1.1001" "2.2.1.1001" "2.2.1.1001" ...
$ Description : chr  "major CHO metabolism.degradation.sucrose.fructose" "major CHO metabolism.degradation.sucrose.fructose" ...
$ geneID.1    : chr  "Vitvi02g00250" "Vitvi03g00088" "Vitvi05g00164" "Vitvi05g00164" ...
$ BINCODE     : chr  "2.1.2.2" "2.2.1.3.1" "2.2.1.3.1" "2.1.2.1" ...
$ NAME        : chr  "major CHO metabolism.synthesis.starch.starch synthase" "major CHO metabolism.synthesis.starch.starch synthase" ...
$ DESCRIPTION : chr  "UDP-Glycosyltransferase superfamily protein | Chr1:1192" "UDP-Glycosyltransferase superfamily protein | Chr1:1192" ...

```

```
> dim(edges)
```

```
[1] 12972    10
```

Export edges file

```
> nettip <- "txt"
```

```
> suffix <- paste(addArgs, collapse="-")
```

```
> pth <- .oroot
```

```
> (netfn <- gsub("\\.", "_", paste0("edges_table-", cutoff, "_", suffix)))
```

```
[1] "edges_table-0_5_18-1-1-2-3-4-5-6-7"
```

```
> (netpath <- file.path(.oroot, paste(netfn, nettip, sep=". ")))
```

```
[1] "../..output/60_Trans-x-Meta-18-1-1-2-3-4-5-6-7/edges_table-0_5_18-1-1-2-3-4-5-6-7.txt"
```

```
> my.write.table(edges, file=netpath,
```

```
+   label=paste("Edges table, cutoff =", cutoff)
```

```
+   )
```

Warning in write.table(x, file = file, col.names = col.names, sep = sep, : append as text

```
Object: edges \\
```

```
Label: Edges table, cutoff = 0.5 \\
```

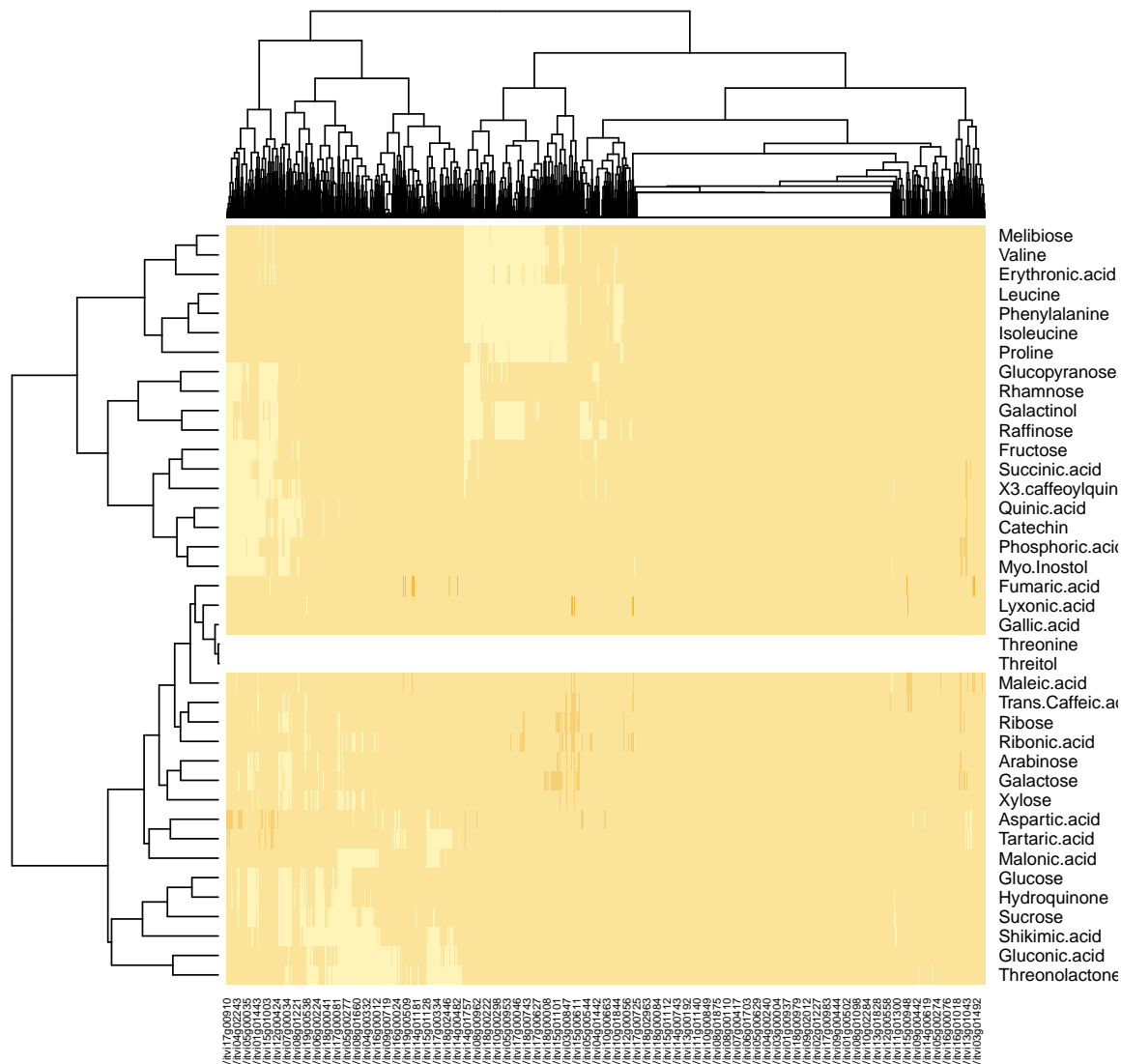
```
File :\\
```

```
\\href{run:D:\\DEJAVNOSTI\\OMIKE\\pISA-projects\\_p_VinskaTrta\\_I_EnViRoS\\_S_
```

```

> names(net)
[1] "gR"      "M"      "cutoff" "E"
> heatmap(net$M)

```



```

> rw <- apply(net$M, 1, function(x, eps=0.1) any(abs(x) >= eps))
> cl <- apply(net$M, 2, function(x, eps=0.1) any(abs(x) >= eps))
> x <- net$M
> x <- x[rw, cl]
> print.blank(t(x))

```

	Fructose	Glucose	Sucrose	Erythronic.acid
Vitvi01g00052	.	.	.	-0.5104274
Vitvi01g00053
Vitvi01g00064	.	0.6471539	0.7447086	-0.5878117
Vitvi01g00932	.	.	0.5359679	.
Vitvi02g00184	.	.	.	0.6014859
Vitvi02g00250	0.5917430	.	.	.
Vitvi02g00512	.	.	.	-0.5032023
Vitvi02g00605
Vitvi03g00088	0.5915617	0.5383286	0.5217524	.

Vitvi03g00304	.	.	0.5757660	-0.5824368	.
Vitvi05g00164	-0.5162027	.	.	-0.6792907	.
Vitvi05g00357	.	-0.5247861	-0.6544628	0.7365837	.
Vitvi05g00442	0.5212446
Vitvi05g01324
Vitvi05g01516	.	0.5641255	0.6671287	-0.6447365	.
Vitvi06g00583	.	.	0.5474486	-0.6569086	.
Vitvi06g01272	.	0.5817600	0.6787532	.	.
Vitvi06g01427
Vitvi07g00353	.	.	-0.6076169	0.7097887	.
Vitvi07g01830	.	0.5967170	0.6641751	.	.
Vitvi08g00225	0.5153951
Vitvi08g00930	.	.	0.5636198	-0.6477947	.
Vitvi08g01307	0.5243501	.	.	0.5844520	.
Vitvi08g01353	0.5527754	.	.	0.6558454	.
Vitvi09g00193	-0.5008085
Vitvi10g00094	0.5769528
Vitvi10g00494	-0.7548289	-0.5875827	-0.5489687	.	.
Vitvi10g00499	.	.	.	0.5591197	.
Vitvi10g00739
Vitvi11g00030	.	.	.	-0.6861736	.
Vitvi11g00260	.	.	.	-0.5916697	.
Vitvi11g00542	.	.	-0.5705351	0.5480646	.
Vitvi11g00903
Vitvi12g00770	0.6090534	.	.	0.5783938	.
Vitvi12g01618	.	.	0.5418723	.	.
Vitvi13g00792	.	.	.	0.7019139	.
Vitvi13g01242	.	0.5280905	0.6108470	.	.
Vitvi14g00070	.	.	.	0.6453745	.
Vitvi14g00950	.	.	-0.5104399	.	.
Vitvi14g01692
Vitvi14g01873	.	.	-0.5188734	.	.
Vitvi14g01968	.	.	.	0.6587398	.
Vitvi15g01127	.	0.5732856	0.5861104	.	.
Vitvi15g01128	.	.	.	0.5818832	.
Vitvi16g00033	.	0.6523071	0.7274074	-0.5178418	.
Vitvi16g01405	0.6756019	0.6392914	0.6221035	.	.
Vitvi17g00738	.	0.6715986	0.7211818	.	.
Vitvi18g00111	0.5781228	.	.	0.5330754	.
Vitvi18g00144
Vitvi18g00454	.	0.5026635	0.6234153	-0.6590534	.
	Galactinol	Galactose	Myo.Inostol	Raffinose	Xylose
Vitvi01g00052
Vitvi01g00053
Vitvi01g00064	.	0.5407503	.	.	.
Vitvi01g00932
Vitvi02g00184
Vitvi02g00250	.	.	0.5336371	.	.
Vitvi02g00512
Vitvi02g00605	-0.5061631	.	.	-0.5411817	.
Vitvi03g00088	.	.	0.6300600	.	.
Vitvi03g00304	-0.5642102	.	.	-0.5661412	.
Vitvi05g00164	-0.6148967	.	.	-0.6065619	.

Vitvi05g00357	0.5489211	.	.	0.5047854	.
Vitvi05g00442	.	.	0.5223330	.	.
Vitvi05g01324	.	.	.	0.5495453	.
Vitvi05g01516
Vitvi06g00583
Vitvi06g01272	0.5182567
Vitvi06g01427	0.5026603	.	.	0.5449772	.
Vitvi07g00353	0.6480885	.	.	0.6396144	.
Vitvi07g01830	.	0.5072740	.	.	.
Vitvi08g00225
Vitvi08g00930	-0.5027461
Vitvi08g01307	0.6879714	.	.	0.7258160	.
Vitvi08g01353
Vitvi09g00193	.	.	-0.5552590	.	.
Vitvi10g00094	.	.	0.5086231	.	.
Vitvi10g00494	.	-0.5280703	-0.7596087	.	.
Vitvi10g00499
Vitvi10g00739
Vitvi11g00030
Vitvi11g00260
Vitvi11g00542	0.5654033	.	.	0.5769190	.
Vitvi11g00903	.	.	.	-0.5281850	.
Vitvi12g00770	0.7223197	.	.	0.7719318	.
Vitvi12g01618
Vitvi13g00792	0.6652972	.	.	0.6644739	.
Vitvi13g01242
Vitvi14g00070
Vitvi14g00950	0.6886845	.	.	0.7579625	-0.5287528
Vitvi14g01692
Vitvi14g01873
Vitvi14g01968	0.6387211	.	.	0.6424206	.
Vitvi15g01127	.	0.5105147	.	.	.
Vitvi15g01128
Vitvi16g00033	.	0.5554995	.	.	.
Vitvi16g01405	.	0.5726032	0.6985457	.	.
Vitvi17g00738	.	0.5747023	0.5261155	.	.
Vitvi18g00111	0.6461834	.	.	0.6865313	.
Vitvi18g00144
Vitvi18g00454	-0.5153908
	Fumaric.acid	Succinic.acid	Arabinose	Melibiose	
Vitvi01g00052	.	.	.	-0.5072784	.
Vitvi01g00053
Vitvi01g00064	.	.	.	-0.6011602	.
Vitvi01g00932
Vitvi02g00184	.	.	.	0.5421271	.
Vitvi02g00250	.	0.5630187	.	.	.
Vitvi02g00512
Vitvi02g00605
Vitvi03g00088	.	0.5167690	.	.	.
Vitvi03g00304	.	.	.	-0.6177412	.
Vitvi05g00164	.	.	.	-0.6426583	.
Vitvi05g00357	.	.	.	0.7296386	.
Vitvi05g00442	.	0.5011379	.	.	.

Vitvi05g01324
Vitvi05g01516	.	.	.	-0.6337813
Vitvi06g00583	.	.	.	-0.6368632
Vitvi06g01272	.	.	.	-0.5208497
Vitvi06g01427
Vitvi07g00353	.	.	.	0.7266886
Vitvi07g01830
Vitvi08g00225
Vitvi08g00930	.	.	.	-0.6441902
Vitvi08g01307	.	.	.	0.6008460
Vitvi08g01353	0.5531890	0.5508801	.	0.5549709
Vitvi09g00193
Vitvi10g00094	.	0.5454805	.	.
Vitvi10g00494	.	-0.6636659	-0.5290075	.
Vitvi10g00499
Vitvi10g00739
Vitvi11g00030	.	.	.	-0.6417518
Vitvi11g00260	.	.	.	-0.5607300
Vitvi11g00542	.	.	.	0.5944912
Vitvi11g00903
Vitvi12g00770	.	0.5373711	.	0.5908220
Vitvi12g01618
Vitvi13g00792	.	.	.	0.7003441
Vitvi13g01242
Vitvi14g00070	.	.	.	0.6006570
Vitvi14g00950	.	.	.	0.5576167
Vitvi14g01692
Vitvi14g01873
Vitvi14g01968	.	.	.	0.6453639
Vitvi15g01127
Vitvi15g01128	.	.	.	0.5325822
Vitvi16g00033	.	.	.	-0.5212467
Vitvi16g01405	.	0.5661916	0.5010829	.
Vitvi17g00738	.	.	0.5373556	.
Vitvi18g00111	.	0.5146926	.	0.5323601
Vitvi18g00144
Vitvi18g00454	.	.	.	-0.6652969
	Rhamnose	Ribonic.acid	Ribose	Aspartic.acid Isoleucine
Vitvi01g00052	.	.	.	-0.5001270
Vitvi01g00053
Vitvi01g00064	.	.	.	-0.7424474
Vitvi01g00932	.	.	.	-0.5270769
Vitvi02g00184	.	.	.	0.6343855
Vitvi02g00250	0.5496647	.	.	.
Vitvi02g00512
Vitvi02g00605
Vitvi03g00088
Vitvi03g00304	.	.	.	-0.6516658
Vitvi05g00164	-0.5996784	.	.	-0.5998493
Vitvi05g00357	.	.	.	0.8025903
Vitvi05g00442
Vitvi05g01324
Vitvi05g01516	.	.	.	-0.7492025

Vitvi06g00583	-0.7020788
Vitvi06g01272	-0.6211757
Vitvi06g01427
Vitvi07g00353	0.7559402
Vitvi07g01830	-0.6128611
Vitvi08g00225
Vitvi08g00930	-0.6994162
Vitvi08g01307	0.5804706	.	.	.	0.5323392
Vitvi08g01353	0.6002550
Vitvi09g00193
Vitvi10g00094	0.5370849
Vitvi10g00494	-0.5495490
Vitvi10g00499
Vitvi10g00739
Vitvi11g00030	-0.6677243
Vitvi11g00260	-0.5597039
Vitvi11g00542	0.6234814
Vitvi11g00903
Vitvi12g00770	0.6337338	.	.	-0.5422296	.
Vitvi12g01618
Vitvi13g00792	0.5608191	.	.	.	0.6884167
Vitvi13g01242	-0.5910614
Vitvi14g00070	0.6183744
Vitvi14g00950	.	-0.5701289	.	-0.5052852	0.5270667
Vitvi14g01692
Vitvi14g01873
Vitvi14g01968	0.5743971	.	.	.	0.6085301
Vitvi15g01127
Vitvi15g01128	0.5934910
Vitvi16g00033	-0.6906635
Vitvi16g01405
Vitvi17g00738	-0.5357129
Vitvi18g00111	0.5924932
Vitvi18g00144
Vitvi18g00454	-0.7329634
	Leucine	Phenylalanine	Proline	Shikimic.acid	
Vitvi01g00052	-0.5375627	-0.5201128	.	.	.
Vitvi01g00053
Vitvi01g00064	-0.7868877	-0.7464952	-0.6815598	0.7438648	.
Vitvi01g00932	-0.5594046	-0.5321456	.	0.5380327	.
Vitvi02g00184	0.6644821	0.6139496	0.5125041	.	.
Vitvi02g00250
Vitvi02g00512
Vitvi02g00605	.	.	.	0.5040728	.
Vitvi03g00088
Vitvi03g00304	-0.7023422	-0.6850097	-0.6056248	0.6637650	.
Vitvi05g00164	-0.6430017	-0.6173325	.	.	.
Vitvi05g00357	0.8547175	0.8153513	0.7043154	-0.7168004	.
Vitvi05g00442
Vitvi05g01324
Vitvi05g01516	-0.7929027	-0.7488376	-0.6608856	0.6815789	.
Vitvi06g00583	-0.7454856	-0.7069859	-0.6034403	0.5956637	.
Vitvi06g01272	-0.6661214	-0.6461686	-0.6057164	0.7090364	.

Vitvi06g01427
Vitvi07g00353	0.8120871	0.7865908	0.6787194	-0.7099583
Vitvi07g01830	-0.6462523	-0.6082535	-0.5683218	0.6312728
Vitvi08g00225
Vitvi08g00930	-0.7459625	-0.7134089	-0.6147025	0.6262617
Vitvi08g01307	0.5806363	0.5753684	.	.
Vitvi08g01353	0.5270417	.	.	.
Vitvi09g00193
Vitvi10g00094
Vitvi10g00494
Vitvi10g00499
Vitvi10g00739
Vitvi11g00030	-0.7085331	-0.6694957	-0.5447569	.
Vitvi11g00260	-0.5969123	-0.5688666	.	.
Vitvi11g00542	0.6741509	0.6615296	0.5905538	-0.6624044
Vitvi11g00903	.	.	.	0.5253341
Vitvi12g00770	0.5393360	0.5382882	.	.
Vitvi12g01618	.	.	.	0.5500592
Vitvi13g00792	0.7405442	0.7175607	0.5962875	-0.5891298
Vitvi13g01242	-0.6289968	-0.6013923	-0.5541727	0.6204702
Vitvi14g00070	0.6562136	0.6198796	0.5001814	.
Vitvi14g00950	0.5828316	0.5941418	0.5399600	-0.6565716
Vitvi14g01692
Vitvi14g01873	0.5082042	0.5023231	.	-0.5719694
Vitvi14g01968	0.6552243	0.6349499	0.5118332	.
Vitvi15g01127
Vitvi15g01128	0.6248760	0.5825940	.	.
Vitvi16g00033	-0.7272217	-0.6822653	-0.6317630	0.6895450
Vitvi16g01405
Vitvi17g00738	-0.5661298	-0.5372944	-0.5398613	0.6629354
Vitvi18g00111
Vitvi18g00144
Vitvi18g00454	-0.7823342	-0.7497481	-0.6549974	0.6839836
	Valine	Trans.Caffeic.acid	Catechin	Gallic.acid
Vitvi01g00052	-0.5094169	.	.	.
Vitvi01g00053	.	0.5120928	.	.
Vitvi01g00064	-0.6332854	.	.	.
Vitvi01g00932
Vitvi02g00184	0.5711039	.	.	.
Vitvi02g00250
Vitvi02g00512
Vitvi02g00605
Vitvi03g00088	.	.	0.7003675	.
Vitvi03g00304	-0.6252328	.	.	.
Vitvi05g00164	-0.6416698	.	.	.
Vitvi05g00357	0.7503898	.	.	.
Vitvi05g00442
Vitvi05g01324
Vitvi05g01516	-0.6632117	.	.	.
Vitvi06g00583	-0.6567409	.	.	.
Vitvi06g01272	-0.5398154	.	.	.
Vitvi06g01427
Vitvi07g00353	0.7358117	.	.	.

Vitvi07g01830
Vitvi08g00225
Vitvi08g00930	-0.6603281	.	.	.
Vitvi08g01307	0.5887340	.	.	.
Vitvi08g01353	0.5574319	.	.	.
Vitvi09g00193
Vitvi10g00094
Vitvi10g00494	.	-0.8387141	.	.
Vitvi10g00499
Vitvi10g00739	.	0.5215284	.	.
Vitvi11g00030	-0.6559606	.	.	.
Vitvi11g00260	-0.5673668	.	.	.
Vitvi11g00542	0.5996506	.	.	.
Vitvi11g00903
Vitvi12g00770	0.5726808	.	.	.
Vitvi12g01618
Vitvi13g00792	0.7025798	.	.	.
Vitvi13g01242	-0.5075587	.	.	.
Vitvi14g00070	0.6128999	.	.	.
Vitvi14g00950	0.5452770	.	.	.
Vitvi14g01692
Vitvi14g01873
Vitvi14g01968	0.6432446	.	.	.
Vitvi15g01127	.	0.6776708	.	.
Vitvi15g01128	0.5538959	.	.	.
Vitvi16g00033	-0.5597653	0.5331796	.	.
Vitvi16g01405	.	0.8447080	.	.
Vitvi17g00738	.	0.6268607	.	.
Vitvi18g00111	0.5157865	.	.	.
Vitvi18g00144
Vitvi18g00454	-0.6831610	.	.	.
	Hydroquinone	Quinic.acid	X3.caffeoylquinic.acid	
Vitvi01g00052
Vitvi01g00053
Vitvi01g00064	0.6994939	.	.	.
Vitvi01g00932	0.5007135	.	.	.
Vitvi02g00184
Vitvi02g00250	.	.	0.5387997	.
Vitvi02g00512
Vitvi02g00605
Vitvi03g00088	0.5630329	0.6939235	0.6698343	.
Vitvi03g00304
Vitvi05g00164
Vitvi05g00357	-0.5725401	.	.	.
Vitvi05g00442
Vitvi05g01324
Vitvi05g01516	0.6197121	.	.	.
Vitvi06g00583
Vitvi06g01272	0.6079765	.	.	.
Vitvi06g01427
Vitvi07g00353
Vitvi07g01830	0.6472017	.	.	.
Vitvi08g00225

Vitvi08g00930	.	.	.
Vitvi08g01307	.	.	.
Vitvi08g01353	.	.	.
Vitvi09g00193	.	-0.5175593	.
Vitvi10g00094	.	.	0.5279115
Vitvi10g00494	-0.6133343	-0.8262798	-0.8289195
Vitvi10g00499	.	.	.
Vitvi10g00739	.	.	.
Vitvi11g00030	.	.	.
Vitvi11g00260	.	.	.
Vitvi11g00542	.	.	.
Vitvi11g00903	.	.	.
Vitvi12g00770	.	.	0.5538163
Vitvi12g01618	.	.	.
Vitvi13g00792	.	.	.
Vitvi13g01242	0.5639726	.	.
Vitvi14g00070	.	.	.
Vitvi14g00950	.	.	.
Vitvi14g01692	.	.	.
Vitvi14g01873	.	.	.
Vitvi14g01968	.	.	.
Vitvi15g01127	0.6359434	0.6343683	0.6107334
Vitvi15g01128	.	.	.
Vitvi16g00033	0.7116590	0.5198050	.
Vitvi16g01405	0.6868407	0.8144652	0.8058608
Vitvi17g00738	0.7126351	0.6295102	0.5082886
Vitvi18g00111	.	.	0.5267319
Vitvi18g00144	.	.	.
Vitvi18g00454	0.5422471	.	.
	Gluconic.acid	Glucopyranose..H2O.	Lyxonic.acid
Vitvi01g00052	.	.	.
Vitvi01g00053	.	.	.
Vitvi01g00064	.	.	.
Vitvi01g00932	.	.	.
Vitvi02g00184	.	.	.
Vitvi02g00250	.	0.6086968	.
Vitvi02g00512	.	-0.5212760	.
Vitvi02g00605	0.5586950	.	.
Vitvi03g00088	.	.	.
Vitvi03g00304	0.6251725	.	.
Vitvi05g00164	0.5283956	-0.6541414	.
Vitvi05g00357	-0.5858553	.	.
Vitvi05g00442	.	0.5147132	.
Vitvi05g01324	.	.	.
Vitvi05g01516	.	.	.
Vitvi06g00583	.	.	.
Vitvi06g01272	0.5252134	.	.
Vitvi06g01427	.	.	.
Vitvi07g00353	-0.6812949	0.5280083	.
Vitvi07g01830	.	.	.
Vitvi08g00225	.	0.5424304	.
Vitvi08g00930	0.5314976	.	.
Vitvi08g01307	-0.6479792	0.6148165	.

Vitvi08g01353	.	0.6720628	.	.
Vitvi09g00193
Vitvi10g00094	.	0.5925482	.	.
Vitvi10g00494	.	-0.5890609	.	.
Vitvi10g00499	.	0.5402904	.	.
Vitvi10g00739
Vitvi11g00030	.	-0.5273154	.	.
Vitvi11g00260
Vitvi11g00542	-0.6366641	.	.	.
Vitvi11g00903	0.5785848	.	.	.
Vitvi12g00770	-0.6544356	0.6699599	.	.
Vitvi12g01618
Vitvi13g00792	-0.6447671	0.6055384	.	.
Vitvi13g01242
Vitvi14g00070	.	0.5096460	.	.
Vitvi14g00950	-0.7725667	.	.	.
Vitvi14g01692	.	0.5025383	.	.
Vitvi14g01873
Vitvi14g01968	-0.5854367	0.6207134	.	.
Vitvi15g01127
Vitvi15g01128
Vitvi16g00033
Vitvi16g01405
Vitvi17g00738
Vitvi18g00111	-0.5650205	0.6295386	.	.
Vitvi18g00144	.	0.5114631	.	.
Vitvi18g00454	0.5654670	.	.	.
	Maleic.acid	Malonic.acid	Phosphoric.acid	Tartaric.acid
Vitvi01g00052
Vitvi01g00053
Vitvi01g00064
Vitvi01g00932
Vitvi02g00184
Vitvi02g00250
Vitvi02g00512
Vitvi02g00605
Vitvi03g00088	.	.	0.5945147	.
Vitvi03g00304	.	0.5451463	.	.
Vitvi05g00164	.	.	.	0.5442610
Vitvi05g00357	.	-0.5545540	.	.
Vitvi05g00442
Vitvi05g01324
Vitvi05g01516
Vitvi06g00583
Vitvi06g01272
Vitvi06g01427
Vitvi07g00353	.	-0.5973837	.	.
Vitvi07g01830
Vitvi08g00225
Vitvi08g00930
Vitvi08g01307	.	-0.5028139	.	-0.5498118
Vitvi08g01353	0.5199761	.	.	-0.5008292
Vitvi09g00193	-0.5283050	.	.	.

Vitvi10g00094
Vitvi10g00494	.	.	-0.7028820	.
Vitvi10g00499
Vitvi10g00739	0.5208453	.	.	.
Vitvi11g00030
Vitvi11g00260
Vitvi11g00542	.	-0.5472642	.	.
Vitvi11g00903
Vitvi12g00770	.	.	.	-0.5887235
Vitvi12g01618
Vitvi13g00792	.	-0.5454725	.	-0.5387237
Vitvi13g01242
Vitvi14g00070
Vitvi14g00950	.	-0.6051287	.	.
Vitvi14g01692
Vitvi14g01873
Vitvi14g01968	.	.	.	-0.5361714
Vitvi15g01127	.	.	0.5323527	.
Vitvi15g01128
Vitvi16g00033
Vitvi16g01405	.	.	0.6886713	.
Vitvi17g00738	.	.	0.5532800	.
Vitvi18g00111	.	.	.	-0.5415554
Vitvi18g00144
Vitvi18g00454	.	0.5282788	.	.
	Threonolactone			
Vitvi01g00052	0.5161791			
Vitvi01g00053	.			
Vitvi01g00064	0.5770477			
Vitvi01g00932	.			
Vitvi02g00184	.			
Vitvi02g00250	.			
Vitvi02g00512	.			
Vitvi02g00605	0.5918576			
Vitvi03g00088	.			
Vitvi03g00304	0.6971782			
Vitvi05g00164	0.5958311			
Vitvi05g00357	-0.6959151			
Vitvi05g00442	.			
Vitvi05g01324	.			
Vitvi05g01516	0.5675827			
Vitvi06g00583	0.5746589			
Vitvi06g01272	0.6050046			
Vitvi06g01427	-0.5102756			
Vitvi07g00353	-0.7689997			
Vitvi07g01830	.			
Vitvi08g00225	.			
Vitvi08g00930	0.6247236			
Vitvi08g01307	-0.6870238			
Vitvi08g01353	.			
Vitvi09g00193	.			
Vitvi10g00094	.			
Vitvi10g00494	.			

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Vitvi10g00499      .
Vitvi10g00739      .
Vitvi11g00030      0.5431952
Vitvi11g00260      0.5043227
Vitvi11g00542      -0.7012422
Vitvi11g00903      0.5892563
Vitvi12g00770      -0.6826653
Vitvi12g01618      .
Vitvi13g00792      -0.7200024
Vitvi13g01242      .
Vitvi14g00070      -0.5044696
Vitvi14g00950      -0.7991375
Vitvi14g01692      .
Vitvi14g01873      -0.5438370
Vitvi14g01968      -0.6488988
Vitvi15g01127      .
Vitvi15g01128      .
Vitvi16g00033      .
Vitvi16g01405      .
Vitvi17g00738      .
Vitvi18g00111      -0.5912697
Vitvi18g00144      .
Vitvi18g00454      0.6627704

```

50 / 1363 rows printed

> *sapply*(*net\$gR*, *print*)

\$Fructose

+ 365/1400 vertices, named, from ac77ef8:

```

[1] Vitvi02g00250 Vitvi03g00088 Vitvi05g00164 Vitvi05g00442
[5] Vitvi08g00225 Vitvi08g01307 Vitvi08g01353 Vitvi09g00193
[9] Vitvi10g00094 Vitvi10g00494 Vitvi12g00770 Vitvi16g01405
[13] Vitvi18g00111 Vitvi18g02417 Vitvi00g01259 Vitvi01g00140
[17] Vitvi02g00514 Vitvi03g00029 Vitvi06g01444 Vitvi07g00721
[21] Vitvi10g02085 Vitvi11g01277 Vitvi14g01717 Vitvi16g01858
[25] Vitvi16g00929 Vitvi17g00192 Vitvi17g00957 Vitvi07g03070
[29] Vitvi01g00274 Vitvi04g01280 Vitvi05g00207 Vitvi07g00599
[33] Vitvi08g01454 Vitvi09g00720 Vitvi10g00554 Vitvi10g01386
[37] Vitvi11g00561 Vitvi14g00138 Vitvi14g01627 Vitvi19g00502

```

+ ... omitted several vertices

\$Glucose

+ 352/1400 vertices, named, from ac77ef8:

```

[1] Vitvi01g00064 Vitvi03g00088 Vitvi05g00357 Vitvi05g01516
[5] Vitvi06g01272 Vitvi07g01830 Vitvi10g00494 Vitvi13g01242
[9] Vitvi15g01127 Vitvi16g00033 Vitvi16g01405 Vitvi17g00738
[13] Vitvi18g00454 Vitvi18g02365 Vitvi19g00704 Vitvi01g00793
[17] Vitvi02g00514 Vitvi04g01689 Vitvi07g01365 Vitvi10g00674
[21] Vitvi14g02461 Vitvi14g01911 Vitvi14g01916 Vitvi16g01858
[25] Vitvi16g01363 Vitvi18g00994 Vitvi01g00698 Vitvi05g00129
[29] Vitvi09g00720 Vitvi10g00073 Vitvi11g00147 Vitvi13g01832
[33] Vitvi01g00456 Vitvi01g00784 Vitvi01g01805 Vitvi02g00125

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[37] Vitvi02g01344 Vitvi02g01230 Vitvi03g01388 Vitvi03g00274
+ ... omitted several vertices

\$Sucrose

+ 535/1400 vertices, named, from ac77ef8:

[1] Vitvi01g00064 Vitvi01g00932 Vitvi03g00088 Vitvi03g00304
[5] Vitvi05g00357 Vitvi05g01516 Vitvi06g00583 Vitvi06g01272
[9] Vitvi07g00353 Vitvi07g01830 Vitvi08g00930 Vitvi10g00494
[13] Vitvi11g00542 Vitvi12g01618 Vitvi13g01242 Vitvi14g00950
[17] Vitvi14g01873 Vitvi15g01127 Vitvi16g00033 Vitvi16g01405
[21] Vitvi17g00738 Vitvi18g00454 Vitvi18g01114 Vitvi18g02365
[25] Vitvi19g00704 Vitvi01g00793 Vitvi02g00191 Vitvi02g00514
[29] Vitvi03g00056 Vitvi04g01689 Vitvi05g00170 Vitvi06g00121
[33] Vitvi07g00431 Vitvi07g00457 Vitvi07g02242 Vitvi07g01365
[37] Vitvi08g01890 Vitvi10g00436 Vitvi10g00674 Vitvi11g00513

+ ... omitted several vertices

\$Erythronic.acid

+ 555/1400 vertices, named, from ac77ef8:

[1] Vitvi01g00052 Vitvi01g00064 Vitvi02g00184 Vitvi02g00512
[5] Vitvi03g00304 Vitvi05g00164 Vitvi05g00357 Vitvi05g01516
[9] Vitvi06g00583 Vitvi07g00353 Vitvi08g00930 Vitvi08g01307
[13] Vitvi08g01353 Vitvi10g00499 Vitvi11g00030 Vitvi11g00260
[17] Vitvi11g00542 Vitvi12g00770 Vitvi13g00792 Vitvi14g00070
[21] Vitvi14g01968 Vitvi15g01128 Vitvi16g00033 Vitvi18g00111
[25] Vitvi18g00454 Vitvi01g00714 Vitvi02g00191 Vitvi03g00029
[29] Vitvi05g00170 Vitvi06g00121 Vitvi06g01444 Vitvi07g00457
[33] Vitvi07g02242 Vitvi07g01777 Vitvi08g01890 Vitvi10g00436
[37] Vitvi10g00625 Vitvi10g00674 Vitvi11g00513 Vitvi12g00307

+ ... omitted several vertices

\$Galactinol

+ 529/1400 vertices, named, from ac77ef8:

[1] Vitvi02g00605 Vitvi03g00304 Vitvi05g00164 Vitvi05g00357
[5] Vitvi06g01427 Vitvi07g00353 Vitvi08g00930 Vitvi08g01307
[9] Vitvi11g00542 Vitvi12g00770 Vitvi13g00792 Vitvi14g00950
[13] Vitvi14g01968 Vitvi18g00111 Vitvi18g00454 Vitvi18g01114
[17] Vitvi01g00714 Vitvi02g00191 Vitvi03g00029 Vitvi04g00117
[21] Vitvi05g00170 Vitvi06g01444 Vitvi07g00457 Vitvi07g02242
[25] Vitvi07g01777 Vitvi08g01890 Vitvi10g00436 Vitvi11g00513
[29] Vitvi12g00307 Vitvi13g00157 Vitvi14g00063 Vitvi14g01717
[33] Vitvi16g00086 Vitvi16g00929 Vitvi16g01363 Vitvi17g00885
[37] Vitvi17g00957 Vitvi19g00016 Vitvi07g03070 Vitvi01g00274

+ ... omitted several vertices

\$Galactose

+ 149/1400 vertices, named, from ac77ef8:

[1] Vitvi01g00064 Vitvi07g01830 Vitvi10g00494 Vitvi15g01127
[5] Vitvi16g00033 Vitvi16g01405 Vitvi17g00738 Vitvi01g00793
[9] Vitvi04g01689 Vitvi07g01365 Vitvi14g02461 Vitvi14g01916
[13] Vitvi18g00994 Vitvi09g00720 Vitvi11g00147 Vitvi13g01832
[17] Vitvi02g01344 Vitvi02g01230 Vitvi03g01388 Vitvi05g00676
[21] Vitvi07g00034 Vitvi07g00525 Vitvi07g00665 Vitvi11g00726

[25] Vitvi12g00024 Vitvi12g02374 Vitvi12g01115 Vitvi12g02167
[29] Vitvi13g01966 Vitvi13g00352 Vitvi13g01731 Vitvi14g00017
[33] Vitvi16g01418 Vitvi17g00480 Vitvi18g00008 Vitvi19g00025
[37] Vitvi00g00869 Vitvi01g00664 Vitvi01g01563 Vitvi04g00328
+ ... omitted several vertices

\$Myo.Inostol

+ 272/1400 vertices, named, from ac77ef8:
[1] Vitvi02g00250 Vitvi03g00088 Vitvi05g00442 Vitvi09g00193
[5] Vitvi10g00094 Vitvi10g00494 Vitvi16g01405 Vitvi17g00738
[9] Vitvi00g01259 Vitvi02g00514 Vitvi07g00016 Vitvi07g01365
[13] Vitvi10g02085 Vitvi11g01277 Vitvi14g02461 Vitvi14g01911
[17] Vitvi16g01858 Vitvi18g00994 Vitvi07g03070 Vitvi01g00698
[21] Vitvi05g00207 Vitvi09g00720 Vitvi10g01386 Vitvi11g00147
[25] Vitvi11g00561 Vitvi13g00719 Vitvi13g01832 Vitvi14g00138
[29] Vitvi19g01739 Vitvi10g02279 Vitvi01g01838 Vitvi02g01341
[33] Vitvi02g01344 Vitvi03g01388 Vitvi03g01138 Vitvi05g00676
[37] Vitvi07g00034 Vitvi07g00376 Vitvi07g02249 Vitvi07g00513
+ ... omitted several vertices

\$Raffinose

+ 584/1400 vertices, named, from ac77ef8:
[1] Vitvi02g00605 Vitvi03g00304 Vitvi05g00164 Vitvi05g00357
[5] Vitvi05g01324 Vitvi06g01427 Vitvi07g00353 Vitvi08g01307
[9] Vitvi11g00542 Vitvi11g00903 Vitvi12g00770 Vitvi13g00792
[13] Vitvi14g00950 Vitvi14g01968 Vitvi18g00111 Vitvi18g01114
[17] Vitvi18g02365 Vitvi01g00140 Vitvi01g01440 Vitvi02g00191
[21] Vitvi03g00029 Vitvi04g00117 Vitvi05g00170 Vitvi06g01444
[25] Vitvi07g00457 Vitvi07g02242 Vitvi07g01777 Vitvi08g02210
[29] Vitvi10g00436 Vitvi11g00513 Vitvi11g01277 Vitvi12g00307
[33] Vitvi13g00157 Vitvi14g00063 Vitvi14g01717 Vitvi15g01026
[37] Vitvi16g00086 Vitvi16g00929 Vitvi16g01363 Vitvi17g00192
+ ... omitted several vertices

\$Xylose

+ 228/1400 vertices, named, from ac77ef8:
[1] Vitvi06g01272 Vitvi14g00950 Vitvi18g01114 Vitvi18g02365
[5] Vitvi19g00538 Vitvi19g00704 Vitvi01g00793 Vitvi01g01440
[9] Vitvi03g00056 Vitvi04g01689 Vitvi07g00431 Vitvi07g00457
[13] Vitvi07g01365 Vitvi14g02461 Vitvi14g01911 Vitvi14g01916
[17] Vitvi16g01363 Vitvi18g00994 Vitvi01g00698 Vitvi13g01832
[21] Vitvi14g01732 Vitvi17g00607 Vitvi02g00125 Vitvi02g00423
[25] Vitvi02g01230 Vitvi03g01388 Vitvi03g00274 Vitvi04g00735
[29] Vitvi04g01334 Vitvi05g00676 Vitvi05g01353 Vitvi06g00435
[33] Vitvi07g00034 Vitvi07g00525 Vitvi08g00233 Vitvi08g00877
[37] Vitvi08g01523 Vitvi08g01897 Vitvi11g01265 Vitvi11g01268
+ ... omitted several vertices

\$Fumaric.acid

+ 30/1400 vertices, named, from ac77ef8:
[1] Vitvi08g01353 Vitvi18g02757 Vitvi05g00503 Vitvi14g01995
[5] Vitvi18g00549 Vitvi04g01854 Vitvi17g00555 Vitvi10g02269
[9] Vitvi01g00444 Vitvi01g01026 Vitvi01g01447 Vitvi02g00089

[13] Vitvi02g00121 Vitvi04g01714 Vitvi05g00104 Vitvi06g00024
[17] Vitvi06g00382 Vitvi06g00800 Vitvi06g01465 Vitvi07g00002
[21] Vitvi07g00452 Vitvi08g00730 Vitvi08g01134 Vitvi10g00270
[25] Vitvi11g01145 Vitvi14g01901 Vitvi15g00948 Vitvi16g01362
[29] Vitvi18g00538 Vitvi19g00322

\$Fructose

+ 365/1400 vertices, named, from ac77ef8:

[1] Vitvi02g00250 Vitvi03g00088 Vitvi05g00164 Vitvi05g00442
[5] Vitvi08g00225 Vitvi08g01307 Vitvi08g01353 Vitvi09g00193
[9] Vitvi10g00094 Vitvi10g00494 Vitvi12g00770 Vitvi16g01405
[13] Vitvi18g00111 Vitvi18g02417 Vitvi00g01259 Vitvi01g00140
[17] Vitvi02g00514 Vitvi03g00029 Vitvi06g01444 Vitvi07g00721
[21] Vitvi10g02085 Vitvi11g01277 Vitvi14g01717 Vitvi16g01858
[25] Vitvi16g00929 Vitvi17g00192 Vitvi17g00957 Vitvi07g03070
[29] Vitvi01g00274 Vitvi04g01280 Vitvi05g00207 Vitvi07g00599
[33] Vitvi08g01454 Vitvi09g00720 Vitvi10g00554 Vitvi10g01386
[37] Vitvi11g00561 Vitvi14g00138 Vitvi14g01627 Vitvi19g00502

+ ... omitted several vertices

\$Glucose

+ 352/1400 vertices, named, from ac77ef8:

[1] Vitvi01g00064 Vitvi03g00088 Vitvi05g00357 Vitvi05g01516
[5] Vitvi06g01272 Vitvi07g01830 Vitvi10g00494 Vitvi13g01242
[9] Vitvi15g01127 Vitvi16g00033 Vitvi16g01405 Vitvi17g00738
[13] Vitvi18g00454 Vitvi18g02365 Vitvi19g00704 Vitvi01g00793
[17] Vitvi02g00514 Vitvi04g01689 Vitvi07g01365 Vitvi10g00674
[21] Vitvi14g02461 Vitvi14g01911 Vitvi14g01916 Vitvi16g01858
[25] Vitvi16g01363 Vitvi18g00994 Vitvi01g00698 Vitvi05g00129
[29] Vitvi09g00720 Vitvi10g00073 Vitvi11g00147 Vitvi13g01832
[33] Vitvi01g00456 Vitvi01g00784 Vitvi01g01805 Vitvi02g00125
[37] Vitvi02g01344 Vitvi02g01230 Vitvi03g01388 Vitvi03g00274

+ ... omitted several vertices

\$Sucrose

+ 535/1400 vertices, named, from ac77ef8:

[1] Vitvi01g00064 Vitvi01g00932 Vitvi03g00088 Vitvi03g00304
[5] Vitvi05g00357 Vitvi05g01516 Vitvi06g00583 Vitvi06g01272
[9] Vitvi07g00353 Vitvi07g01830 Vitvi08g00930 Vitvi10g00494
[13] Vitvi11g00542 Vitvi12g01618 Vitvi13g01242 Vitvi14g00950
[17] Vitvi14g01873 Vitvi15g01127 Vitvi16g00033 Vitvi16g01405
[21] Vitvi17g00738 Vitvi18g00454 Vitvi18g01114 Vitvi18g02365
[25] Vitvi19g00704 Vitvi01g00793 Vitvi02g00191 Vitvi02g00514
[29] Vitvi03g00056 Vitvi04g01689 Vitvi05g00170 Vitvi06g00121
[33] Vitvi07g00431 Vitvi07g00457 Vitvi07g02242 Vitvi07g01365
[37] Vitvi08g01890 Vitvi10g00436 Vitvi10g00674 Vitvi11g00513

+ ... omitted several vertices

\$Erythronic.acid

+ 555/1400 vertices, named, from ac77ef8:

[1] Vitvi01g00052 Vitvi01g00064 Vitvi02g00184 Vitvi02g00512
[5] Vitvi03g00304 Vitvi05g00164 Vitvi05g00357 Vitvi05g01516
[9] Vitvi06g00583 Vitvi07g00353 Vitvi08g00930 Vitvi08g01307
[13] Vitvi08g01353 Vitvi10g00499 Vitvi11g00030 Vitvi11g00260

[17] Vitvi11g00542 Vitvi12g00770 Vitvi13g00792 Vitvi14g00070
[21] Vitvi14g01968 Vitvi15g01128 Vitvi16g00033 Vitvi18g00111
[25] Vitvi18g00454 Vitvi01g00714 Vitvi02g00191 Vitvi03g00029
[29] Vitvi05g00170 Vitvi06g00121 Vitvi06g01444 Vitvi07g00457
[33] Vitvi07g02242 Vitvi07g01777 Vitvi08g01890 Vitvi10g00436
[37] Vitvi10g00625 Vitvi10g00674 Vitvi11g00513 Vitvi12g00307
+ ... omitted several vertices

\$Galactinol

+ 529/1400 vertices, named, from ac77ef8:

[1] Vitvi02g00605 Vitvi03g00304 Vitvi05g00164 Vitvi05g00357
[5] Vitvi06g01427 Vitvi07g00353 Vitvi08g00930 Vitvi08g01307
[9] Vitvi11g00542 Vitvi12g00770 Vitvi13g00792 Vitvi14g00950
[13] Vitvi14g01968 Vitvi18g00111 Vitvi18g00454 Vitvi18g01114
[17] Vitvi01g00714 Vitvi02g00191 Vitvi03g00029 Vitvi04g00117
[21] Vitvi05g00170 Vitvi06g01444 Vitvi07g00457 Vitvi07g02242
[25] Vitvi07g01777 Vitvi08g01890 Vitvi10g00436 Vitvi11g00513
[29] Vitvi12g00307 Vitvi13g00157 Vitvi14g00063 Vitvi14g01717
[33] Vitvi16g00086 Vitvi16g00929 Vitvi16g01363 Vitvi17g00885
[37] Vitvi17g00957 Vitvi19g00016 Vitvi07g03070 Vitvi01g00274
+ ... omitted several vertices

\$Galactose

+ 149/1400 vertices, named, from ac77ef8:

[1] Vitvi01g00064 Vitvi07g01830 Vitvi10g00494 Vitvi15g01127
[5] Vitvi16g00033 Vitvi16g01405 Vitvi17g00738 Vitvi01g00793
[9] Vitvi04g01689 Vitvi07g01365 Vitvi14g02461 Vitvi14g01916
[13] Vitvi18g00994 Vitvi09g00720 Vitvi11g00147 Vitvi13g01832
[17] Vitvi02g01344 Vitvi02g01230 Vitvi03g01388 Vitvi05g00676
[21] Vitvi07g00034 Vitvi07g00525 Vitvi07g00665 Vitvi11g00726
[25] Vitvi12g00024 Vitvi12g02374 Vitvi12g01115 Vitvi12g02167
[29] Vitvi13g01966 Vitvi13g00352 Vitvi13g01731 Vitvi14g00017
[33] Vitvi16g01418 Vitvi17g00480 Vitvi18g00008 Vitvi19g00025
[37] Vitvi00g00869 Vitvi01g00664 Vitvi01g01563 Vitvi04g00328
+ ... omitted several vertices

\$Myo.Inositol

+ 272/1400 vertices, named, from ac77ef8:

[1] Vitvi02g00250 Vitvi03g00088 Vitvi05g00442 Vitvi09g00193
[5] Vitvi10g00094 Vitvi10g00494 Vitvi16g01405 Vitvi17g00738
[9] Vitvi00g01259 Vitvi02g00514 Vitvi07g00016 Vitvi07g01365
[13] Vitvi10g02085 Vitvi11g01277 Vitvi14g02461 Vitvi14g01911
[17] Vitvi16g01858 Vitvi18g00994 Vitvi07g03070 Vitvi01g00698
[21] Vitvi05g00207 Vitvi09g00720 Vitvi10g01386 Vitvi11g00147
[25] Vitvi11g00561 Vitvi13g00719 Vitvi13g01832 Vitvi14g00138
[29] Vitvi19g01739 Vitvi10g02279 Vitvi01g01838 Vitvi02g01341
[33] Vitvi02g01344 Vitvi03g01388 Vitvi03g01138 Vitvi05g00676
[37] Vitvi07g00034 Vitvi07g00376 Vitvi07g02249 Vitvi07g00513
+ ... omitted several vertices

\$Raffinose

+ 584/1400 vertices, named, from ac77ef8:

[1] Vitvi02g00605 Vitvi03g00304 Vitvi05g00164 Vitvi05g00357


```

[5] Vitvi05g01324 Vitvi06g01427 Vitvi07g00353 Vitvi08g01307
[9] Vitvi11g00542 Vitvi11g00903 Vitvi12g00770 Vitvi13g00792
[13] Vitvi14g00950 Vitvi14g01968 Vitvi18g00111 Vitvi18g01114
[17] Vitvi18g02365 Vitvi01g00140 Vitvi01g01440 Vitvi02g00191
[21] Vitvi03g00029 Vitvi04g00117 Vitvi05g00170 Vitvi06g01444
[25] Vitvi07g00457 Vitvi07g02242 Vitvi07g01777 Vitvi08g02210
[29] Vitvi10g00436 Vitvi11g00513 Vitvi11g01277 Vitvi12g00307
[33] Vitvi13g00157 Vitvi14g00063 Vitvi14g01717 Vitvi15g01026
[37] Vitvi16g00086 Vitvi16g00929 Vitvi16g01363 Vitvi17g00192
+ ... omitted several vertices

```

\$Xylose

```

+ 228/1400 vertices, named, from ac77ef8:
[1] Vitvi06g01272 Vitvi14g00950 Vitvi18g01114 Vitvi18g02365
[5] Vitvi19g00538 Vitvi19g00704 Vitvi01g00793 Vitvi01g01440
[9] Vitvi03g00056 Vitvi04g01689 Vitvi07g00431 Vitvi07g00457
[13] Vitvi07g01365 Vitvi14g02461 Vitvi14g01911 Vitvi14g01916
[17] Vitvi16g01363 Vitvi18g00994 Vitvi01g00698 Vitvi13g01832
[21] Vitvi14g01732 Vitvi17g00607 Vitvi02g00125 Vitvi02g00423
[25] Vitvi02g01230 Vitvi03g01388 Vitvi03g00274 Vitvi04g00735
[29] Vitvi04g01334 Vitvi05g00676 Vitvi05g01353 Vitvi06g00435
[33] Vitvi07g00034 Vitvi07g00525 Vitvi08g00233 Vitvi08g00877
[37] Vitvi08g01523 Vitvi08g01897 Vitvi11g01265 Vitvi11g01268
+ ... omitted several vertices

```

\$Fumaric.acid

```

+ 30/1400 vertices, named, from ac77ef8:
[1] Vitvi08g01353 Vitvi18g02757 Vitvi05g00503 Vitvi14g01995
[5] Vitvi18g00549 Vitvi04g01854 Vitvi17g00555 Vitvi10g02269
[9] Vitvi01g00444 Vitvi01g01026 Vitvi01g01447 Vitvi02g00089
[13] Vitvi02g00121 Vitvi04g01714 Vitvi05g00104 Vitvi06g00024
[17] Vitvi06g00382 Vitvi06g00800 Vitvi06g01465 Vitvi07g00002
[21] Vitvi07g00452 Vitvi08g00730 Vitvi08g01134 Vitvi10g00270
[25] Vitvi11g01145 Vitvi14g01901 Vitvi15g00948 Vitvi16g01362
[29] Vitvi18g00538 Vitvi19g00322

```

6.1 Save network with igraph

```

> library(igraph)
> nettip <- "gml"
> (netfn<- gsub("\\.", "_", paste0("network-", cutoff, "_", suffix)))
[1] "network-0_5_18-1-1-2-3-4-5-6-7"
> (netpath <- file.path(.oroot, paste(netfn, nettip, sep=".")))
[1] ".././output/60_Trans-x-Meta-18-1-1-2-3-4-5-6-7/network-0_5_18-1-1-2-3-4-5-6-7"
> write.graph(net$gR, file=netpath, format=nettip)

```

Save correlation matrix (M) used for network construction.

```

> (Mfn <- file.path(.oroot, "network-M.txt"))
[1] ".././output/60_Trans-x-Meta-18-1-1-2-3-4-5-6-7/network-M.txt"
> my.write.table(t(net$M), file=Mfn, label="Correlation matrix (M) used for ne
Warning in write.table(x, file = file, col.names = col.names, sep = sep, : app
Object: t(net$M) \\
Label: Correlation matrix (M) used for network construction \\
File :\\
\\href{run:D:\\DEJAVNOSTI\\OMIKE\\pISA-projects\\_p_VinskaTrta\\_I_EnViRoS\\_S_

```

A Metadata files

A.1 Project metadata

Table 1: Project metadata

Item	Value
project:	_p_VinskaTrta
Short Name:	VinskaTrta
Title:	Vine related research
Description:	*
pISA projects path:	D:/DEJAVNOSTI/OMIKE/pISA-projects
Local pISA-tree organisation:	NIB
pISA project creation date:	2020-11-13
pISA project creator:	AB
Project funding code:	*
Project coordinator:	Marusa Pompe Novak
Project partners:	*
Project start date:	*
Project end date:	*
Principal investigator:	*
License:	CC BY 4.0
Sharing permission:	Private
Upload to FAIRDOMHub:	Yes

A.2 Investigation metadata

Table 2: Investigation metadata

Item	Value
Investigation:	_I_EnViRoS
Short Name:	EnViRoS
Title:	Integration of data from different platforms
Description:	*
Phenodata:	./phenodata_20201113.txt
pISA Investigation creation date:	2020-11-13
pISA Investigation creator:	AB
Principal investigator:	*
License:	CC BY 4.0
Sharing permission:	Private
Upload to FAIRDOMHub:	Yes

A.3 Study metadata

Table 3: Study metadata

Item	Value
Study:	_S_01_Integ
Short Name:	01_Integ
Title:	*
Description:	*
Raw Data:	
pISA Study creation date:	2020-11-13
pISA Study creator:	AB
Principal investigator:	*
License:	CC BY 4.0
Sharing permission:	Private
Upload to FAIRDOMHub:	Yes

A.4 Assay metadata

Table 4: Assay metadata

Item	Value
Assay:	_A_01_Desc-R
Short Name:	01_Desc-R
Assay Class:	DRY
Assay Type:	R
Title:	Data overview and descriptive statistical presentation
Description:	First we will organize the data and perform preliminary data analyses to overview what we have.
pISA Assay creation date:	2020-11-13
pISA Assay creator:	Andrej Blejec
Analyst:	Andrej Blejec
Phenodata:	Phenodata_20201109.txt
Featuredata:	Transcripts1819-02.txt
Featuredata metabolites:	Featuredata_metabolites_210127.txt
Transcript data 18:	/input/Transcripts 2018.txt
Transcript data 19:	/input/Transcripts 2019.txt
Metabolite data 18:	/input/Metabolites_2018_raw_-data.txt
Metabolite data 19:	/input/Metabolites_2019_raw_-data.txt
Water potential data:	/input/Stem water potential 2018 2019.txt
Interesting bins:	Bin_selection_2.txt
Transcript stats:	/output/50_Expression-water-stress-0.05-1.5-0.5-1-1/lm-fit-means-swpXvariety-statistics.txt

B SessionInfo

Windows 10 x64 (build 19042)

- R version 4.0.2 (2020-06-22), x86_64-w64-mingw32
- Locale: LC_COLLATE=Slovenian_Slovenia.1250, LC_CTYPE=Slovenian_Slovenia.1250, LC_MONETARY=Slovenian_Slovenia.1250, LC_NUMERIC=C, LC_TIME=Slovenian_Slovenia.1250
- Running under: Windows 10 x64 (build 19042)
- Matrix products: default
- Base packages: base, datasets, graphics, grDevices, grid, methods, parallel, splines, stats, utils
- Other packages: amisc 0.1.0, Biobase 2.50.0, BiocGenerics 0.36.0, dotCall64 1.0-0, fda 5.1.9, fds 1.8, fields 11.6, Formula 1.2-4, ggplot2 3.3.3, gplots 3.1.1, Hmisc 4.4-1, igraph 1.2.6, knitr 1.30, lattice 0.20-41, lubridate 1.7.9.2, MASS 7.3-51.6, Matrix 1.2-18, mixOmics 6.14.0, pcaPP 1.9-73, rainbow 3.6, RCurl 1.98-1.2, reshape2 1.4.4, spam 2.6-0, survival 3.2-7, xtable 1.8-4
- Loaded via a namespace (and not attached): backports 1.2.0, base64enc 0.1-3, bitops 1.0-6, caTools 1.18.1, checkmate 2.0.0, cluster 2.1.0, colorspace 1.4-1, compiler 4.0.2, corpcor 1.6.9, crayon 1.3.4, data.table 1.13.2, digest 0.6.27, dplyr 1.0.2, ellipse 0.4.2, ellipsis 0.3.1, evaluate 0.14, farver 2.0.3, foreign 0.8-80, generics 0.1.0, ggrepel 0.9.0, glue 1.4.2, gridExtra 2.3, gtable 0.3.0, gtools 3.8.2, hdrclde 3.4, highr 0.8, htmlTable 2.1.0, htmltools 0.5.0, htmlwidgets 1.5.3, jpeg 0.1-8.1, KernSmooth 2.23-17, ks 1.12.0, labeling 0.4.2, latticeExtra 0.6-29, lifecycle 0.2.0, magrittr 2.0.1, maps 3.3.0, matrixStats 0.57.0, mclust 5.4.7, munsell 0.5.0, mvtnorm 1.1-1, nnet 7.3-14, pillar 1.4.7, pkgconfig 2.0.3, plyr 1.8.6, png 0.1-7, purrr 0.3.4, R6 2.5.0, rARPACK 0.11-0, RColorBrewer 1.1-2, Rcpp 1.0.5, rlang 0.4.10, rpart 4.1-15, RSpectra 0.16-0, rstudioapi 0.13, scales 1.1.1, stringi 1.5.3, stringr 1.4.0, tibble 3.0.4, tidyr 1.1.2, tidyselect 1.1.0, tools 4.0.2, vctrs 0.3.6, withr 2.3.0, xfun 0.19

Analysis project path: [\[link\]](#)

Local

D:

DEJAVNOSTI

OMIKE

pISA-projects

_p_VinskaTrta

_I_EnViRoS

_S_01_Integ

_A_01_Desc-R

other

Network

O:

DEJAVNOSTI

OMIKE

pISA-projects

_p_VinskaTrta

_I_EnViRoS

_S_01_Integ

_A_01_Desc-R

other

Main file : [../doc/60_Trans-x-Meta.Rnw](#)

Main file : [../scripts/60_Trans-x-Meta.Rnw](#)

Project file: [\[link\]](#)

View as vignette

Source files can be viewed by pasting this code to R console:

```
projectName <-"other"
mainFile <-"60_Trans-x-Meta"
commandArgs()
library(tkWidgets)
openPDF(file.path(dirname(getwd()), "doc",
paste(mainFile, "PDF", sep=".")))
viewVignette("viewVignette", projectName,
file.path("../doc", paste(mainFile, "Rnw", sep=".")))
```